

# Yale Liver Center

## 2023 LIVER CENTER RETREAT



The Yale Liver Center held its annual retreat at the Water's Edge Resort and Spa, in Westbrook, Conn., on September 10, 2023.

Topics discussed at the event included important resources for Yale Liver Center members including the new academic unit [Biomedical Informatics and Data Science](#) and the [Yale Center for Clinical Investigation](#), cutting-edge liver research at Yale in metabolism and in immunobiology, and the projects undertaken by junior faculty and postdoctoral fellows through the center's pilot grant program and institutional liver training grant.

There were over 100 registrants and 30 poster presentations at the event, according to [Michael H. Nathanson, MD, PhD](#), director of the Yale Liver Center and Gladys Crofoot Professor of Medicine (Digestive Diseases) and professor of cell biology, who added that on-site daycare was provided to maximize the ability of individuals to attend the retreat.

A new feature of the event this year was a session that focused on the accomplishments of disease-aligned research teams, said [Mario Strazzabosco, MD, PhD](#), professor of medicine (digestive diseases), co-director of the Yale Liver Center, and organizer of the retreat. "These 10 teams exploit new discoveries and innovations to advance the care of patients with complex liver conditions and sustain destination programs able to provide highly differentiated world class care," he said. "What was evident was the richness of the research, the large number of patients treated, and the ability to cover many of the unmet needs of patients with liver disease—the tripartite mission of an academic medical center and school realized at his best."

Strazzabosco noted the participation of members from a variety of departments, both clinical and basic, which illustrates the interdisciplinary breadth of liver research at Yale.

The retreat helped raise awareness about recent scientific advances that resulted from Liver Center initiatives, said [Guadalupe Garcia-Tsao, MD, FRCP](#), professor of medicine (digestive diseases), who led a session on Liver Cen-

ter core offerings. "It also made participants more aware of what everybody else is doing within the Liver Center and—perhaps more importantly—outside of the Liver Center," she said.

The occasion is a forum for people with different interests and backgrounds to share ideas and information, added [Catherine Mezzacappa, MD, MPH](#), clinical fellow in the Section of Digestive Diseases, who presented at the event. "For example, I'm interested in the interactions between metabolism and serious liver diseases, and I learned a great deal from [Gerald Shulman, MD, PhD](#), who spoke about peripheral insulin resistance and how this can be modified," she said. "This kind of sharing across disciplines generates new questions and opportunities."

Nathanson hopes the retreat allowed participants to network with colleagues and improve their understanding of current resources at Yale to support their research. "The annual event is a wonderful opportunity to further enhance the already extensive degree of collaborations among our membership and beyond," he said.

A main highlight of the event was the keynote lecture given by [Nancy J. Brown, MD](#), Jean and David W. Wallace Dean of the Yale School of Medicine and C.N.H. Long Professor of Internal Medicine, who spoke about strategic planning for discovery and the ways Liver Center members can participate in many new initiatives at the school level.



[This article](#) was written by Serena Crawford

## 2022-2023 LIVER CENTER PILOT PROJECTS



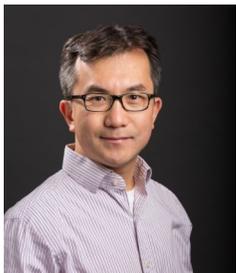
The morning session began with presentations from the recipients of our 2022-2023 Pilot Project Awards. The Pilot Feasibility Program seeks to introduce new investigators and ideas to the Center, to promote novel ideas that may advance the field of hepatology, and to enable investigators to obtain data for future grant submissions. This program has enabled the Center to continually revitalize its membership and to capitalize on new technologies and research opportunities of importance to the continued development of the Center's research base. For example, the program has enabled us to bring new investigators into the Center with expertise in diverse but cutting-edge areas. Pilot Feasibility projects also were pivotal for the development of novel technologies such as iPSC-derived cholangiocytes, as well as biliary organoids derived from liver tissue or bile.



**Vikas Gupta, MD, PhD**  
Assistant Professor of Medicine  
Digestive Diseases

Project Title: *Role of Cholangiocyte Derived Wnts During Cholestatic Injury*

This pilot proposal will utilize murine biliary injury models, genetics, and sequencing to discover how changes in the expression of cholangiocyte derived Wnts affects the cellular phenotype of adjacent mesenchyme during biliary injury in the large ducts using cell type specific gain and loss of function.



**Xiaoyong Yang, PhD**  
Professor  
Comparative Medicine

Project Title: *Characterizing novel paracrine factors in chronic liver disease*

The intercellular crosstalk between the parenchymal cell of the liver, the hepatocyte, and other cell types such as immune cells, stellate cells, and endothelial cells, is much undiscovered. Our long-term goal is to examine paracrine factors that mediate cell-cell communication in disease progression from NASH, fibrosis to HCC. The objective of this pilot proposal is to characterize the role of the hepatocyte-derived paracrine factor TFF2 in intercellular signaling during liver fibrogenesis.



**Won Jae Huh, PhD**  
Assistant Professor  
Pathology

Project Title: *Sex-Hormone-Induced Differential Expression of EGF Receptor (EGFR) in Hepatocytes and Its Implication for Hepatic Steatosis*

The main hypothesis of this study is that testosterone regulated hepatic EGFR expression is an underlying mechanism of sex disparity in NAFLD. We will utilize mouse models and hepatocyte organoids to dissect the mechanism how testosterone regulate EGFR and how EGFR signaling regulates lipid metabolism and accumulation in hepatocytes.



**Jason Bini, PhD**  
Associate Research Scientist  
Radiology & Biomedical Imaging

Project Title: *Positron Emission Tomography imaging of  $11\beta$ -hydroxysteroid dehydrogenase type 1 in NAFLD*

This pilot project will allow examination of  $11\beta$ -HSD1 whole-body distribution levels in human individuals with non-alcoholic steatohepatitis (NASH) using the [18F]FMOZAT PET radioligand. The results of this study will be important to understand the metabolic changes associated with steatotic liver disease.

## HEPATOLOGY T32 TRAINEES



The second session began with presentations from the current Hepatology T32 trainees.

This NIDDK-sponsored training program has been continuously supported for the past 44 years. The goal of this training program is to provide basic laboratory, translational, or clinical research training for physicians who have completed clinical training in gastroenterology in preparation for careers as independent investigators in academic hepatology and to provide research training for recent PhD graduates to prepare them for careers as independent investigators in basic liver-related research.



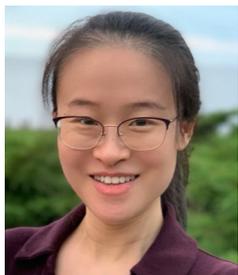
### **Jeremy Puthumana, MD**

Clinical Fellow | Digestive Diseases

Project Title: *Biomarkers for the Diagnosis and Prognosis of AKI in Patients with Cirrhosis*

Primary Mentor: Pramod Mistry

Faculty Committee: Pramod Mistry, Chirag Parikh, Dennis Moledina, and Michael Nathanson



### **Xing Feng, PhD**

Postdoctoral Fellow | Immunobiology

Project Title: *Role of Gpr56 and Gpr97 signaling in fetal liver hematopoiesis*

Primary Mentors: Joao Pereira & Silvia Vilarinho

Faculty Committee: Yasuko Iwakiri, Silvia Vilarinho, and Joao Pereira



### **Catherine Mezzacappa, MD, MPH**

Clinical Fellow | Digestive Diseases

Project Title: *Investigating gene-environment interactions in steatotic liver disease and HIV*

Mentors: Tamar Taddei & Amy Justice

Faculty Committee: Tamar Taddei, Amy Justice, Xiaomei Ma, Rachel Perry, Michael Nathanson, Chen Liu



### **Jonathan Pascale, PhD**

Postdoctoral Fellow | Pharmacology

Project Title: *Characterization of a novel knockin mouse to assess MKP5 druggability in tissue fibrosis*

Primary Mentor: Anton Bennett

Faculty Committee: Anton Bennett, Elias Lolis, Barbara Ehrlich, Naftali Kaminski



The retreat also featured three invited talks which highlighted some of the developments in the School of Medicine that may be relevant to our Liver Center members.

## CLINICAL RESEARCH



**David Coleman, MD**  
Interim Director, YCCI  
Emeritus Professor, Yale School of Medicine  
*How YCCI can help Clinical Research in Hepatology*

## BIOINFORMATICS & DATA SCIENCE FOR THE STUDY OF LIVER DISEASE



**Hua Xu, PhD**  
Robert T. McCluskey Professor of Biomedical Informatics and  
Data Science  
Vice Chair for Research and Development  
Assistant Dean for Biomedical Informatics  
*Biomedical Informatics and Data Science for  
Clinical And Translational Research at Yale*

## LIVER VASCULAR BIOLOGY



**Silvia Vilarinho, MD, PhD**  
Associate Professor of Medicine  
Digestive Diseases and of Pathology  
Associate Director, Yale MD-PhD Program  
Director, Internal Medicine Physician Scientist Training Program  
*Liver Endothelial Cell Biology*

# LIVER CENTER THEMES

Three invited speakers updated the membership on the latest developments in each of the three main themes of the Liver Center as shown below.

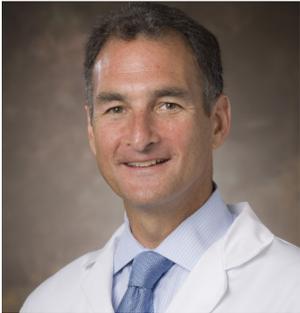


## Theme 1: Immunobiology & Inflammation



**Andres Hidalgo, PhD**  
 Professor of Medicine (Immunobiology)  
*Neutrophils: new paradigms*

## Theme 2: Hepatic Metabolism



**Gerald I. Shulman, MD, PhD**  
 George R. Cowgill Professor of Medicine (Endocrinology)  
 Professor of Cellular & Molecular Physiology  
 Co-Director, Yale Diabetes Research Center  
*Targeting hepatic mitochondrial fat oxidation to treat MASLD, MASH, hepatocellular cancer and cardiometabolic disease*

## Theme 3: Epithelial Biology & Pathobiology



**Romina Fiorotto, PhD**  
 Assistant Professor of Medicine (Digestive Diseases)  
 Associate Director, Cellular & Molecular Physiology Core  
*Novel and Multi-Omics approaches for cholestatic liver diseases and beyond*

## AVAILABLE CENTER CORE SERVICES

Another portion of the retreat was a roundtable chaired by Dr. Garcia-Tsao. Each of the Core Directors and Associate Directors discussed with the members the offerings of each core and potential new technologies needed by the members. Also during this session, a questionnaire was distributed among the members to better understand their evolving needs. Below is a list of current available core services.



### ADMINISTRATIVE

#### PILOT FEASIBILITY PROGRAM

Pilot grants given annually to promote studies of liver disease

#### ENRICHMENT PROGRAM

Monthly seminar series, annual Klatskin/Boyer Lectureship, annual Center retreat, extended visiting professorship, research in progress series

### MORPHOLOGY

#### CONFOCAL, SUPER-RESOLUTION, MULTIPHOTON IMAGING

Leica SP5 Confocal Microscope  
Stellaris 8 DIVE Multiphoton Microscope  
Zeiss LSM 880 Airyscan Confocal Microscope  
Leica SP8 Gated STED 3X Super Resolution  
Bruker Opterra II Swept Field Microscope  
Bruker Luxendo Light Sheet Microscope  
Bruker Vutara 352 Super Resolution Microscope

#### ELECTRON MICROSCOPY

Tecnai 12. biotwinFEI Tecnai TF20 FEG

#### OTHER MICROSCOPY TOOLS

Zeiss Axio Observer epifluorescence microscope  
Olympus BX51 multi-headed brightfield microscope  
Dissecting microscope  
Zeiss Discovery 8 SteReo

#### IMAGE ANALYSIS WORKSTATIONS

4 PC workstations dedicated to image analysis. Software from Zeiss (ZEN blue), Leica (LAS X), Bitplane (Imaris), SVI (Huygens Deconvolution) and Perkin Elmer (Volocity) are available to users.

### CELLULAR-MOLECULAR

#### ISOLATED CELL PREPARATIONS

Hepatocytes, cholangiocytes, endothelial cells, stellate cells, portal fibroblasts and hepatic lymphocytes, primarily from mice and rats. Human hepatocytes when available.

#### PROTEIN & GENE EXPRESSION

Quantitative real time PCR and infrared imaging detection. Altering gene expression in these cells using siRNA transfection and adenovirus infection technologies

#### IPSC/LIVER ORGANOID

On request, PBMCs are transferred to the Yale Stem Cell Center (YSCC) for reprogramming into iPSC. YSCC will generate at least 3 clones of iPSCs for each PBMC sample. iPSCs can be differentiated into liver cells (biliary cells or hepatocytes) and made available. Liver organoids available upon request.

#### CELL CULTURE FACILITIES

Available for short- and long- term cultures and cell lines.

### CLINICAL-TRANSLATIONAL

#### BIOSTATISTICAL SUPPORT

Two biostatisticians available for expertise in the design, conduct, and analysis of patient-oriented studies, as well as methodological development, education, and training

#### BIOINFORMATICS SUPPORT

Bioinformatics analysis support for Liver Center members through the Yale Center for Genome Analysis (YCGA) bioinformatics core.

#### RESEARCH COORDINATOR

Recruitment of patients and collection of blood samples and other biospecimens

#### PATIENT REGISTRY

Patient databases on diagnoses including: chronic hepatitis C, cirrhosis, chronic hepatitis B, PBC, autoimmune hepatitis, PSC, hepatocellular carcinoma, NAFLD, and cholangiocarcinoma

## POSTER SESSION



## NEW! CHILDCARE @ RETREAT



*This year, an exciting new addition to our event was that we were able to offer childcare for anyone who attended the retreat. We would like to thank the staff from the Medical Schools Bodel Childcare Center for spending the day with us and taking great care of the kids!*



# FEATURED PUBLICATIONS FROM LIVER CENTER MEMBERS FURTHER HIGHLIGHT THE BREATH OF SCIENTIFIC AND TECHNICAL INNOVATION



## Dean Yimlamai, MD, PhD

Assistant Professor of Pediatrics & Experimental Pathology  
Director of Pediatric Hepatology Research

### ***Hepatocyte CYR61 polarizes profibrotic macrophages to orchestrate NASH fibrosis***

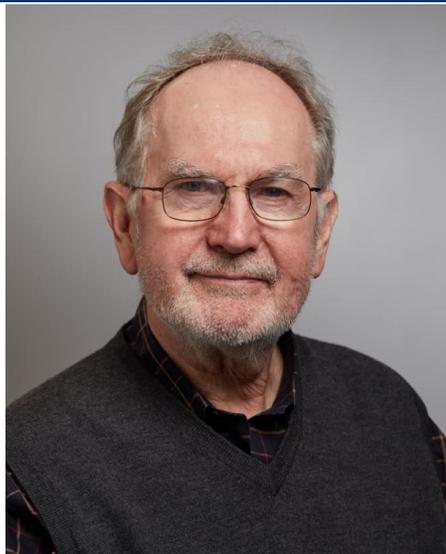
Mooring M, Yeung GA, Luukkonen P, Liu S, Akbar MW, Zhang GJ, Balogun O, Yu X, Mo R, Nejak-Bowen K, Poyurovsky MV, Booth CJ, Konnikova L, Shulman GI, Yimlamai D.

**Science Translational Medicine.** 2023;15(715):eade3157. PMID: 37756381

Nonalcoholic steatohepatitis is a severe form of liver disease that has no cure and whose etiology is incompletely understood. Mooring et al. show that the protein cysteine-rich angiogenic inducer 61 (CYR61) promotes liver-infiltrating macrophage polarization in NASH, leading to inflammation and fibrosis via NF- $\kappa$ B-linked and PDGF $\alpha$ /b-linked signaling pathways, respectively. The authors also developed a CYR61-blocking antibody that ameliorated inflammatory and profibrotic gene expression in macrophages in vitro and reduced fibrotic collagen deposition in a NASH model in vivo, demonstrating proof-of-principle targeting of this signaling axis.



## FEATURED PUBLICATION



**Richard A. Flavell, PhD**  
Sterling Professor of  
Immunobiology



**Wajahat Z. Mehal, MD**  
Professor of Medicine  
(Digestive Diseases)



**Eleanna Kaffe, PhD**  
Associate Research  
Scientist (Immunobiology)

### ***Humanized mouse liver reveals endothelial control of essential hepatic metabolic functions***

Kaffe E, Roulis M, Zhao J, Qu R, Sefik E, Mirza H, Zhou J, Zheng Y, Charkoftaki G, Vasiliou V, Vatter DF, Mehal WZ; AlcHepNet; Yuval Kluger, Flavell RA.

**Cell.** 2023; 186(18):3793-3809.e26. PMID: 37562401

A comprehensive human liver tissue was established in a mouse host that consists of all human-relevant parenchymal and non-parenchymal cell types and mimics the cellular composition, histological architecture, and functional properties of a human liver. This highly human-relevant murine model allows investigation of human-specific metabolic features and liver cell type interactions. Using this model, it was found that essential hepatocyte functions like cholesterol uptake and bile acid conjugation are not cell autonomous but are highly dependent on signals from the adjacent endothelial cells. This model can develop human fibrosis upon damaging insult in liver epithelial cells and NAFLD upon western diet feeding. This model can be used to study mechanisms driving human liver fibrosis and human NAFLD and their consequences on human specific liver functions.

## FEATURED PUBLICATION

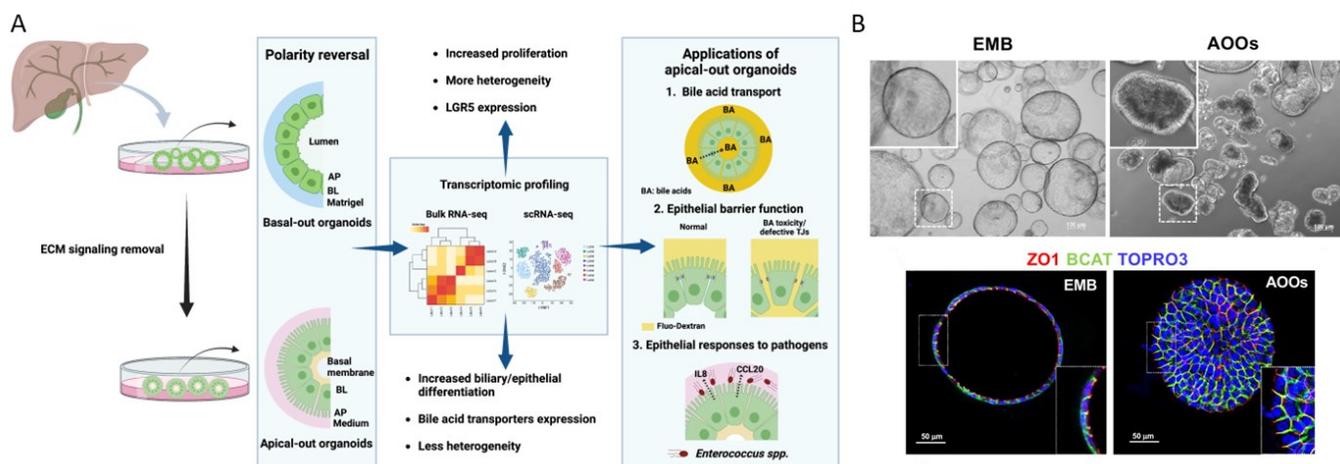


**Romina Fiorotto, PhD**  
Assistant Professor of Medicine  
Digestive Diseases

### ***Cell-matrix interactions control biliary organoid polarity, architecture, and differentiation.***

Fiorotto R, Mariotti V, Taleb SA, Zehra SA, Nguyen M, Amenduni M, Strazzabosco M.  
**Hepatology Communications.** 2023; 7(4):e0094. PMID: 36972396

Human biliary organoids are an attractive technology to study the pathobiology of biliary diseases, but current models are limited by the inaccessibility of their apical pole and the presence of extracellular matrix (ECM). This study published in *Hepatology Communications*, describes a new approach that overcomes these limitations by generating liver derived 3D human biliary organoids, in which the cell polarity is inverted, and the apical membrane is easily accessible. This approach is based on the modulation of  $\beta_1$ -integrin signaling, that controls the apical-basal orientation. Apical-out organoids generated in this study exhibit less cellular heterogeneity and improved functional maturation of cells and reproduce essential functions of the biliary epithelium such as bile acid transport and epithelial barrier function. Moreover, the exposure of apical-out biliary organoids to different pathogenic bacterial strains elicit specific innate immune responses making them ideal to study epithelial pathogen interactions. This novel approach represents a powerful new model to study liver biology and especially cholangiocyte biology, cell-cell and cell-ECM interactions, epithelial-pathogen interactions, and the pathobiology of various cholangiopathies.



(A)The cartoon illustrates the experimental design and the main findings of the study. (B)Brightfield and confocal images showing respectively the change of morphology in apical-out organoids (AOOs) and the distribution of markers of cell polarization (ZO-1, tight-junctions marker; b-catenin, basolateral membrane)