

# Relating Protein Abundance & mRNA Expression

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NIDA site visit at Yale  
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# Why relate amounts of protein & mRNA

Gene expression -  
major place for **regulation**  
(easy to measure)

VS.

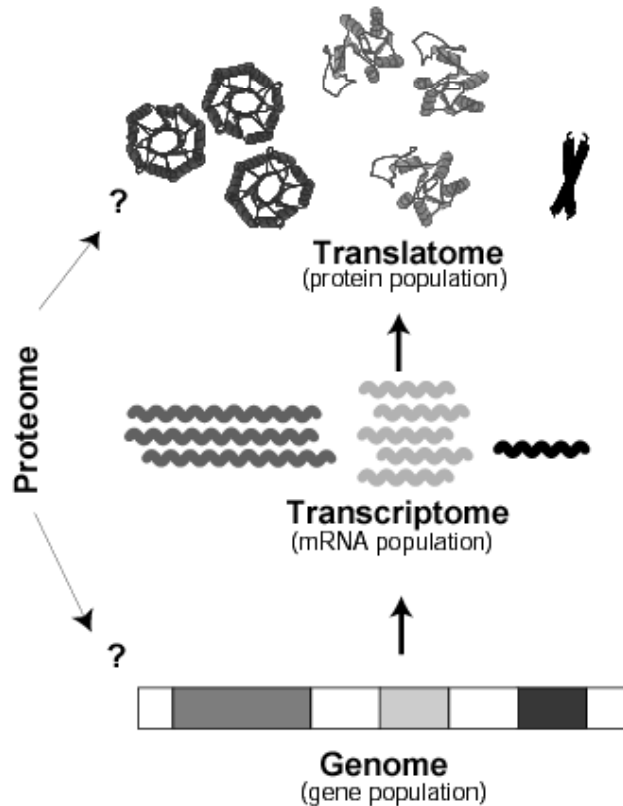
Concentration of protein -  
major determinant of **activity**

**Expectations** from simple kinetic models:

$$\frac{dP_i}{dt} = k_{s,i}[\text{mRNA}_i] - k_{d,i} P_i$$

$$\text{At steady state: } P_i = \frac{k_{s,i}[\text{mRNA}_i]}{k_{d,i}}$$

where  $k_{s,i}$  and  $k_{d,i}$  are the protein synthesis and degradation rate constants

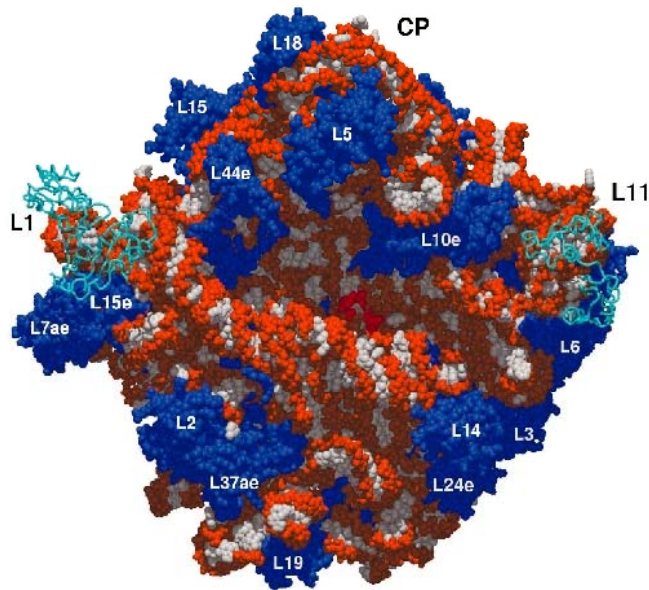


**Outliers** from trend interesting

# Relationship of Protein Abundance to Complexes and Pathways

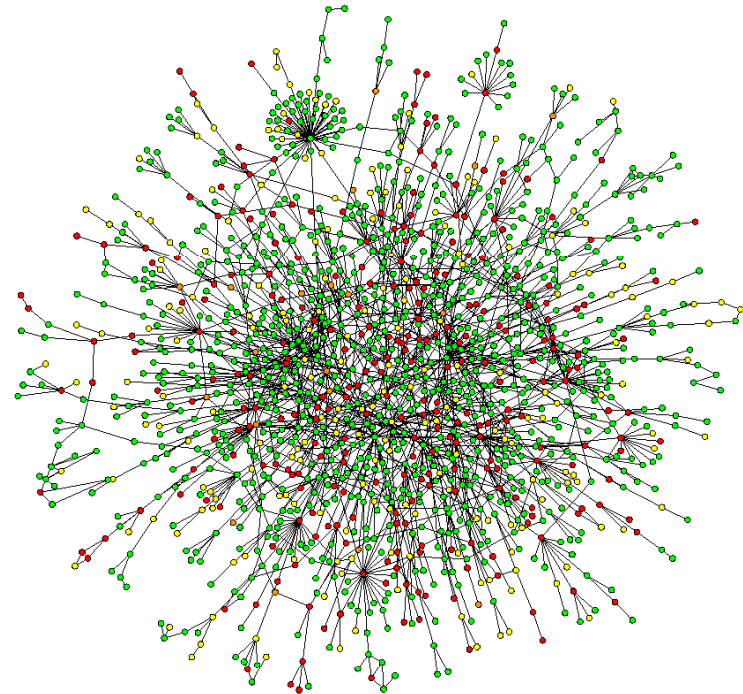
In protein complexes, one expects stoichiometric abundance of component proteins and that mRNA expression levels should be correlated with protein abundance

...Among pathways, this is expected to a lesser degree between interacting proteins



**Protein complexes**

[Graphic: <http://proton.chem.yale.edu>]



**Protein interaction networks**

[Graphic: Jeong et al, Nature, 41:411]

# Sources of experimental data

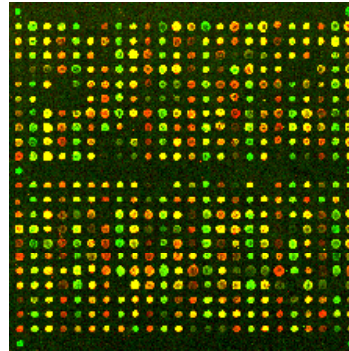
## mRNA expression levels

Microarrays

Affymetrix

PCR

SAGE



## Protein abundance

2D Gel Electrophoresis

Multiple staining options

Small dynamic range

DIGE

Cy3 vs. Cy5 labeling

Large dynamic range

ICAT, iTRAQ

MS-based

Relative abundance (ratio of isotopically labeled species)

Large dynamic range

MudPIT

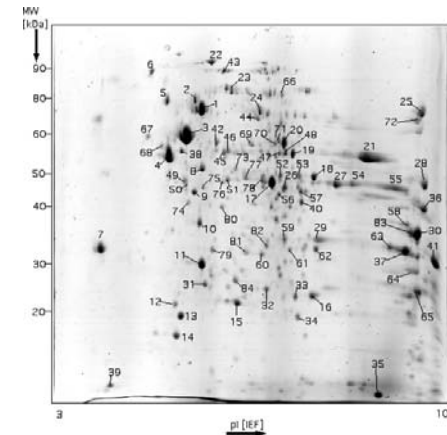
LC-MS/MS

SILAC

Stable isotope labeling with amino acids in cell culture - for MS analysis

TAP-Tag

Weissman and O'Shea (Oct. 2003)



# PARE: [proteomics.gersteinlab.org](http://proteomics.gersteinlab.org)

proteomics.gersteinlab.org  
**PARE: Protein Abundance and mRNA Expression Correlation Tool**

**Choose datasets** [help](#)

**1. Select organism:**    
[required if selecting datasets from menus in (2) and (3) below]

**2. mRNA expression**  
Select [complete citations](#)  
   
or upload [example file](#)

**3. Protein abundance**  
Select [complete citations](#)  
   
or upload [example file](#)

Please see [external mRNA expression and protein abundance databases](#) to retrieve additional datasets for analysis.

**Analyze** [help](#) [download PARE](#)

**Correlate everything**  
Perform correlation for selected categories (subsets for selection appear on next page)

- pick a MIPS complex of proteins for the analysis
- pick a GO biological process subset
- pick a GO molecular function subset
- pick a GO cellular component subset
- upload your own subset [example file](#)

**Other tools**  
[Sequence variation \(SNP\) substitution generator](#)

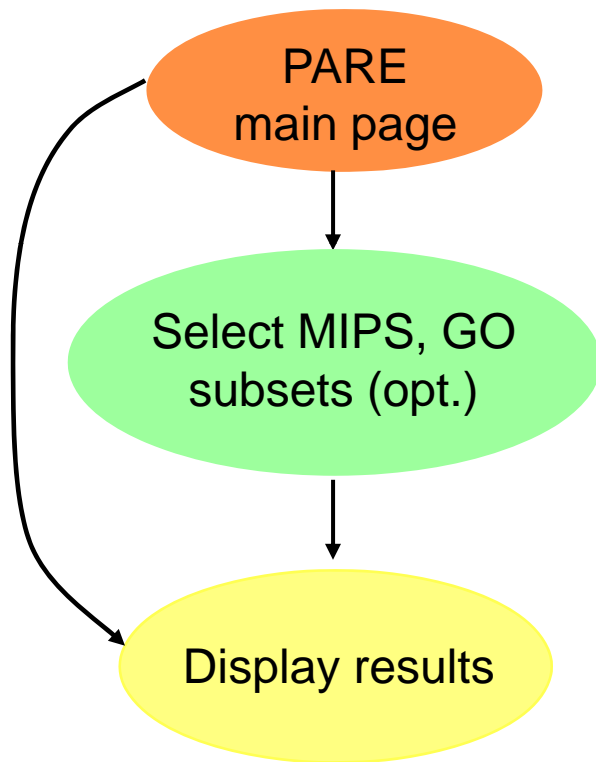
Upload or use pre-loaded mRNA, protein datasets

Open-source code  
Downloadable

Analyze all or  
analyze MIPS or GO subset

[Yu et al., BMC Bioinfo. '07]

# PARE: a web-based tool for correlating mRNA expression and protein abundance



(1) Select mRNA, protein datasets:

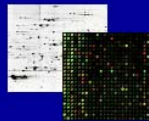
- use pre-loaded datasets
- upload datasets

(2) Choose categorization method:

- correlate all
- MIPS complexes
- GO biological processes
- GO molecular function
- GO cellular component

(3) Display

- Linear or log-log correlation for selected subset(s)
- Tabulate data, correlation values for selected subset(s)
- Label (on plot) and tabulate outlying datapoints



# PARE output

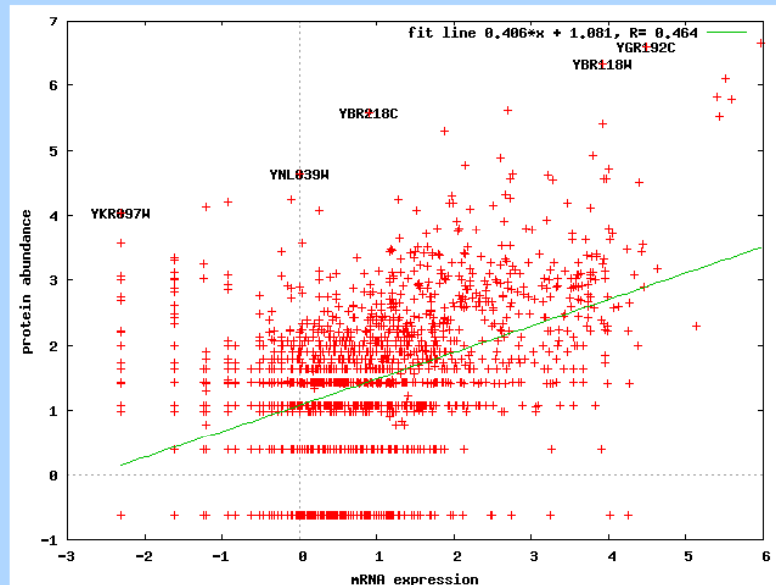
The following analysis is a log-log correlation. Switch to a linear correlation?

**Combined mRNA-protein file** (sorted by perpendicular distance to fit line)

ORF_id	mRNA	Protein	Dist_to_fit
YBR218C	0.899	5.580	3.830
YKR097W	-2.303	4.041	3.608
YGR192C	4.489	6.580	3.406
YBR118W	3.928	6.325	3.381
YNL039W	0.000	4.636	3.294
YOR347C	-1.204	4.130	3.277
YIL136W	-0.916	4.210	3.244
YJR104C	2.688	5.617	3.192
YPL231W	1.882	5.286	3.188
YKR057W	4.255	-0.629	3.185

### mRNA-protein overall correlation figure

Please note that the plot is loaded as an image file; you may need to refresh your browser to obtain the most recent plot.



customize the number of outliers shown in the plot (the top 5 shown by default)

absolute number: 
  
 percentage:  % out of 2041

Mutual information [help](#) = 10.66

Calculated using 204 bins for the mRNA and protein data

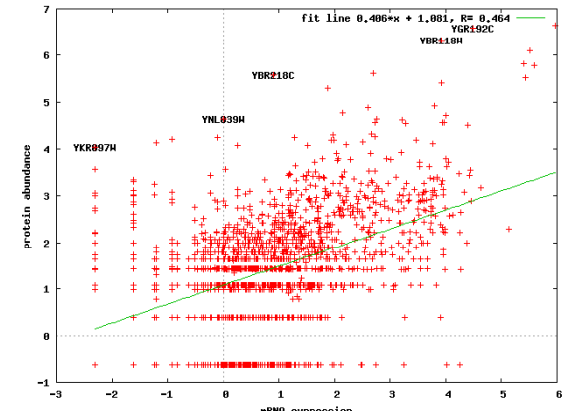
Correlated data

Log-log plot of correlation  
 -linear fit  
 -outliers labeled

Calculation of  
 mutual information

# Correlation of subsets (GO, MIPS)

Yeast ref. datasets:  
 “Correlate all”  
 VS.  
 GO cellular component subsets  
 for particular cellular locations



PARE: Protein-Abundance-vs-MRNA - Mozilla Firefox

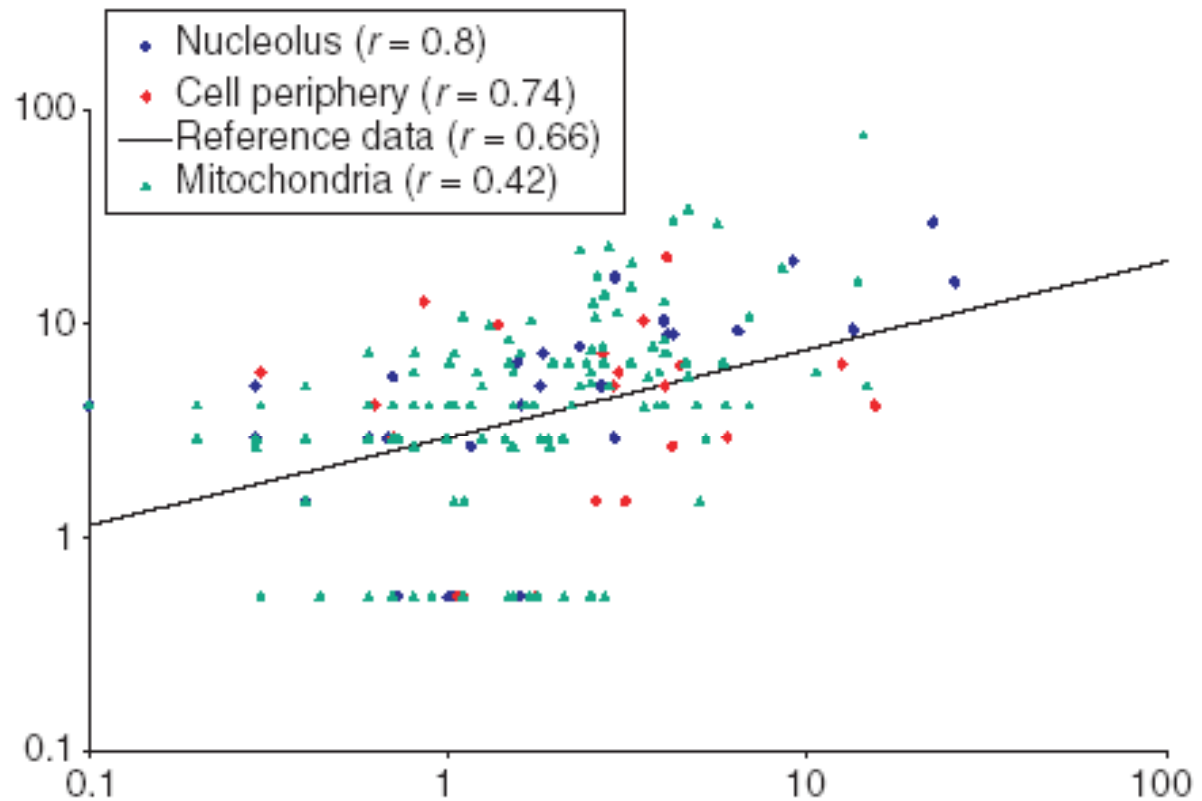
http://proteomics.gersteinlab.org

proteomics.gersteinlab.org  
**PARE: Protein Abundance**

The following 252 complexes are available. Please choose the ones on which you want to correlate.

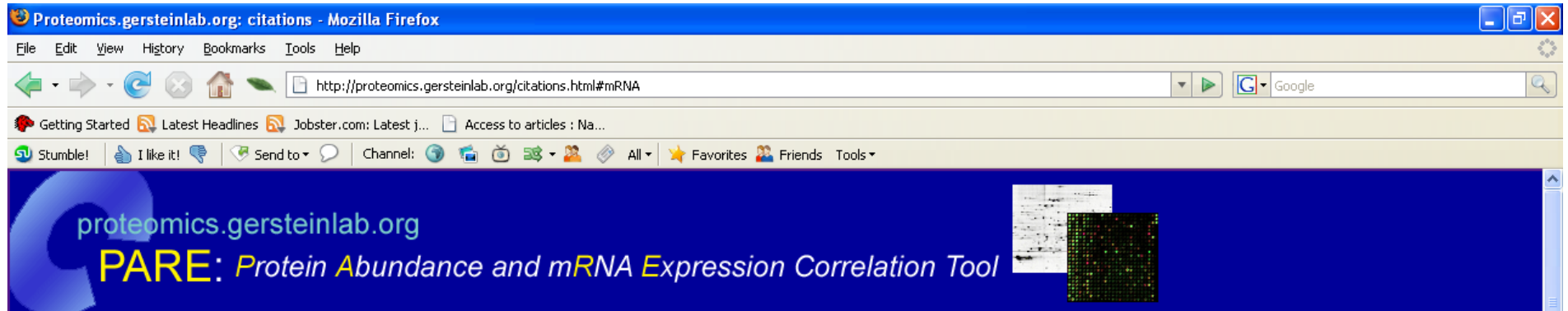
select or deselect all

- cytoplasmic ribosomal large subunit (80 entries)
- cytoplasmic ribosomal small subunit (57 entries)
- mitochondrial ribosomal large subunit (43 entries)
- mRNA splicing (37 entries)
- mitochondrial ribosomal small subunit (29 entries)
- Actin-associated proteins (25 entries)
- rRNA splicing (22 entries)
- Kornberg's mediator (SRB) complex (20 entries)
- F0/F1 ATP synthase (complex V) (18 entries)
- 19/22S regulator (18 entries)
- H<sup>+</sup>-transporting ATPase, vacuolar (15 entries)
- 20S proteasome (15 entries)
- Tubulin-associated proteins (14 entries)
- RNA polymerase I (14 entries)
- Nuclear pore complex (NPC) (13 entries)
- other respiration chain complexes (13 entries)
- Mitochondrial splicing complexes (12 entries)
- CCR4 complex (12 entries)
- Anaphase promoting complex (APC) (11 entries)
- SPB components (11 entries)
- tRNA splicing (11 entries)
- COPII (11 entries)
- Cytochrome c oxidase (complex IV) (11 entries)
- Pre-replication complex (pre-RC) (10 entries)
- Cytochrome bc1 complex (Ubiquinol-cytochrome c reductase complex) (10 entries)
- SAGA complex (10 entries)
- RSC complex (Remodel the structure of chromatin) (10 entries)
- Cdc28p complexes (10 entries)
- TRAPP (Transport Protein Particle) complex (10 entries)
- Oligosaccharyltransferase (9 entries)
- TOM - transport across the outer membrane (9 entries)
- Dam1 protein complex (9 entries)
- RNase MRP (9 entries)





# PARE: pre-loaded datasets



## mRNA expression datasets

Data type	Organism	Number of ORFs	Method	Rel./Abs.	Citation
mRNA	yeast	6250	Affymetrix GeneChip	A	<a href="#">Greenbaum, D., Colangelo, C., Williams, K. and Gerstein, M. (2003) Comparing protein abundance and mRNA expression levels on a genomic scale. Genome Biology, 4, 117</a>
mRNA	yeast	5442	Affymetrix GeneChip	A	<a href="#">Holstege, F.C. and Jennings, E.G., et al. (1998) Dissecting the regulatory circuitry of a eukaryotic genome. Cell, 95, 717-728.</a>
mRNA	yeast	6090	Affymetrix GeneChip	A	<a href="#">Jelinsky, S.A. and Samson, L.D. (1999) Global response of Saccharomyces cerevisiae to an alkylating agent. PNAS USA, 96, 1486-1491.</a>
mRNA	yeast	6214	Affymetrix GeneChip	A	<a href="#">Roth, F.P. and Hughes, J.D., et al. (1998) Finding DNA regulatory motifs within unaligned noncoding sequences clustered by whole-genome mRNA quantitation. Nature Biotechnology, 16, 939-945.</a>
mRNA	yeast	2961	SAGE	A	<a href="#">Velculescu, V.E. and Zhang, L., et al. (1997) Characterization of the yeast transcriptome. Cell, 88, 243-251.</a>
mRNA	rat	8799	Affymetrix RGU34A arrays	A	<a href="#">Walker, J.R., Su, A.I., Self, D.W., Hogenesch, J.B., Lapp, H., Maier, R., Hoyer, D. and Bilbe, G. (2004) Applications of a rat multiple tissue gene expression data set. Genome Research, 14, 742-749.</a>
mRNA	yeast	5935	DNA microarray	R	<a href="#">Ideker T, Thorsson V, Ranish JA, Christmas R, Buhler J, Eng JK, Bumgarner R, Goodlett DR, Aebersold R, Hood L. (2001) Integrated genomic and proteomic analyses of a systematically perturbed metabolic network. Science, 292, 929-934.</a>
					<a href="#">Griffin TJ, Gygi SP, Ideker T, Rist B, Eng J, Hood L, Aebersold R. (2002) Complementary profiling of gene</a>

[Yu et al., BMC Bioinfo. '07]

# Connecting PARE with datasets from NIDA investigators

**Rat** (Nairn lab) - samples from 3 brain regions:  
cortex, striatum, hippocampus

*Protein abundance* - MudPIT datasets

(protein profiling core)

samples for each brain region pooled from 4 rats

*mRNA expression* - Affymetrix microarray

(microarray core)

microarrays for brain regions for each of

4 rat individuals + replicate measurements (13 datasets)

**Mouse** (Nairn lab) - 3 brain regions

only iTRAQ data available; no matched mRNA datasets

# Rat brain: mRNA expression

mRNA samples

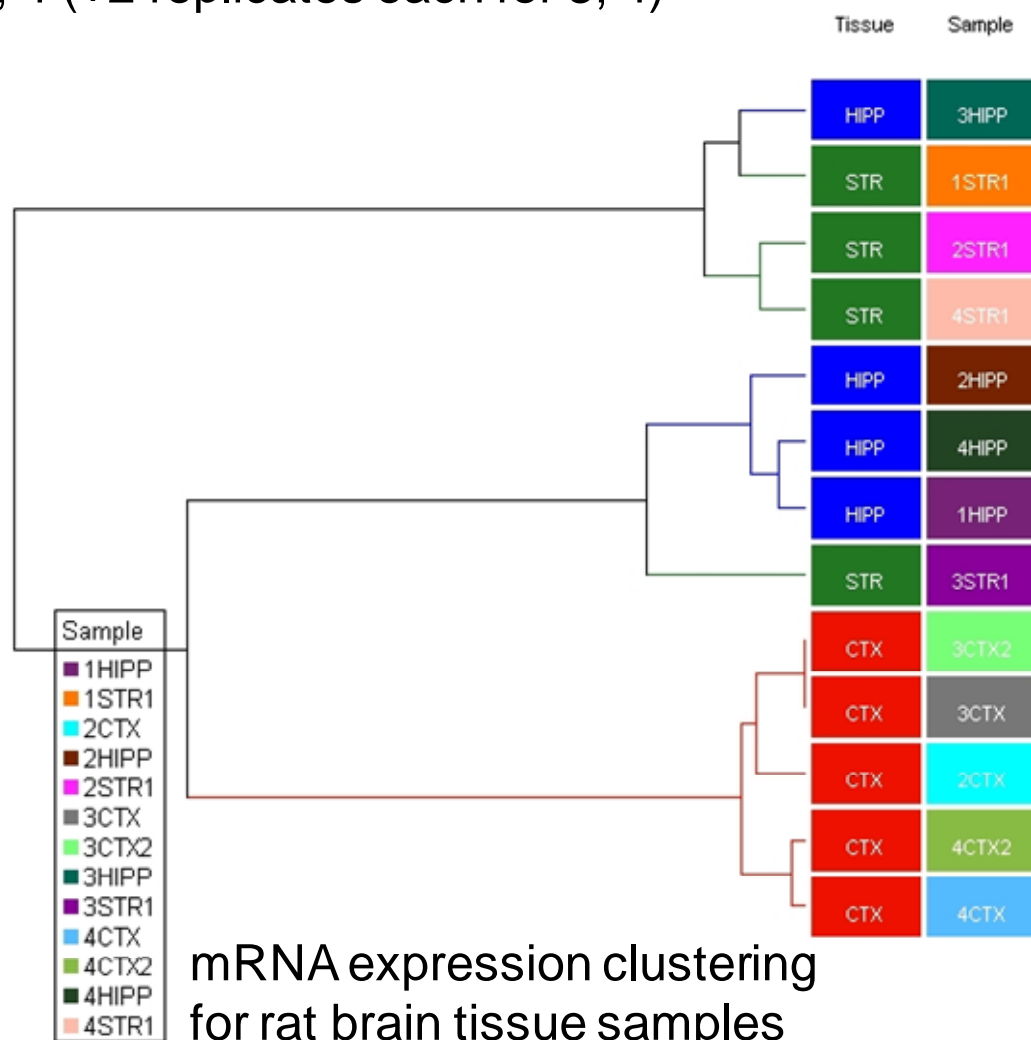
4 individual rats

CTX: indiv. 2, 3, 4 (+2 replicates each for 3, 4)

HIPP: indiv. 1-4

STR: indiv. 1-4

28,894 probes



# Rat brain: protein abundance

Rat brain

3 tissues: Cortex (CTX), striatum (STR), hippocampus (HIPP)

Number of matched mRNA/proteins pairs between Affymetrix and MudPIT data:

CTX: 830 proteins

STR: 926 proteins

HIPP: 701 proteins

Number of proteins observed in 2 or more tissues (for MudPIT data):

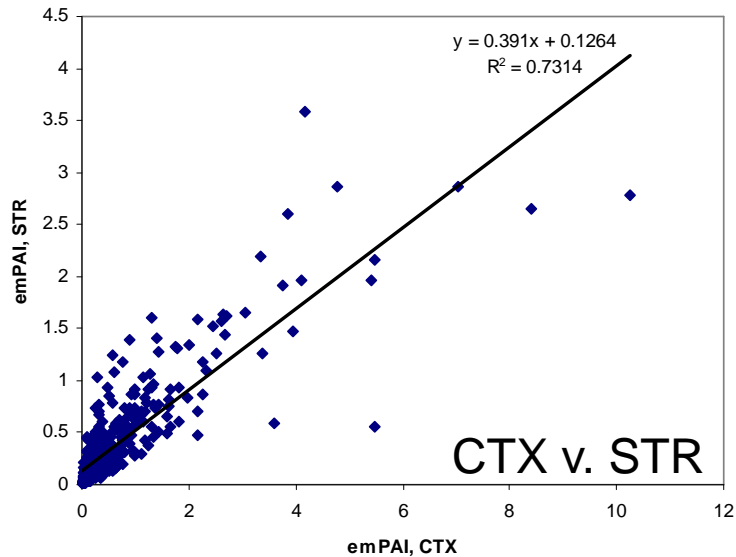
CTX and STR: 517 proteins

CTX and HIPP: 538 proteins

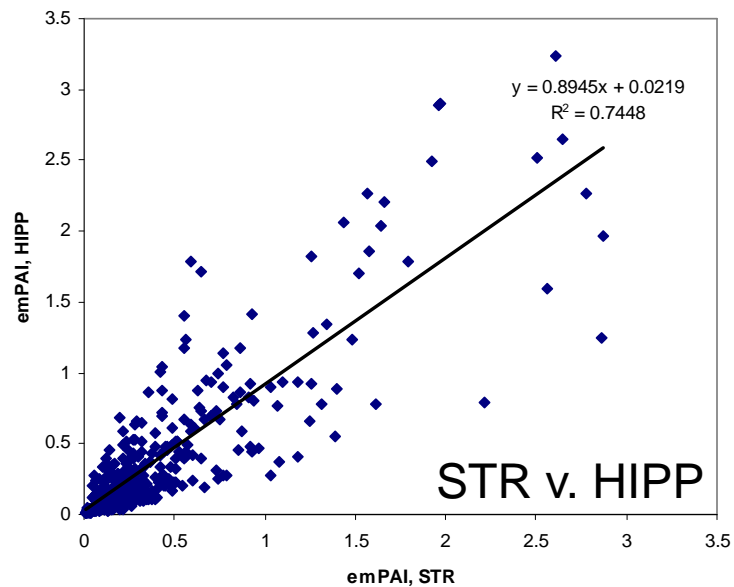
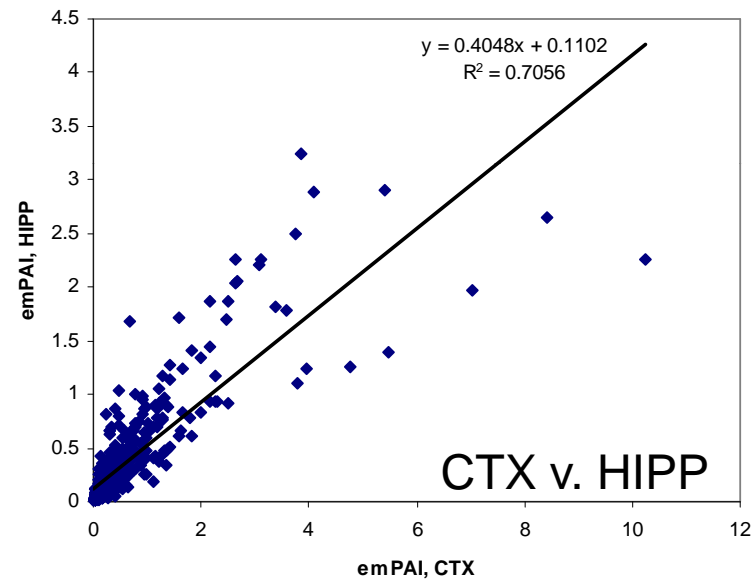
STR and HIPP: 485 proteins

CTX, STR, and HIPP: 421 proteins

# Rat brain: protein abundance

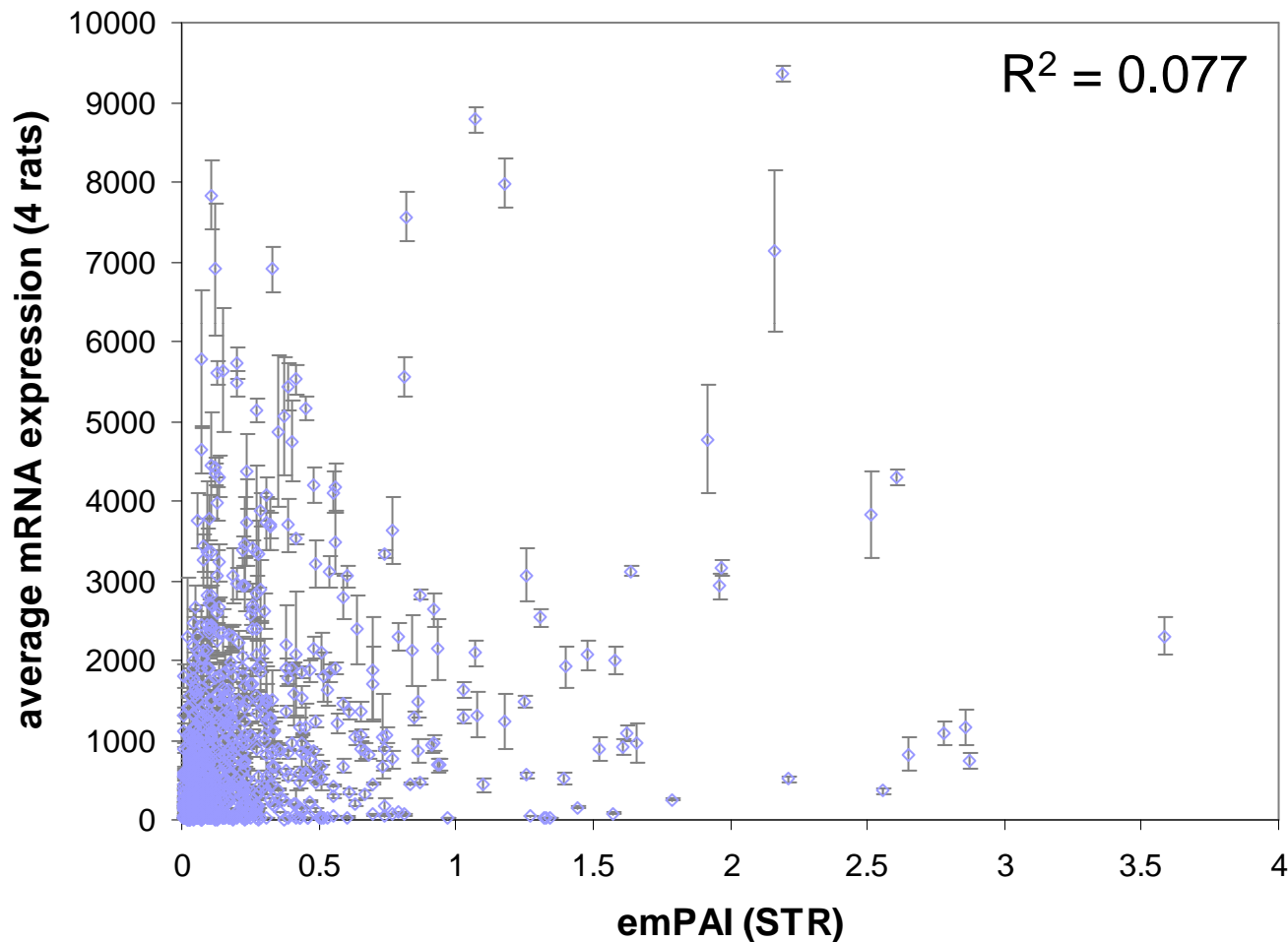


emPAI correlations for proteins observed in multiple tissues



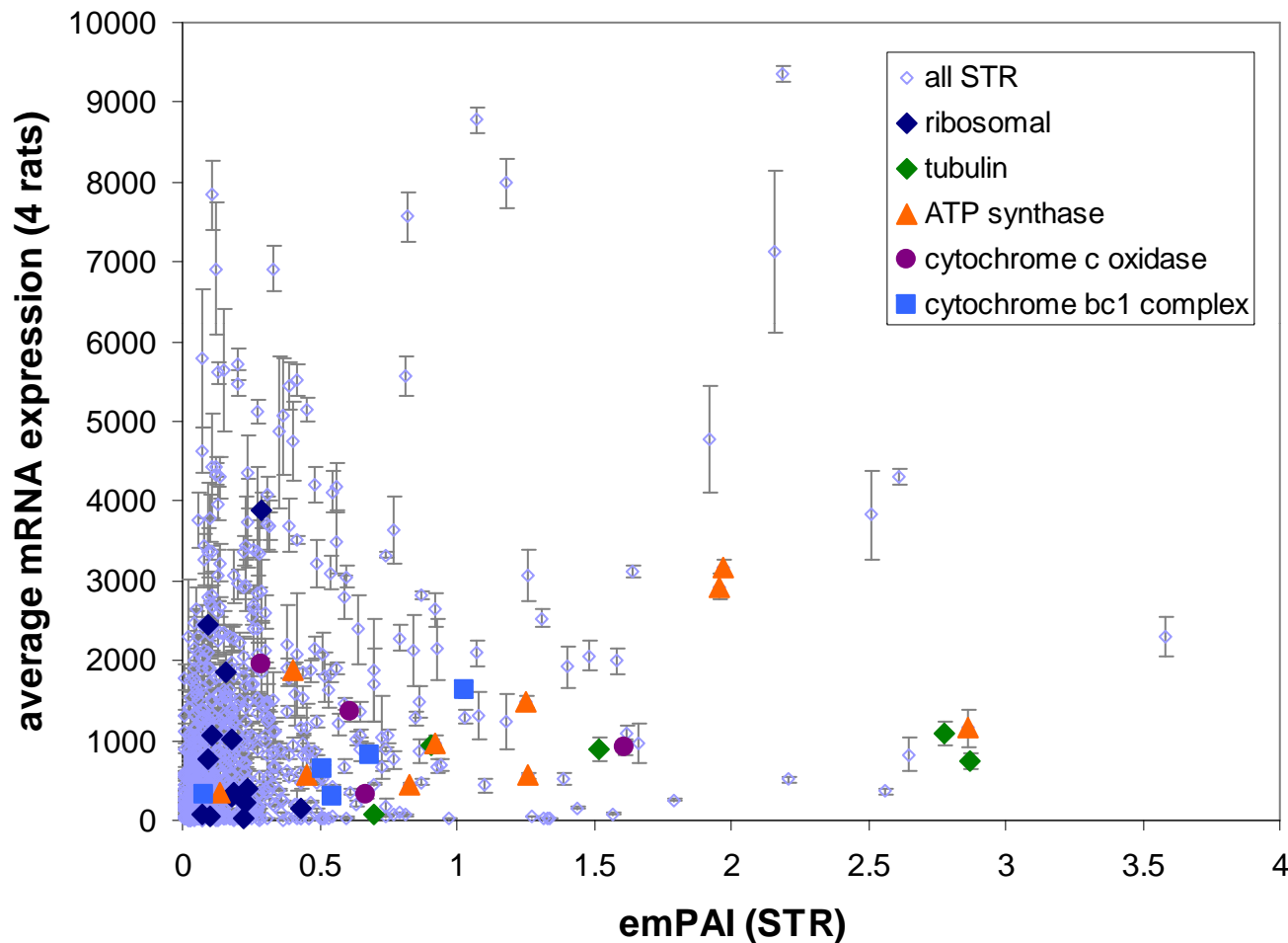
emPAI = exponentially modified protein abundance index, metric proportional to protein abundance (Ishihama et al. Mol. Cell. Proteomics (2005) 4, 1265-1272.)

# Rat brain: correlation of mRNA expression and protein abundance



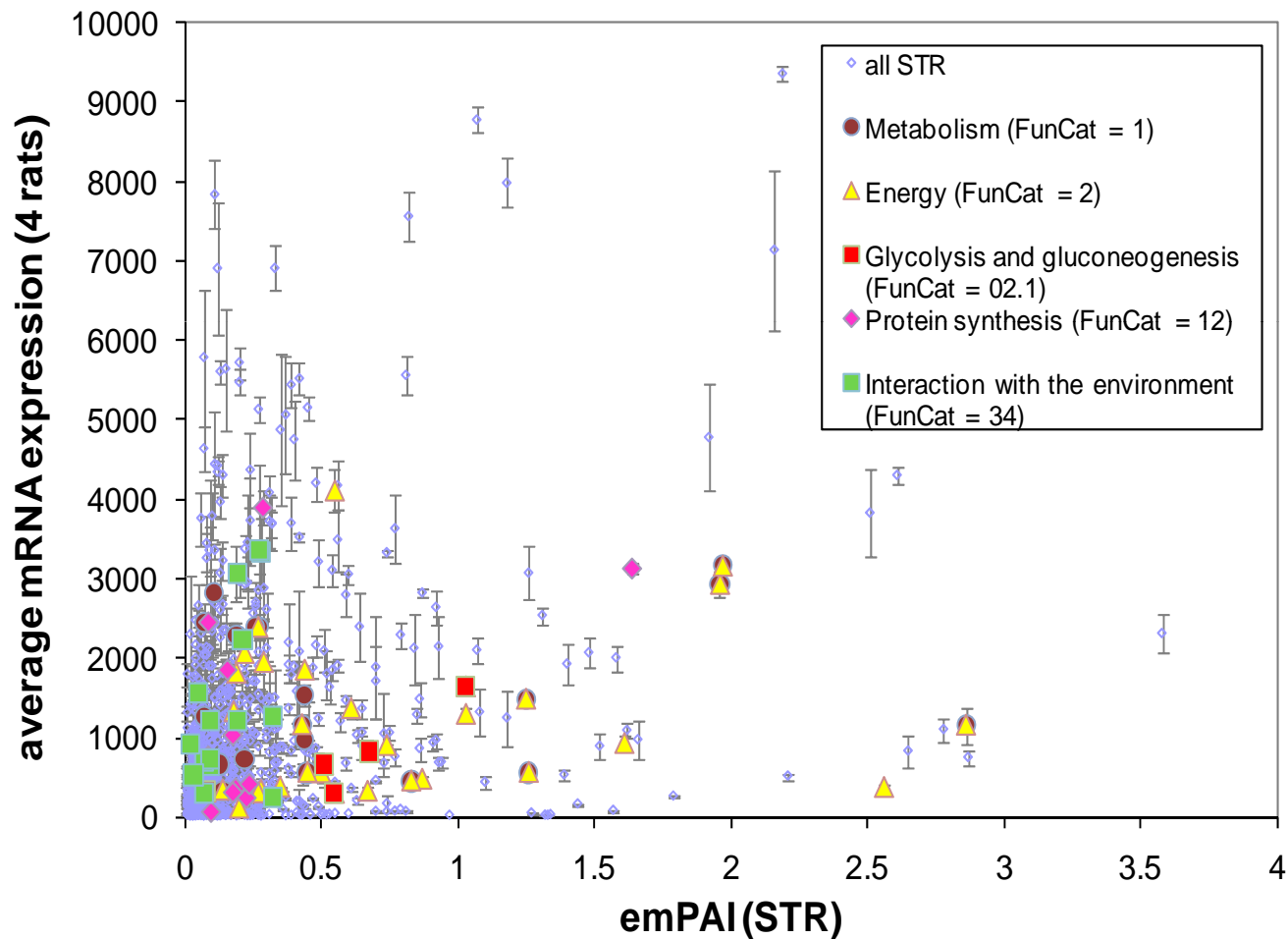
Rat brain: emPAI(STR) v. *average* mRNA expression (4 rats: 1STR to 4STR)  
926 proteins matched with corresponding mRNA

# Rat brain: mRNA expression and protein abundance for protein complexes



- Ribosomal proteins observed for STR appear to have low emPAI and a wide range of mRNA expression.
- Proteins that comprise tubulin have a wide range of emPAI (including 2 of the 4 highest values), but a narrow range of mRNA expression.

# Rat brain: mRNA expression and protein abundance for functional categories





# Acknowledgements

MudPIT datasets:

Angus Nairn, Erika Andrade

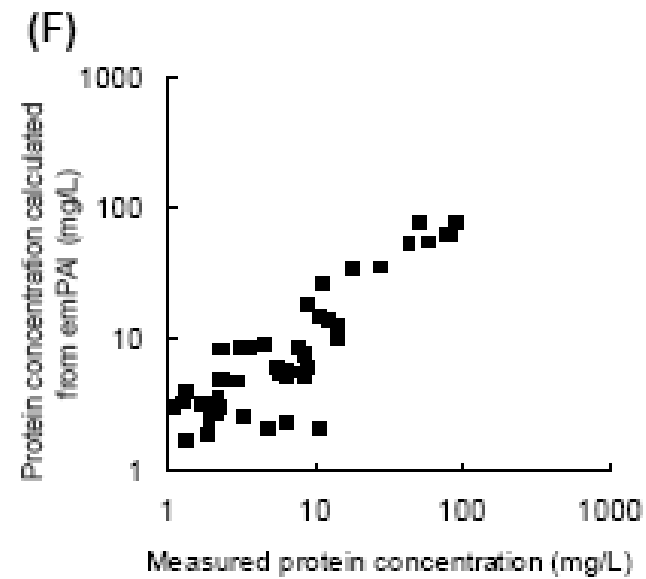
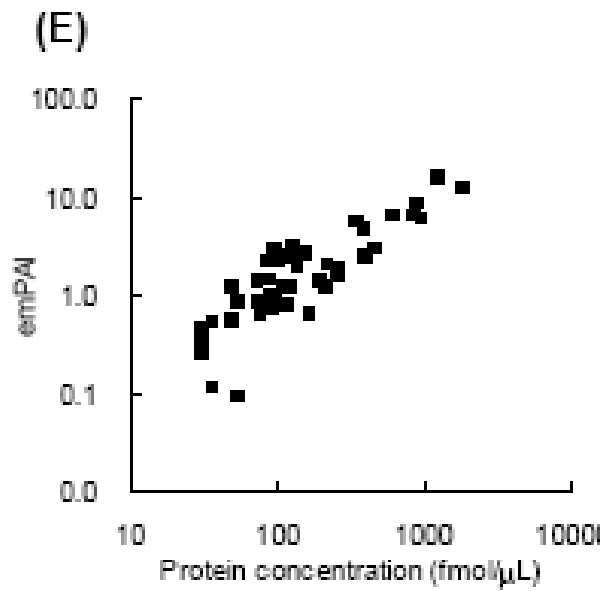
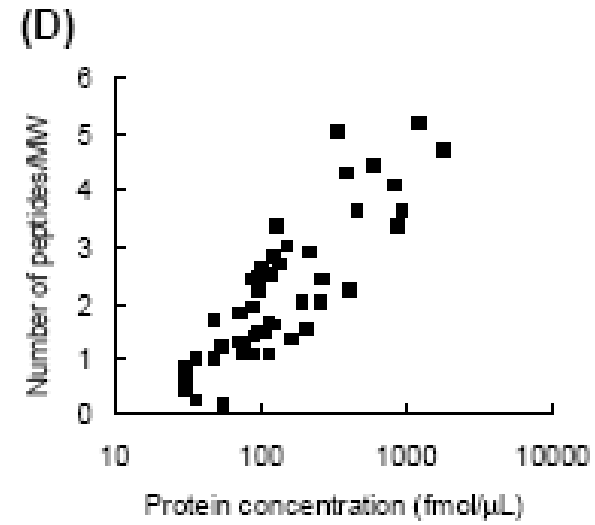
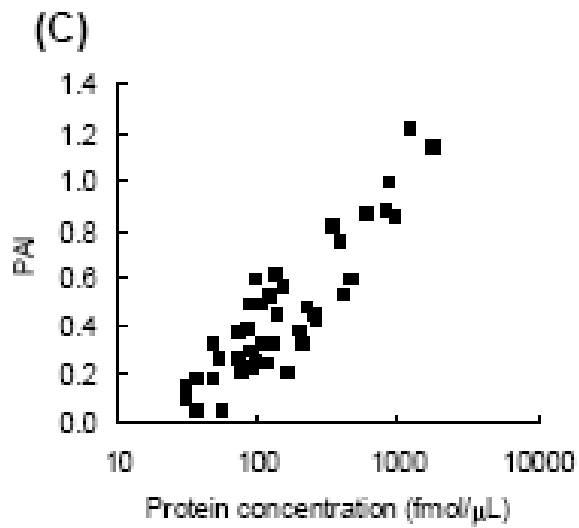
Kathy Stone, Chris Colangelo, Mark Shifman (YPED)

mRNA datasets:

Shrikant Mane, Aiping (Amy) Lin

PARE:

Anne Burba, Eric Yu



Ishihama et al. *Mol. Cell. Proteomics* (2005) 4, 1265-1272.