



Nuclear-Import Receptors Counter Deleterious Phase Transitions in Neurodegenerative Disease

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<https://doi.org/10.1016/j.jmb.2021.167220>

Edited by Monika Fuxreiter

Abstract

Nuclear-import receptors (NIRs) engage nuclear-localization signals (NLSs) of polypeptides in the cytoplasm and transport these cargo across the size-selective barrier of the nuclear-pore complex into the nucleoplasm. Beyond this canonical role in nuclear transport, NIRs operate in the cytoplasm to chaperone and disaggregate NLS-bearing clients. Indeed, NIRs can inhibit and reverse functional and deleterious phase transitions of their cargo, including several prominent neurodegenerative disease-linked RNA-binding proteins (RBPs) with prion-like domains (PrLDs), such as TDP-43, FUS, EWSR1, TAF15, hnRNPA1, and hnRNPA2. Importantly, elevated NIR expression can mitigate degenerative phenotypes connected to aberrant cytoplasmic aggregation of RBPs with PrLDs. Here, we review recent discoveries that NIRs can also antagonize aberrant interactions and toxicity of arginine-rich, dipeptide-repeat proteins that are associated with amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) caused by G₄C₂ hexanucleotide repeat expansions in the first intron of *C9ORF72*. We also highlight recent findings that multiple NIR family members can prevent and reverse liquid–liquid phase separation of specific clients bearing RGG motifs in an NLS-independent manner. Finally, we discuss strategies to enhance NIR activity or expression, which could have therapeutic utility for several neurodegenerative disorders, including ALS, FTD, multisystem proteinopathy, limbic-predominant age-related TDP-43 encephalopathy, tauopathies, and related diseases.

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Introduction

A distinctive feature of several fatal and presently incurable neurodegenerative diseases, including amyotrophic lateral sclerosis (ALS), frontotemporal dementia (FTD), multisystem proteinopathy (MSP), and limbic-predominant age-related TDP-43 encephalopathy (LATE), is the depletion of specific RNA-binding proteins (RBPs) with prion-like domains (PrLDs) from the nucleus and

their accumulation in cytoplasmic aggregates in degenerating neurons.^{1–8} For example, TDP-43 exhibits this pathological phenotype in ~97% of ALS cases and ~50% of FTD cases, whereas another RBP with a PrLD, FUS, displays nuclear depletion and cytoplasmic aggregation in ~1% of ALS cases and ~9% of FTD cases.^{1,8} It is suggested that the loss of nuclear function of these RBPs coupled to a gain of toxic function due to cytoplasmic accumulation and aggregation may syner-

gize to elicit neurodegeneration.^{1,6,8–10} We have suggested that agents that reverse the cytoplasmic mislocalization and aggregation of these RBPs and restore their nuclear localization and function could be powerful therapeutics.^{6,11–18} Remarkably, nuclear-import receptors (NIRs) have emerged as agents capable of eliciting such a therapeutic effect.^{16,19}

NIRs are members of the karyopherin family of proteins that bind tightly to nuclear-localization signals (NLSs) of polypeptide cargo in the cytoplasm.^{20–22} NIRs are subdivided into a small family of karyopherin- α (Kap α) proteins, which engage classical NLSs and subsequently bind to a member of the larger karyopherin- β (Kap β) family that enables nuclear import.²² However, Kap β s can directly recognize non-classical NLSs (e.g. proline tyrosine [PY]-NLSs) independent of Kap α s.²² Kap β s are flexible, superhelical proteins that typically comprise ~ 20 consecutive HEAT (Huntingtin, elongation factor 3 (EF3), protein phosphatase 2A (PP2A), and the yeast kinase TOR1) repeats, a type of protein tandem repeat structural motif composed of two alpha helices linked by a short loop (Figure 1(A)).²² Once bound to the NLS, NIRs can transport their cargo across the nuclear-pore complex (NPC) and into the nucleoplasm.²²

The NPC is an intricate structure²³ which operates as a size-selective barrier to prevent macromolecules with a molecular weight greater than

~ 30 kDa (or a Stokes radius greater than ~ 3 nm) from passively diffusing in and out of the nucleus.²⁴ This barrier is established by FG-rich nucleoporins, which form a phase-separated state inside the NPC channel.^{25–28} NIRs can penetrate rapidly through this phase and transport cargo into the nucleus.²⁹ Once inside the nucleoplasm, the small GTPase Ran in its GTP-bound state binds to the incoming NIR, dissociating the NIR-cargo complex.²² Cargo is thus released into the nucleus where a high concentration of RNA keeps the incoming RBP soluble so it can perform its regular function, and the NIR is recycled for further rounds of nuclear transport.^{22,30–32} By contrast, in the cytoplasm Ran is found in the GDP-bound state, which has a low affinity for NIRs, permitting NIR-cargo interactions.²²

Beyond this classical function in nuclear transport, NIRs are now understood to operate in the cytoplasm to chaperone and disaggregate NLS-bearing clients.^{19,33–43} In this context, NIRs engage cognate NLSs to inhibit and reverse physiological and deleterious phase transitions of their cargo (Figure 1(B)), which include several prominent neurodegenerative disease-linked RBPs with PrLDs, including wild-type and disease-linked mutant forms of TDP-43, FUS, EWSR1, TAF15, hnRNPA1, and hnRNPA2.^{19,33–44} For example, Karyopherin- $\beta 2$ (Kap $\beta 2$; also known as Transportin 1) can prevent and reverse fibrillization of wild-type

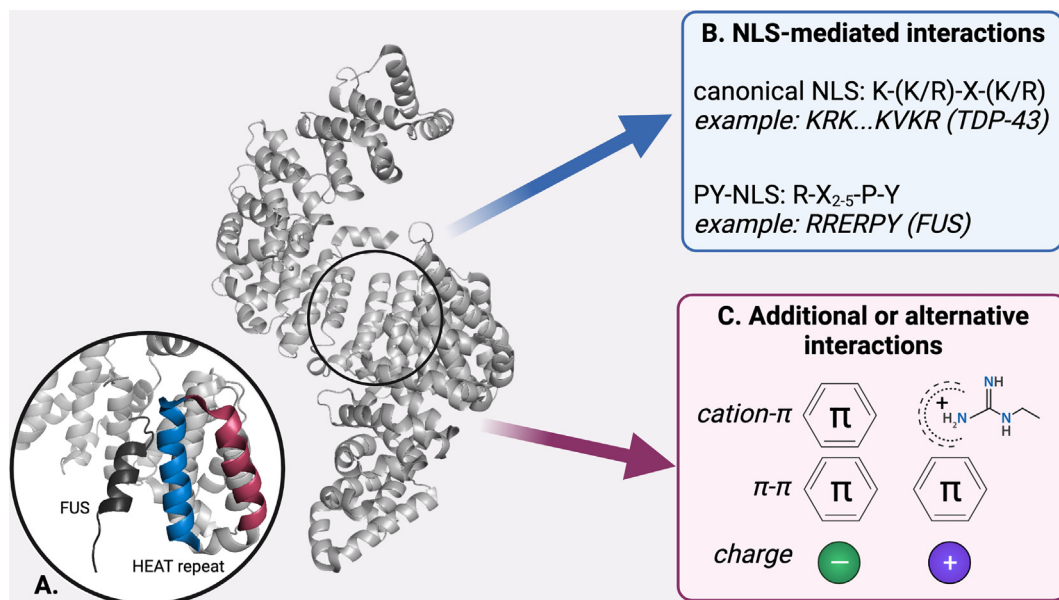


Figure 1. Specific and general interactions enable NIRs to chaperone cargo. (A) NIRs like Kap $\beta 2$ (PDB: 4FDD) share a similar structure, with roughly 20 paired helical HEAT repeats coiled into an alpha solenoid structure. HEAT repeats comprise an outer helix (raspberry) and an inner, cargo-facing helix (blue; FUS-PY-NLS cargo, black). NIRs interact with cargo via two non-exclusive mechanisms. (B) Many NIRs will specifically interact with cargo bearing an NLS. For example, Kap $\beta 2$ will interact with cargo bearing a PY-NLS, like FUS and hnRNPA1. (C) NIRs can also bind to cargo through additional or alternative interactions, including cation- π , π - π , and electrostatic interactions. Such interactions can occur with cargo that bears an NLS, and with cargo that has no known NLS sequence, such as the arginine-rich DPRs produced in c9ALS/FTD. This figure was made with BioRender.

FUS, EWSR1, TAF15, hnRNPA1, hnRNPA2, and several disease-linked variants.¹⁹ Kap β 2 also prevents and reverses FUS liquid–liquid phase separation (LLPS).^{19,33,34,36} Moreover, Importin α (Imp α and Kap β 1 (also known as importin β) cooperate to prevent and reverse TDP-43 condensation and fibrillization.^{19,37} Importantly, elevated NIR expression can mitigate degenerative phenotypes connected with aberrant aggregation of RBPs with PrLDs in model systems.^{19,34,45} Indeed, NIRs can disaggregate cytoplasmic inclusions formed by RBPs with PrLDs and return these proteins to the nucleus, thereby restoring their native function.¹⁹ In this way, NIRs may simultaneously eliminate: (1) any gain of toxic function due to cytoplasmic mislocalization and aggregation of the RBP; and (2) any loss of nuclear RBP function, two facets of disease that likely conspire to drive neurodegeneration.¹⁹ Thus, NIRs join a growing class of ATP-independent protein disaggregases.^{14,46–49} These exciting advances have been reviewed in detail elsewhere.^{16–18,20,50–52}

In this review, we focus on recent developments concerning how NIRs can antagonize aberrant interactions and toxicity of dipeptide-repeat proteins (DPRs) that are produced via repeat-associated non-AUG (RAN) translation⁵³ of the G₄C₂ hexanucleotide repeat expansion (HRE) in *C9ORF72* that cause ALS and FTD.^{37,54} We also highlight recent findings that multiple NIR family members can prevent and reverse the LLPS of specific cargo bearing RGG motifs.³⁵ In ALS/FTD and related degenerative disorders, NIRs can be mutated,⁵⁵ expressed at lower levels,^{52,56} sequestered in stress granules (SGs)⁵⁷ and aggregated structures,^{52,58–60} or fail to effectively recognize post-translationally modified cargo⁶¹ or disease-linked mutant NLSs.^{19,62–68} Moreover, NIRs are critical for neuronal maintenance and function, and mutations in NIRs are associated with human developmental delays, neurologic deficits, and dysmorphic features.⁶⁹ Thus, we close the review by discussing strategies to enhance NIR activity or expression, which could have therapeutic utility for several presently untreatable disorders.¹⁶

NIRs as Safeguards against Toxic DPRs

A large G₄C₂ HRE in the first intron of the *C9ORF72* gene is the most common genetic cause of ALS and FTD (termed c9ALS/FTD).^{10,70–73} Patients with c9ALS/FTD can have hundreds to thousands of G₄C₂ repeats in the first intron of *C9ORF72*, whereas healthy individuals typically harbor ~2–23 repeats.^{10,70–74} In c9ALS/FTD, the G₄C₂ HRE is bidirectionally transcribed into toxic repeat RNAs, which are RAN-translated to yield five different DPRs: poly(GA), poly(GP), poly(PR), poly(GR), and poly(PA).^{71,75–77} In c9ALS/FTD models, arginine-rich DPRs (R-DPRs) are particularly toxic

to neurons due to their positive charge and wide range of interacting partners.^{78–81} More specifically, poly(GR) and poly(PR) can directly interact with the PrLD-containing RBP TDP-43, altering its phase-separation behavior and accelerating its aggregation both *in vitro* and in cells.^{37,82} In fact, poly(GR) and poly(PR) are notorious for their ability to disrupt the LLPS of multiple RBPs through interactions with low-complexity domains (LCDs), and their ability to disturb the dynamics of several membraneless organelles, including SGs, nucleoli, nuclear speckles, Cajal bodies, and heterochromatin.^{71,80,81}

Given the high affinity of NIRs to arginine- or lysine-rich NLSs, it was postulated that NIRs might also target R-DPRs.³⁷ Several NIRs were identified as modulators of R-DPR toxicity in c9ALS/FTD models, suggesting a mechanistic link between NIRs and R-DPRs.^{54,83–85} More recent studies found that R-DPRs directly interact with multiple NIRs, including Imp α , Kap β 1, and Kap β 2, causing an interruption in nucleocytoplasmic trafficking.^{37,86} Specifically, high concentrations of R-DPRs promote the insolubility of NIRs, disrupting their ability to bind and import their NLS-containing cargo.^{37,86} As such, this mechanism provides an explanation for why TDP-43 nuclear import deteriorates in c9ALS/FTD.³⁷

With the direct link between DPRs and NIRs now revealed, can we steer the chaperoning power of NIRs to combat R-DPR-associated toxicity? R-DPRs undergo RNA-stimulated phase separation,⁸⁷ which likely leads to R-DPR accumulation in aggregated structures in c9ALS/FTD.^{76,77} Importantly, Kap β 1 and Kap β 2 suppress RNA-stimulated poly(GR) condensation, whereas Imp α 3 is ineffective.³⁷ Interestingly, while R-DPRs in molar excess can directly interact with NIRs and impair TDP-43 nuclear import, equimolar or elevated levels of Kap β 1 or Kap β 2 can shield R-DPRs, thereby suppressing their pathological interactions with TDP-43.³⁷ Thus, increasing the concentration of NIRs prevents R-DPR phase separation, prevents R-DPRs from engaging in deleterious interactions, and also restores the nuclear localization of TDP-43 as it becomes available to interact with its own NIRs.³⁷ These findings suggest that the reported reduction in the endogenous concentrations of NIRs associated with neurodegenerative disease may contribute to the pathogenesis of c9ALS/FTD, and that interventions aiming to elevate NIR levels are a promising therapeutic strategy to combat R-DPR toxicity in c9ALS/FTD.⁵²

Besides R-DPRs, another c9ALS/FTD-linked DPR, poly(GA), as well as chimeric DPR species such as GA:GP can form cytoplasmic inclusions that may inhibit nuclear import of TDP-43.^{88,89} Indeed, in hippocampal neurons, poly(GA) expression results in robust TDP-43 cytoplasmic mislocalization.⁸⁸ However, overexpression of Imp α 3 or Imp α 4, which may be involved in nuclear import of TDP-43,^{56,90} can likely restore TDP-43 to the

nucleus.⁸⁸ Other repeat-expansion disorders can also produce DPRs, such as spinocerebellar ataxia 36 (SCA36), which presents with poly(PR) and poly(GP).^{89,91} Thus, NIRs may also be promising therapeutics to mitigate DPR toxicity in SCA36.

Multiple NIRs Exhibit Chaperone and Disaggregation Activity

The family of Kap β proteins is large, containing over a dozen subfamilies.⁹² These proteins vary in their structure, directionality of transport (nuclear-export factors such as Crm1 are also members of the Kap β family), and cargo repertoire.⁹² And, whereas some cargo proteins show a clear preference for a single NIR, others can be ferried by multiple karyopherins, either individually or in concert with one another.^{93–99} In addition to trafficking cargo proteins in and out of the nucleus, Kap β family members like Kap β 2 can also act to prevent and reverse the self-assembly and aggregation of proteins, including those implicated in neurodegenerative disease.¹⁶ However, it had been an unexplored question as to whether this disaggregation activity was a feature of karyopherin proteins in general, or an exclusive capability of only some. Focusing on the disease-associated RBP FUS, recent work from Baade et al. now establishes that multiple Kap β family members can act as potent chaperones both *in vitro* and in cells.³⁵

To uncover the network of Kap β proteins that FUS interacts with, Baade et al. performed pull-down assays using cell lysates or purified proteins and found that, in addition to Kap β 2, FUS also binds to Kap β 1, transportin-3, importin-7, importin-13, and exportin-4.³⁵ Like Kap β 2, transportin-3, and importin-7 (either on its own or as a heterodimer with Kap β 1) interact stably with FUS, forming complexes that were sensitive to the addition of Ran-GTP.³⁵ Although these complexes were stable, of the NIRs tested, Kap β 2 demonstrated the strongest binding to FUS.³⁵ It is well established that Kap β 2 binds to FUS via its PY-NLS,⁶² but it was unclear if these other NIRs were interacting with FUS in the same way. Thus, Baade and colleagues assessed the binding of each NIR to truncated constructs of FUS.³⁵ They found that instead of interacting with the PY-NLS, Kap β 1, transportin-3, importin-7, and Kap β 1/importin-7 interact with FUS via its arginine- and glycine-rich RGG domains (Figure 1(C)).³⁵ Interestingly, when the arginine residues of the RGG domains were mutated to lysine, the binding of Kap β 1, transportin-3, importin-7, and Kap β 1/importin-7 was impaired, underscoring the specific importance of arginine in mediating the interaction between Kap β proteins and their cargo.^{35,36,40,100}

Not only do multiple Kap β proteins bind to FUS, but they can also chaperone its material state.³⁵ In the absence of any NIR, purified FUS protein will undergo LLPS.^{19,33,37,101} Addition of equimolar

levels of Kap β 1, transportin-3, importin 7, or Kap β 1/importin-7 each prevented and reversed FUS LLPS, indicating that these NIRs both bind to and chaperone FUS.³⁵ Baade et al. also observed this chaperoning activity in cells, where addition of any of the NIRs tested was able to suppress the association of exogenous MBP-FUS with stress granules.³⁵ The work from Baade and colleagues illustrates that FUS self-assembly can be modulated by multiple Kap β proteins and suggests that the mechanism by which this chaperone activity occurs may be a universal feature of Kap β family members.³⁵ Although Kap β 1, transportin-3, importin 7, or Kap β 1/importin-7 can prevent and reverse FUS LLPS,³⁵ it remains unclear whether Kap β 1, transportin-3, importin 7, or importin β /7 can reverse the formation of FUS fibrils like Kap β 2.¹⁹ Kap β 2 interacts more avidly with FUS than the other NIRs, which may confer stronger FUS chaperone and nuclear-import activity.³⁵ Further studies into the many Kap β family members and their respective abilities to chaperone disease-related cargo proteins will therefore be an intriguing area for future research.

Of particular interest is Transportin-3. Intriguingly, mutations in transportin-3 have been connected to congenital limb-girdle myopathy,^{55,102–107} indicating that transportin-3 is critical for human health. Transportin-3 binds to multiple proteins with RGG-motifs, including the cold-inducible RNA-binding protein which is involved in responding to cell stress.¹⁰⁰ Transportin-3 has also been shown to chaperone another disease-related cargo, the arginine-rich nuclear-speckle protein SRRM2.^{108,109} In tauopathies, including FTD and Alzheimer's disease, tau disrupts nuclear speckles and sequesters SRRM2 in cytoplasmic inclusions, which reduces SRRM2 splicing activity.^{110,111} Thus, upregulation of transportin-3 might enable extraction of SRRM2 from cytoplasmic tau aggregates and restore SRRM2 to the nucleus, which may mitigate degenerative phenotypes in tauopathies.¹¹²

NIRs also affect phase transitions in physiological contexts. For example, Imp α /Kap β 1 regulate the material state of Targeting Protein for XKlp2 (TPX2), a spindle-assembly factor whose activity must be tightly regulated for proper cell cycle progression.^{44,113} TPX2 condensation promotes its activity, whereas Imp α and Kap β 1, either alone or combined, inhibit TPX2 condensation through a combination of specific binding and general low-affinity interactions.^{44,113}

Interestingly, there are NIR family members that bind to cargo with no defined consensus NLS. For example, Transportin-3 and importin 13 can each bind dozens of cargo proteins with no annotated NLS.¹¹⁴ Surprisingly, despite their close similarity, Transportin-3 and importin 13 have little overlap in which cargo they recognize, and for each NIR, the cargo vary in terms of sequence and structure.¹¹⁴ Hence, there is yet much to uncover with respect to potential NIR: cargo interactions.

Leveraging NIRs as Therapeutic Agents

As newly appreciated dissolvases,¹⁸ Kap β family members represent an exciting target to pursue in developing drugs for diseases related to the adoption of aberrant material states by various proteins.¹⁶ There already exist several compounds that affect Kap β s, however these mainly work to prevent activity by occupying the cargo-binding surface of their target.¹¹⁵ As such, there is an opening to develop compounds that can stimulate Kap β activity (Figure 2A).

What would such compounds look like? One approach could be to target disassembly of Ran-GTP:Kap β complexes in the nucleus to promote Kap β turnover (Figure 2(A, i)).¹¹⁶ Increasing the efficiency with which free Kap β proteins are made available would increase the relative levels of unbound Kap β , thereby potentially increasing their apparent activity.

There is also the opportunity to discover novel small molecules to enhance Kap β activity directly (Figure 2(A, ii)). To this end, employing a high-throughput screening approach to identify compounds that stimulate Kap β activity would be an exciting avenue to pursue. Such a screen could be done *in vitro* using purified proteins. For example, it will be of great interest to screen for drug-like compounds that enhance the ability of Kap β 2 to prevent or reverse FUS fibrillization, or Imp α /Kap β 1 to prevent and reverse TDP-43 fibrillization. Likewise, screening campaigns might also be considered in cell-based models to find drug-like compounds that enhance the ability of NIRs to restore nuclear localization of TDP-43 or FUS in response to stress or disease conditions.

Several structures of Kap β 2 bound to disease-linked cargo, including hnRNPA1, FUS, and ALS-linked FUS variants are available.^{40,62,117} These structures could facilitate the design of small mole-

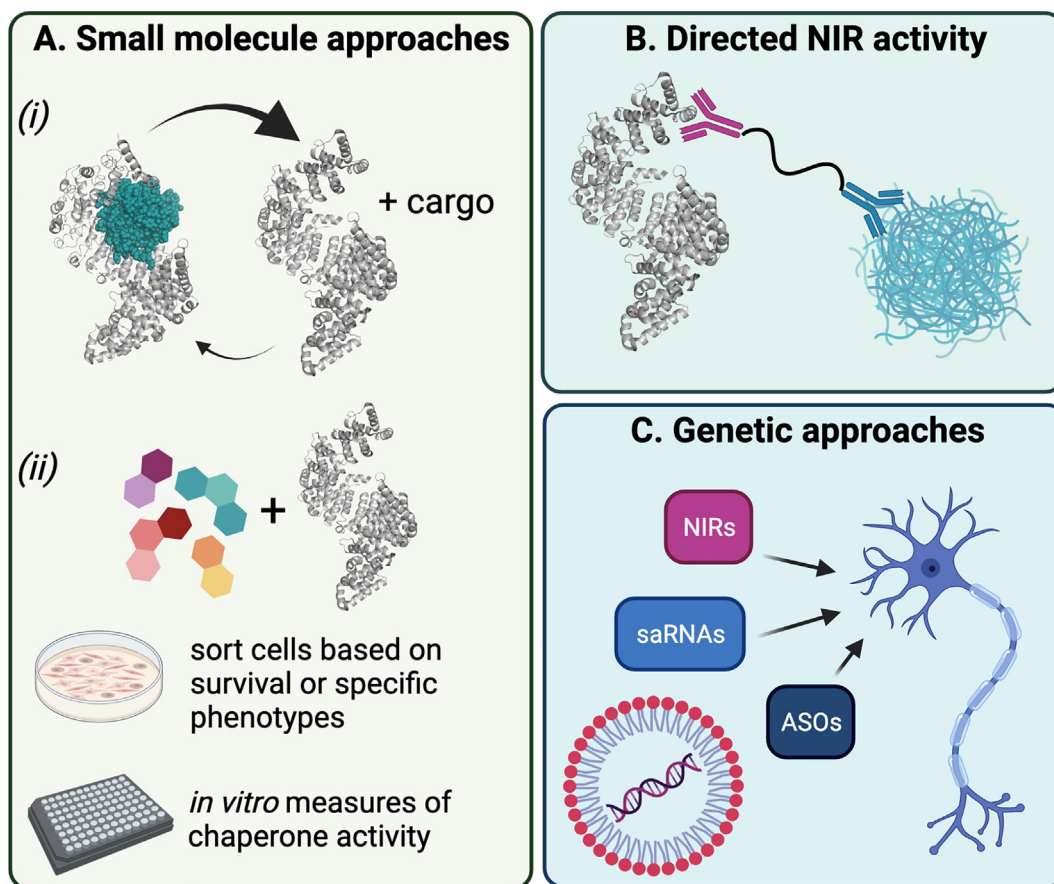


Figure 2. Therapeutic strategies to enhance NIR activity. (A) One potential means by which NIR activity could be enhanced is through the use of small-molecule drugs. For example, a molecule that promotes the release of Ran-GTP (shown in teal) from NIRs in the nucleus would increase the pool of free NIRs available for binding to cargo (i). Alternatively, small molecule libraries or engineered compounds could be used to perform *in vivo* or *in vitro* screens for chaperone activity (ii). (B) Another approach could use a PROTAC-like molecule to direct NIRs to specified targets. (C) Finally, NIR activity could be augmented using genetic approaches to deliver saRNAs or ASOs to elevate NIR expression, or to deliver sequences via AAVs or lipid-containing nanoparticles to express NIRs directly. This figure was made with BioRender.

cules that increase the affinity of Kap β 2 for these specific cargo. This approach may be particularly important for disease-linked hnRNP A1 and FUS variants with mutations in the PY-NLS that weaken the interaction with Kap β 2.^{62,67,68} Ideally, compounds could be uncovered that restore the affinity of Kap β 2 for disease-linked cargo to similar levels observed with wild-type cargo. Although structures of Imp α /Kap β 1 bound to the TDP-43 NLS are not yet available, other structures of Imp α family members bound to cargo have been solved,^{118–123} and these could also inform drug design. Here, it will be important to ensure that small-molecules do not make NIR-cargo interactions so tight that they cannot be dissociated by Ran-GTP, as release of cargo is essential for restoring nuclear activity and NIR recycling.¹⁶

Small-molecule drugs that increase the expression of specific NIRs or even globally upregulate NIRs may also be an interesting therapeutic strategy. How NIR expression is regulated in response to nuclear-transport stress remains poorly understood, and further studies in this area are likely to be informative. For example, it would be important to uncover compounds that globally upregulate nuclear-transport pathways akin to compounds that induce the heat-shock response to upregulate a battery of molecular chaperones.^{124–130} Such compounds could provide a critical boost to nuclear transport to combat neurodegeneration.

Another strategy to boost NIR chaperone activity could resemble recent work with proteolysis-targeting chimeras (PROTACs).¹³¹ PROTACs are small molecules that comprise a moiety that targets an E3-ubiquitin ligase tethered to a recruiting compound that binds to a protein of interest.¹³¹ Thus, PROTACs enable specific ubiquitination and subsequent degradation of a target protein, and have been successfully used in a wide range of applications.¹³¹ However, in ALS/FTD, the disease-relevant proteins TDP-43 and FUS serve vital roles in biology, including in RNA metabolism, axonal transport, and responding to DNA damage and other stressors.^{6,8,34,132–136} A PROTAC-type model that disaggregates but does not degrade its target protein is therefore a potential adaptation well suited to Kap β activity. Here, instead of an E3 ligase, Kap β proteins could be utilized to target phase-separated and aggregated proteins, including those with a mutated NLS or no NLS at all (Figure 2(B)). In promoting a locally high concentration of NIRs, this strategy would liberate the target protein from deleterious assemblies, allowing it to resume its normal activities.

In addition to a compound-based treatment, genetic approaches are also becoming an increasingly tractable strategy for preventing or reversing disease states (Figure 2(C)). For example, delivery of specific NIRs to neurons could be achieved using adeno-associated viruses (AAVs).^{137,138} After a single administration of AAV,

neurons can be successfully transduced, enabling stable expression of a therapeutic gene of interest.¹³⁹ The safety of this approach has been established in numerous clinical trials for neurodegenerative diseases^{140–143} and AAVs delivering specific genes are now FDA-approved drugs.^{144,145} Nonetheless, caution is still needed.^{146,147}

Additional approaches for gene delivery might also be considered, including lipid-containing nanoparticle-mediated delivery of chemically-modified mRNAs to afflicted neurons akin to the technology that has produced highly effective mRNA vaccines (Figure 2(C)).^{15,148,149} Antisense oligonucleotides (ASOs) have also been used to increase the expression of certain proteins by facilitating specific splicing events for productive gene expression (Figure 2(C)).^{150,151} Alternatively, small-activating RNAs (saRNAs) can be used to activate the expression of target genes with the RNA-induced transcriptional activation complex (Figure 2(C)).^{152,153} These approaches could be used to supplement production of NIRs, which undergo age- and disease-related changes in expression levels.^{52,56,154}

Perspectives

It is now clear that NIRs perform a range of beneficial chaperone and dissolvase activities that could, if channeled appropriately, provide profound therapeutic effects for several presently fatal neurodegenerative disorders, including ALS, FTD, MSP, LATE, and tauopathies.^{16,112} However, several challenges lie ahead. These include ensuring that elevating or enhancing NIR activity does not result in unanticipated off-target effects or undesirable effects such as disturbing the nuclear:cytoplasmic ratio of various cargo in a manner that is detrimental to cell viability. Likewise, some cytoplasmic condensates formed by RBPs with PrLDs, such as TDP-43 myogranules¹⁵⁵ or RNA-transport granules^{34,135,156,157}, serve beneficial functions, which ideally would not be perturbed by elevated NIR activity. Indeed, it will be important to disperse pathological condensates and simultaneously preserve beneficial condensates. Despite these challenges, the ability of NIRs to reverse the cytoplasmic mislocalization and aggregation of specific cargo and restore their nuclear localization and function could enable the development of powerful therapeutics, which warrants intense investigation.

CRedit authorship contribution statement

Hana M. Odeh: Visualization, Writing – original draft, Writing – review & editing. **Charlotte M. Fare:** Visualization, Writing – original draft, Writing

– review & editing. **James Shorter:** Visualization, Writing – original draft, Writing – review & editing.

Acknowledgements

We thank Katie Copley and Zarin Tabassum for feedback on the manuscript. H.M.O. is supported by an AstraZeneca post-doctoral fellowship. C.M.F. is supported by National Institutes of Health (NIH) grants T32GM008275 and F31NS111870. J.S. is supported by grants from Target ALS, The Association for Frontotemporal Degeneration, the Amyotrophic Lateral Sclerosis Association, the Office of the Assistant Secretary of Defense for Health Affairs through the Amyotrophic Lateral Sclerosis Research Program (W81XWH-20-1-0242 and W81XWH-17-1-0237), the G. Harold and Leila Y. Mathers Foundation, Sanofi, and NIH grants R01GM099836, R21AG061784, R21AG065854, and R21NS090205.

Declaration of Competing Interest

H.M.O. and C.M.F. declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper. J.S. is a consultant for Dewpoint Therapeutics, Maze Therapeutics, Vivid Sciences, Korro Bio, ADRx, and RBNC Therapeutics.

Received 19 July 2021;

Accepted 24 August 2021;

Available online 28 August 2021

Keywords:

ALS/FTD;
nuclear-import receptor;
FUS;
phase separation;
chaperone

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References

- Portz, B., Lee, B.L., Shorter, J., (2021). FUS and TDP-43 phases in health and disease. *Trends Biochem. Sci.*, **46**, 550–563.
- Cushman, M., Johnson, B.S., King, O.D., Gitler, A.D., Shorter, J., (2010). Prion-like disorders: blurring the divide between transmissibility and infectivity. *J. Cell Sci.*, **123**, 1191–1201.
- King, O.D., Gitler, A.D., Shorter, J., (2012). The tip of the iceberg: RNA-binding proteins with prion-like domains in neurodegenerative disease. *Brain Res.*, **1462**, 61–80.
- Li, Y.R., King, O.D., Shorter, J., Gitler, A.D., (2013). Stress granules as crucibles of ALS pathogenesis. *J. Cell Biol.*, **201**, 361–372.
- March, Z.M., King, O.D., Shorter, J., (2016). Prion-like domains as epigenetic regulators, scaffolds for subcellular organization, and drivers of neurodegenerative disease. *Brain Res.*, **1647**, 9–18.
- Guo, L., Shorter, J., (2017). Biology and pathobiology of TDP-43 and emergent therapeutic strategies. *Cold Spring Harb. Perspect. Med.*, **7**, a024554.
- Harrison, A.F., Shorter, J., (2017). RNA-binding proteins with prion-like domains in health and disease. *Biochem. J.*, **474**, 1417–1438.
- Ling, S.C., Polymenidou, M., Cleveland, D.W., (2013). Converging mechanisms in ALS and FTD: disrupted RNA and protein homeostasis. *Neuron*, **79**, 416–438.
- Lee, E.B., Lee, V.M., Trojanowski, J.Q., (2011). Gains or losses: molecular mechanisms of TDP43-mediated neurodegeneration. *Nature Rev. Neurosci.*, **13**, 38–50.
- Kim, G., Gautier, O., Tassoni-Tsuchida, E., Ma, X.R., Gitler, A.D., (2020). ALS Genetics: Gains, Losses, and Implications for Future Therapies. *Neuron*, **108**, 822–842.
- Jackrel, M.E., Shorter, J., (2015). Engineering enhanced protein disaggregases for neurodegenerative disease. *Prion.*, **9**, 90–109.
- Jackrel, M.E., Shorter, J., (2017). Protein-remodeling factors as potential therapeutics for neurodegenerative disease. *Front. Neurosci.*, **11**, 99.
- Shorter, J., (2008). Hsp104: a weapon to combat diverse neurodegenerative disorders. *Neurosignals*, **16**, 63–74.
- Shorter, J., (2017). Designer protein disaggregases to counter neurodegenerative disease. *Curr. Opin. Genet. Dev.*, **44**, 1–8.
- Shorter, J., (2016). Engineering therapeutic protein disaggregases. *Mol. Biol. Cell*, **27**, 1556–1560.
- Guo, L., Fare, C.M., Shorter, J., (2019). Therapeutic Dissolution of Aberrant Phases by Nuclear-Import Receptors. *Trends Cell Biol.*, **29**, 308–322.
- Darling, A.L., Shorter, J., (2021). Combating deleterious phase transitions in neurodegenerative disease. *Biochim. Biophys. Acta, Mol. Cell. Biol. Res.*, **1868**, 118984.
- Fare, C.M., Shorter, J., (2021). (Dis)Solving the problem of aberrant protein states. *Dis. Model Mech.*, **14**.
- Guo, L., Kim, H.J., Wang, H., Monaghan, J., Freyermuth, F., Sung, J.C., et al., (2018). Nuclear-import receptors reverse aberrant phase transitions of RNA-binding proteins with prion-like domains. *Cell*, **173**, (677–92) e20.
- Springhower, C.E., Rosen, M.K., Chook, Y.M., (2020). Karyopherins and condensates. *Curr. Opin. Cell Biol.*, **64**, 112–123.
- Chook, Y.M., Suel, K.E., (2011). Nuclear import by karyopherin-betas: recognition and inhibition. *Biochim. Biophys. Acta, Mol. Cell. Biol. Lipids*, **1813**, 1593–1606.
- Soniat, M., Chook, Y.M., (2015). Nuclear localization signals for four distinct karyopherin-beta nuclear import systems. *Biochem. J.*, **468**, 353–362.
- Allegretti, M., Zimmerli, C.E., Rantos, V., Wilfling, F., Ronchi, P., Fung, H.K.H., et al., (2020). In-cell architecture of the nuclear pore and snapshots of its turnover. *Nature*, **586**, 796–800.
- Mohr, D., Frey, S., Fischer, T., Guttler, T., Gorlich, D., (2009). Characterisation of the passive permeability barrier of nuclear pore complexes. *EMBO J.*, **28**, 2541–2553.
- Frey, S., Gorlich, D., (2007). A saturated FG-repeat hydrogel can reproduce the permeability properties of nuclear pore complexes. *Cell*, **130**, 512–523.

26. Frey, S., Richter, R.P., Gorlich, D., (2006). FG-rich repeats of nuclear pore proteins form a three-dimensional meshwork with hydrogel-like properties. *Science*, **314**, 815–817.
27. Labokha, A.A., Gradmann, S., Frey, S., Hulsman, B.B., Urlaub, H., Baldus, M., et al., (2013). Systematic analysis of barrier-forming FG hydrogels from *Xenopus* nuclear pore complexes. *EMBO J.*, **32**, 204–218.
28. Celetti, G., Paci, G., Caria, J., VanDelinder, V., Bachand, G., Lemke, E.A., (2020). The liquid state of FG-nucleoporins mimics permeability barrier properties of nuclear pore complexes. *J. Cell Biol.*, **219**, e201907157.
29. Frey, S., Rees, R., Schunemann, J., Ng, S.C., Funfgeld, K., Huyton, T., et al., (2018). Surface properties determining passage rates of proteins through nuclear pores. *Cell*, **174** 202–17 e9.
30. Maharana, S., Wang, J., Papadopoulos, D.K., Richter, D., Pozniakovsky, A., Poser, I., et al., (2018). RNA buffers the phase separation behavior of prion-like RNA binding proteins. *Science*, **360**, 918–921.
31. Mann, J.R., Gleixner, A.M., Mauna, J.C., Gomes, E., DeChellis-Marks, M.R., Needham, P.G., et al., (2019). RNA binding antagonizes neurotoxic phase transitions of TDP-43. *Neuron*, **102** 321–38 e8.
32. Yu, H., Lu, S., Gasior, K., Singh, D., Vazquez-Sanchez, S., Tapia, O., et al., (2021). HSP70 chaperones RNA-free TDP-43 into anisotropic intranuclear liquid spherical shells. *Science*, **371**, eabb4309.
33. Yoshizawa, T., Ali, R., Jiou, J., Fung, H.Y.J., Burke, K.A., Kim, S.J., et al., (2018). Nuclear import receptor inhibits phase separation of FUS through binding to multiple sites. *Cell*, **173** 693–705 e22.
34. Qamar, S., Wang, G., Randle, S.J., Ruggeri, F.S., Varela, J.A., Lin, J.Q., et al., (2018). FUS phase separation is modulated by a molecular chaperone and methylation of arginine cation- π interactions. *Cell*, **173** 720–34 e15.
35. Baade, I., Hutten, S., Sternburg, E.L., Porschke, M., Hofweber, M., Dormann, D., et al., (2021). The RNA-binding protein FUS is chaperoned and imported into the nucleus by a network of import receptors. *J. Biol. Chem.*, **296**, 100659.
36. Hofweber, M., Hutten, S., Bourgeois, B., Spreitzer, E., Niedner-Boblenz, A., Schifferer, M., et al., (2018). Phase separation of FUS is suppressed by its nuclear import receptor and arginine methylation. *Cell*, **173** 706–19 e13.
37. Hutten, S., Usluer, S., Bourgeois, B., Simonetti, F., Odeh, H.M., Fare, C.M., et al., (2020). Nuclear import receptors directly bind to arginine-rich dipeptide repeat proteins and suppress their pathological interactions. *Cell Rep.*, **33**, 108538.
38. Niaki, A.G., Sarkar, J., Cai, X., Rhine, K., Vidaurre, V., Guy, B., et al., (2020). Loss of dynamic RNA interaction and aberrant phase separation induced by two distinct types of ALS/FTD-Linked FUS mutations. *Mol. Cell*, **77** 82–94 e4.
39. Rhine, K., Makurath, M.A., Liu, J., Skanchy, S., Lopez, C., Catalan, K.F., et al., (2020). ALS/FTLD-linked mutations in FUS glycine residues cause accelerated gelation and reduced interactions with wild-type FUS. *Mol. Cell*, **80** 666–81 e8.
40. Gonzalez, A., Mannen, T., Cagatay, T., Fujiwara, A., Matsumura, H., Niesman, A.B., et al., (2021). Mechanism of karyopherin-beta2 binding and nuclear import of ALS variants FUS(P525L) and FUS(R495X). *Sci. Rep.*, **11**, 3754.
41. Jakel, S., Mingot, J.M., Schwarzmaier, P., Hartmann, E., Gorlich, D., (2002). Importins fulfil a dual function as nuclear import receptors and cytoplasmic chaperones for exposed basic domains. *EMBO J.*, **21**, 377–386.
42. Chou, C.C., Zhang, Y., Umoh, M.E., Vaughan, S.W., Lorenzini, I., Liu, F., et al., (2018). TDP-43 pathology disrupts nuclear pore complexes and nucleocytoplasmic transport in ALS/FTD. *Nature Neurosci.*, **21**, 228–239.
43. Miyake, Y., Keusch, J.J., Decamps, L., Ho-Xuan, H., Iketani, S., Gut, H., et al., (2019). Influenza virus uses transportin 1 for vRNP debundling during cell entry. *Nature Microbiol.*, **4**, 578–586.
44. King, M.R., Petry, S., (2020). Phase separation of TPX2 enhances and spatially coordinates microtubule nucleation. *Nature Commun.*, **11**, 270.
45. Jackel, S., Summerer, A.K., Thommes, C.M., Pan, X., Voigt, A., Schulz, J.B., et al., (2015). Nuclear import factor transportin and arginine methyltransferase 1 modify FUS neurotoxicity in *Drosophila*. *Neurobiol. Dis.*, **74**, 76–88.
46. Jaru-Ampornpan, P., Shen, K., Lam, V.Q., Ali, M., Doniach, S., Jia, T.Z., et al., (2010). ATP-independent reversal of a membrane protein aggregate by a chloroplast SRP subunit. *Nature Struct. Mol. Biol.*, **17**, 696–702.
47. Huang, L., Agrawal, T., Zhu, G., Yu, S., Tao, L., Lin, J., et al., (2021). DAXX represents a new type of protein-folding enabler. *Nature*, **597**, 132–137.
48. Zhu, G., Harischandra, D.S., Ghaisas, S., Zhang, P., Prall, W., Huang, L., et al., (2020). TRIM11 prevents and reverses protein aggregation and rescues a mouse model of Parkinson's disease. *Cell Rep.*, **33**, 108418.
49. Baker, J.D., Shelton, L.B., Zheng, D., Favretto, F., Nordhues, B.A., Darling, A., et al., (2017). Human cyclophilin 40 unravels neurotoxic amyloids. *PLoS Biol.*, **15**, e2001336.
50. Yoshizawa, T., Guo, L., (2021). Karyopherin-betas play a key role as a phase separation regulator. *J. Biochem.*,.
51. Mikhaleva, S., Lemke, E.A., (2018). Beyond the transport function of import receptors: what's all the FUS about? *Cell*, **173**, 549–553.
52. Pasha, T., Zatorska, A., Sharipov, D., Rogelj, B., Hortobagyi, T., Hirth, F., (2021). Karyopherin abnormalities in neurodegenerative proteinopathies. *Brain*,. <https://doi.org/10.1093/brain/awab201>.
53. Zu, T., Gibbens, B., Doty, N.S., Gomes-Pereira, M., Huguet, A., Stone, M.D., et al., (2011). Non-ATG-initiated translation directed by microsatellite expansions. *PNAS*, **108**, 260–265.
54. Jovicic, A., Mertens, J., Boeynaems, S., Bogaert, E., Chai, N., Yamada, S.B., et al., (2015). Modifiers of C9orf72 dipeptide repeat toxicity connect nucleocytoplasmic transport defects to FTD/ALS. *Nature Neurosci.*, **18**, 1226–1229.
55. Vihola, A., Palmio, J., Danielsson, O., Penttilä, S., Louiselle, D., Pittman, S., et al., (2019). Novel mutation in TNPO3 causes congenital limb-girdle myopathy with slow progression. *Neurol Genet.*, **5**, e337.
56. Nishimura, A.L., Zupunski, V., Troakes, C., Kathe, C., Fratta, P., Howell, M., et al., (2010). Nuclear import impairment causes cytoplasmic trans-activation response DNA-binding protein accumulation and is associated with

- frontotemporal lobar degeneration. *Brain*, **133**, 1763–1771.
57. Zhang, K., Daigle, J.G., Cunningham, K.M., Coyne, A.N., Ruan, K., Grima, J.C., et al., (2018). Stress granule assembly disrupts nucleocytoplasmic transport. *Cell*, **173**, 958–71 e17.
 58. Solomon, D.A., Stepto, A., Au, W.H., Adachi, Y., Diaper, D.C., Hall, R., et al., (2018). A feedback loop between dipeptide-repeat protein, TDP-43 and karyopherin- α mediates C9orf72-related neurodegeneration. *Brain*, **141**, 2908–2924.
 59. Troakes, C., Hortobagyi, T., Vance, C., Al-Sarraj, S., Rogelj, B., Shaw, C.E., (2013). Transportin 1 colocalization with Fused in Sarcoma (FUS) inclusions is not characteristic for amyotrophic lateral sclerosis-FUS confirming disrupted nuclear import of mutant FUS and distinguishing it from frontotemporal lobar degeneration with FUS inclusions. *Neuropathol. Appl. Neurobiol.*, **39**, 553–561.
 60. Neumann, M., Valori, C.F., Ansorge, O., Kretschmar, H. A., Munoz, D.G., Kusaka, H., et al., (2012). Transportin 1 accumulates specifically with FET proteins but no other transportin cargos in FTL-D-FUS and is absent in FUS inclusions in ALS with FUS mutations. *Acta Neuropathol.*, **124**, 705–716.
 61. Dormann, D., Madl, T., Valori, C.F., Bentmann, E., Tahirovic, S., Abou-Ajam, C., et al., (2012). Arginine methylation next to the PY-NLS modulates Transportin binding and nuclear import of FUS. *EMBO J.*, **31**, 4258–4275.
 62. Zhang, Z.C., Chook, Y.M., (2012). Structural and energetic basis of ALS-causing mutations in the atypical proline-tyrosine nuclear localization signal of the Fused in Sarcoma protein (FUS). *Proc. Natl. Acad. Sci. USA*, **109**, 12017–12021.
 63. Dormann, D., Rodde, R., Edbauer, D., Bentmann, E., Fischer, I., Hruscha, A., et al., (2010). ALS-associated fused in sarcoma (FUS) mutations disrupt Transportin-mediated nuclear import. *EMBO J.*, **29**, 2841–2857.
 64. Naruse, H., Ishiura, H., Mitsui, J., Date, H., Takahashi, Y., Matsukawa, T., et al., (2018). Molecular epidemiological study of familial amyotrophic lateral sclerosis in Japanese population by whole-exome sequencing and identification of novel HNRNPA1 mutation. *Neurobiol. Aging*, **61** 255.e9–e16.
 65. Liu, Q., Shu, S., Wang, R.R., Liu, F., Cui, B., Guo, X.N., et al., (2016). Whole-exome sequencing identifies a missense mutation in hnRNPA1 in a family with flail arm ALS. *Neurology*, **87**, 1763–1769.
 66. Bosco, D.A., Lemay, N., Ko, H.K., Zhou, H., Burke, C., Kwiatkowski Jr, T.J., et al., (2010). Mutant FUS proteins that cause amyotrophic lateral sclerosis incorporate into stress granules. *Hum. Mol. Genet.*, **19**, 4160–4175.
 67. Beijer, D., Kim, H., Guo, L., O'Donovan, K., Mademan, I., Deconinck, T., et al., (2021). Characterization of HNRNPA1 mutations defines diversity in pathogenic mechanisms and clinical presentation. *JCI Insight*, **6**, e148363. 2021.02.02.21250330.
 68. Kim, H.J., Mohassel, P., Donkervoort, S., Guo, L., O'Donovan, K., Coughlin, M., et al., (2021). Specific heterozygous frameshift variants in hnRNPA2B1 cause early-onset oculopharyngeal muscular dystrophy. *medRxiv*, 2021.04.08.21254942.
 69. Goodman, L.D., Cope, H., Nil, Z., Ravenscroft, T.A., Charnig, W.L., Lu, S., et al., (2021). TNPO2 variants associate with human developmental delays, neurologic deficits, and dysmorphic features and alter TNPO2 activity in *Drosophila*. *Am. J. Hum. Genet.*, **108**, 1669–1691.
 70. DeJesus-Hernandez, M., Mackenzie, I.R., Boeve, B.F., Boxer, A.L., Baker, M., Rutherford, N.J., et al., (2011). Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS. *Neuron*, **72**, 245–256.
 71. Odeh, H.M., Shorter, J., (2020). Arginine-rich dipeptide-repeat proteins as phase disruptors in C9-ALS/FTD. *Emerg Top Life Sci.*, **4**, 293–305.
 72. Renton, A.E., Majounie, E., Waite, A., Simon-Sanchez, J., Rollinson, S., Gibbs, J.R., et al., (2011). A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron*, **72**, 257–268.
 73. Gitler, A.D., Tsuiji, H., (2016). There has been an awakening: Emerging mechanisms of C9orf72 mutations in FTD/ALS. *Brain Res.*, **1647**, 19–29.
 74. Iacoangeli, A., Al Khleifat, A., Jones, A.R., Sproviero, W., Shatunov, A., Opie-Martin, S., et al., (2019). C9orf72 intermediate expansions of 24–30 repeats are associated with ALS. *Acta Neuropathol. Commun.*, **7**, 115.
 75. Mori, K., Arzberger, T., Grässer, F.A., Gijssels, I., May, S., Rentzsch, K., et al., (2013). Bidirectional transcripts of the expanded C9orf72 hexanucleotide repeat are translated into aggregating dipeptide repeat proteins. *Acta Neuropathol.*, **126**, 881–893.
 76. Mori, K., Weng, S.M., Arzberger, T., May, S., Rentzsch, K., Kremmer, E., et al., (2013). The C9orf72 GGGGCC repeat is translated into aggregating dipeptide-repeat proteins in FTL/D/ALS. *Science*, **339**, 1335–1338.
 77. Zu, T., Liu, Y., Bañez-Coronel, M., Reid, T., Pletnikova, O., Lewis, J., et al., (2013). RAN proteins and RNA foci from antisense transcripts in C9ORF72 ALS and frontotemporal dementia. *PNAS*, **110**, E4968–E4977.
 78. Mizielinska, S., Grönke, S., Niccoli, T., Ridler, C.E., Clayton, E.L., Devoy, A., et al., (2014). C9orf72 repeat expansions cause neurodegeneration in *Drosophila* through arginine-rich proteins. *Science (New York, NY)*, **345**, 1192–1194.
 79. Wen, X., Tan, W., Westergard, T., Krishnamurthy, K., Markandaiah, S.S., Shi, Y., et al., (2014). Antisense proline-arginine RAN dipeptides linked to C9ORF72-ALS/FTD form toxic nuclear aggregates that initiate in vitro and in vivo neuronal death. *Neuron*, **84**, 1213–1225.
 80. Lee, K.-H., Zhang, P., Kim, H.J., Mitrea, D.M., Sarkar, M., Freibaum, B.D., et al., (2016). C9orf72 dipeptide repeats impair the assembly, dynamics, and function of membrane-less organelles. *Cell*, **167** 774–88.e17.
 81. Zhang, Y.J., Guo, L., Gonzales, P.K., Gendron, T.F., Wu, Y., Jansen-West, K., et al., (2019). Heterochromatin anomalies and double-stranded RNA accumulation underlie C9orf72 poly(PR) toxicity. *Science*, **363**.
 82. Cook, C.N., Wu, Y., Odeh, H.M., Gendron, T.F., Jansen-West, K., Del Rosso, G., et al., (2020). C9orf72 poly(GR) aggregation induces TDP-43 proteinopathy. *Sci. Transl. Med.*, **12**.
 83. Boeynaems, S., Bogaert, E., Michiels, E., Gijssels, I., Sieben, A., Jovicic, A., et al., (2016). *Drosophila* screen connects nuclear transport genes to DPR pathology in c9ALS/FTD. *Sci. Rep.*, **6**, 20877.

84. Freibaum, B.D., Lu, Y., Lopez-Gonzalez, R., Kim, N.C., Almeida, S., Lee, K.-H., et al., (2015). GGGGCC repeat expansion in C9orf72 compromises nucleocytoplasmic transport. *Nature*, **525**, 129–133.
85. Zhang, K., Donnelly, C.J., Haeusler, A.R., Grima, J.C., Machamer, J.B., Steinwald, P., et al., (2015). The C9orf72 repeat expansion disrupts nucleocytoplasmic transport. *Nature*, **525**, 56–61.
86. Hayes, L.R., Duan, L., Bowen, K., Kalab, P., Rothstein, J. D., (2020). C9orf72 arginine-rich dipeptide repeat proteins disrupt karyopherin-mediated nuclear import. *eLife*, **9**, e51685.
87. Boeynaems, S., Holehouse, A.S., Weinhardt, V., Kovacs, D., Van Lindt, J., Larabell, C., et al., (2019). Spontaneous driving forces give rise to protein-RNA condensates with coexisting phases and complex material properties. *Proc. Natl. Acad. Sci. USA*, **116**, 7889–7898.
88. Khosravi, B., Hartmann, H., May, S., Möhl, C., Ederle, H., Michaelsen, M., et al., (2017). Cytoplasmic poly-GA aggregates impair nuclear import of TDP-43 in C9orf72 ALS/FTLD. *Hum. Mol. Genet.*, **26**, 790–800.
89. McEachin, Z.T., Gendron, T.F., Raj, N., García-Murias, M., Banerjee, A., Purcell, R.H., et al., (2020). Chimeric peptide species contribute to divergent dipeptide repeat pathology in c9ALS/FTD and SCA36. *Neuron*, **107**, 292–305.e6.
90. Park, J.H., Chung, C.G., Park, S.S., Lee, D., Kim, K.M., Jeong, Y., et al., (2020). Cytosolic calcium regulates cytoplasmic accumulation of TDP-43 through Calpain-A and Importin α 3. *eLife*, **9**, e60132.
91. Todd, T.W., McEachin, Z.T., Chew, J., Burch, A.R., Jansen-West, K., Tong, J., et al., (2020). Hexanucleotide repeat expansions in c9FTD/ALS and SCA36 Confer selective patterns of neurodegeneration in vivo. *Cell Rep.*, **31**, 107616.
92. O'Reilly, A.J., Dacks, J.B., Field, M.C., (2011). Evolution of the karyopherin- β family of nucleocytoplasmic transport factors; ancient origins and continued specialization. *PLoS ONE*, **6**, e19308.
93. Arnold, M., Nath, A., Hauber, J., Kehlenbach, R.H., (2006). Multiple importins function as nuclear transport receptors for the Rev protein of human immunodeficiency virus type 1. *J. Biol. Chem.*, **281**, 20883–20890.
94. Baake, M., Bauerle, M., Doenecke, D., Albig, W., (2001). Core histones and linker histones are imported into the nucleus by different pathways. *Eur. J. Cell Biol.*, **80**, 669–677.
95. Ivic, N., Potocnjak, M., Solis-Mezarino, V., Herzog, F., Bilokapic, S., Halic, M., (2019). Fuzzy interactions form and shape the histone transport complex. *Mol. Cell*, **73**, 1191–203 e6.
96. Jakel, S., Albig, W., Kutay, U., Bischoff, F.R., Schwamborn, K., Doenecke, D., et al., (1999). The importin beta/importin 7 heterodimer is a functional nuclear import receptor for histone H1. *EMBO J.*, **18**, 2411–2423.
97. Muhlhäusser, P., Müller, E.C., Otto, A., Kutay, U., (2001). Multiple pathways contribute to nuclear import of core histones. *EMBO Rep.*, **2**, 690–696.
98. Waldmann, I., Walde, S., Kehlenbach, R.H., (2007). Nuclear import of c-Jun is mediated by multiple transport receptors. *J. Biol. Chem.*, **282**, 27685–27692.
99. Freedman, N.D., Yamamoto, K.R., (2004). Importin 7 and importin alpha/importin beta are nuclear import receptors for the glucocorticoid receptor. *Mol. Biol. Cell*, **15**, 2276–2286.
100. Bourgeois, B., Hutten, S., Gottschalk, B., Hofweber, M., Richter, G., Sternat, J., et al., (2020). Nonclassical nuclear localization signals mediate nuclear import of CIRBP. *Proc. Natl. Acad. Sci. USA*, **117**, 8503–8514.
101. Patel, A., Lee, H.O., Jawerth, L., Maharana, S., Jahnel, M., Hein, M.Y., et al., (2015). A Liquid-to-solid phase transition of the ALS protein FUS accelerated by disease mutation. *Cell*, **162**, 1066–1077.
102. Angelini, C., Marozzo, R., Pinzan, E., Pegoraro, V., Molnar, M.J., Torella, A., et al., (2019). A new family with transportinopathy: increased clinical heterogeneity. *Ther. Adv. Neurol. Disord.*, **12** 1756286419850433.
103. Pál, E., Zima, J., Hadzsiev, K., Ito, Y.A., Hartley, T., Boycott, K.M., et al., (2019). A novel pathogenic variant in TNPO3 in a Hungarian family with limb-girdle muscular dystrophy 1F. *Eur. J. Med. Genet.*, **62**, 103662.
104. Rodríguez-Mora, S., De Wit, F., García-Pérez, J., Bermejo, M., López-Huertas, M.R., Mateos, E., et al., (2019). The mutation of Transportin 3 gene that causes limb girdle muscular dystrophy 1F induces protection against HIV-1 infection. *PLoS Pathog.*, **15**, e1007958.
105. Melià, M.J., Kubota, A., Ortolano, S., Vílchez, J.J., Gámez, J., Tanji, K., et al., (2013). Limb-girdle muscular dystrophy 1F is caused by a microdeletion in the transportin 3 gene. *Brain*, **136**, 1508–1517.
106. Torella, A., Fanin, M., Mutarelli, M., Peterle, E., Del Vecchio, B.F., Rispoli, R., et al., (2013). Next-generation sequencing identifies transportin 3 as the causative gene for LGMD1F. *PLoS ONE*, **8**, e63536.
107. Costa, R., Rodia, M.T., Vianello, S., Santi, S., Lattanzi, G., Angelini, C., et al., (2020). Transportin 3 (TNPO3) and related proteins in limb girdle muscular dystrophy D2 muscle biopsies: a morphological study and pathogenetic hypothesis. *Neuromuscul. Disord.*, **30**, 685–692.
108. Kimura, M., Morinaka, Y., Imai, K., Kose, S., Horton, P., Imamoto, N., (2017). Extensive cargo identification reveals distinct biological roles of the 12 importin pathways. *Elife*, **6**.
109. Kataoka, N., Bachorik, J.L., Dreyfuss, G., (1999). Transportin-SR, a nuclear import receptor for SR proteins. *J. Cell Biol.*, **145**, 1145–1152.
110. Lester, E., Ooi, F.K., Bakkar, N., Ayers, J., Woerman, A. L., Wheeler, J., et al., (2021). Tau aggregates are RNA-protein assemblies that mislocalize multiple nuclear speckle components. *Neuron*, **109**, 1675–91 e9.
111. McMillan, P.J., Strovass, T.J., Baum, M., Mitchell, B.K., Eck, R.J., Hendricks, N., et al., (2021). Pathological tau drives ectopic nuclear speckle scaffold protein SRRM2 accumulation in neuron cytoplasm in Alzheimer's disease. *Acta Neuropathol. Commun.*, **9**, 117.
112. Gorenberg, E.L., Shorter, J., (2021). Tau heckles speckles: A pathogenic mechanism in tauopathy? *Neuron*, **109**, 1585–1587.
113. Safari, M.S., King, M.R., Brangwynne, C.P., Petry, S., (2021). Interaction of spindle assembly factor TPX2 with importins-alpha/beta inhibit protein phase separation. *J. Biol. Chem.*, 100998.
114. Kimura, M., Imai, K., Morinaka, Y., Hosono-Sakuma, Y., Horton, P., Imamoto, N., (2021). Distinct mutations in importin- β family nucleocytoplasmic transport receptors transportin-SR and importin-13 affect specific cargo binding. *Sci. Rep.*, **11**, 15649.

115. Kosyna, F.K., Depping, R., (2018). Controlling the gatekeeper: therapeutic targeting of nuclear transport. *Cells*, **7**.
116. Floer, M., Blobel, G., Rexach, M., (1997). Disassembly of RanGTP-karyopherin beta complex, an intermediate in nuclear protein import. *J. Biol. Chem.*, **272**, 19538–19546.
117. Lee, B.J., Cansizoglu, A.E., Suel, K.E., Louis, T.H., Zhang, Z., Chook, Y.M., (2006). Rules for nuclear localization sequence recognition by karyopherin beta 2. *Cell*, **126**, 543–558.
118. Smith, K.M., Tsimbalyuk, S., Edwards, M.R., Cross, E.M., Batra, J., Soares da Costa, T.P., et al., (2018). Structural basis for importin alpha 3 specificity of W proteins in Hendra and Nipah viruses. *Nature Commun.*, **9**, 3703.
119. Koyama, M., Matsuura, Y., (2017). Crystal structure of importin- α bound to the nuclear localization signal of Ran-binding protein 3. *Biochem. Biophys. Res. Commun.*, **491**, 609–613.
120. Jung, H., Takeshima, T., Nakagawa, T., MacMillan, K.S., Wynn, R.M., Wang, H., et al., (2020). The structure of importin α and the nuclear localization peptide of ChREBP, and small compound inhibitors of ChREBP-importin α interactions. *Biochem. J.*, **477**, 3253–3269.
121. Kirby, T.W., Pedersen, L.C., Gabel, S.A., Gassman, N.R., London, R.E., (2018). Variations in nuclear localization strategies among pol X family enzymes. *Traffic*.
122. Kirby, T.W., Gassman, N.R., Smith, C.E., Pedersen, L.C., Gabel, S.A., Sobhany, M., et al., (2015). Nuclear Localization of the DNA Repair Scaffold XRCC1: Uncovering the Functional Role of a Bipartite NLS. *Sci. Rep.*, **5**, 13405.
123. Jeong, S.A., Kim, K., Lee, J.H., Cha, J.S., Khadka, P., Cho, H.S., et al., (2015). Akt-mediated phosphorylation increases the binding affinity of hTERT for importin α to promote nuclear translocation. *J. Cell Sci.*, **128**, 2287–2301.
124. Neef, D.W., Jaeger, A.M., Gomez-Pastor, R., Willmund, F., Frydman, J., Thiele, D.J., (2014). A direct regulatory interaction between chaperonin TRiC and stress-responsive transcription factor HSF1. *Cell Rep.*, **9**, 955–966.
125. Vigh, L., Literati, P.N., Horvath, I., Torok, Z., Balogh, G., Glatz, A., et al., (1997). Bimocromol: a nontoxic, hydroxylamine derivative with stress protein-inducing activity and cytoprotective effects. *Nature Med.*, **3**, 1150–1154.
126. Hargitai, J., Lewis, H., Boros, I., Rácz, T., Fiser, A., Kurucz, I., et al., (2003). Bimocromol, a heat shock protein co-inducer, acts by the prolonged activation of heat shock factor-1. *Biochem. Biophys. Res. Commun.*, **307**, 689–695.
127. Kieran, D., Kalmar, B., Dick, J.R., Riddoch-Contreras, J., Burnstock, G., Greensmith, L., (2004). Treatment with arimocromol, a coinducer of heat shock proteins, delays disease progression in ALS mice. *Nature Med.*, **10**, 402–405.
128. Ahmed, M., Machado, P.M., Miller, A., Spicer, C., Herbelin, L., He, J., et al., (2016). Targeting protein homeostasis in sporadic inclusion body myositis. *Sci. Transl. Med.*, **8**, 331ra41.
129. Jackrel, M.E., Shorter, J., (2011). Shock and awe: unleashing the heat shock response to treat Huntington disease. *J. Clin. Invest.*, **121**, 2972–2975.
130. Auluck, P.K., Bonini, N.M., (2002). Pharmacological prevention of Parkinson disease in *Drosophila*. *Nature Med.*, **8**, 1185–1186.
131. Burslem, G.M., Crews, C.M., (2020). Proteolysis-targeting chimeras as therapeutics and tools for biological discovery. *Cell*, **181**, 102–114.
132. Levone, B.R., Lenzken, S.C., Antonaci, M., Maiser, A., Rapp, A., Conte, F., et al., (2021). FUS-dependent liquid-liquid phase separation is important for DNA repair initiation. *J. Cell Biol.*, **220**.
133. Mitra, J., Guerrero, E.N., Hegde, P.M., Liachko, N.F., Wang, H., Vasquez, V., et al., (2019). Motor neuron disease-associated loss of nuclear TDP-43 is linked to DNA double-strand break repair defects. *Proc. Natl. Acad. Sci. USA*, **116**, 4696–4705.
134. Alami, N.H., Smith, R.B., Carrasco, M.A., Williams, L.A., Winborn, C.S., Han, S.S.W., et al., (2014). Axonal transport of TDP-43 mRNA granules is impaired by ALS-causing mutations. *Neuron*, **81**, 536–543.
135. Gopal, P.P., Nirschl, J.J., Klinman, E., Holzbaur, E.L., (2017). Amyotrophic lateral sclerosis-linked mutations increase the viscosity of liquid-like TDP-43 RNP granules in neurons. *Proc. Natl. Acad. Sci. USA*, **114**, E2466–E2475.
136. Hallegger, M., Chakrabarti, A.M., Lee, F.C.Y., Lee, B.L., Amaliotti, A.G., Odeh, H.M., et al., (2021). TDP-43 condensation properties specify its RNA-binding and regulatory repertoire. *Cell*, **184**, 4680–4696.e22.
137. Deverman, B.E., Ravina, B.M., Bankiewicz, K.S., Paul, S. M., Sah, D.W.Y., (2018). Gene therapy for neurological disorders: progress and prospects. *Nature Rev. Drug Discovery*, **17**, 641–659.
138. Bravo-Hernandez, M., Tadokoro, T., Navarro, M.R., Platoshyn, O., Kobayashi, Y., Marsala, S., et al., (2020). Spinal subpial delivery of AAV9 enables widespread gene silencing and blocks motoneuron degeneration in ALS. *Nature Med.*, **26**, 118–130.
139. Naldini, L., (2015). Gene therapy returns to centre stage. *Nature*, **526**, 351–360.
140. LeWitt, P.A., Rezai, A.R., Leehey, M.A., Ojemann, S.G., Flaherty, A.W., Eskandar, E.N., et al., (2011). AAV2-GAD gene therapy for advanced Parkinson's disease: a double-blind, sham-surgery controlled, randomised trial. *Lancet Neurol.*, **10**, 309–319.
141. Mendell, J.R., Al-Zaidy, S., Shell, R., Arnold, W.D., Rodino-Klapac, L.R., Prior, T.W., et al., (2017). Single-dose gene-replacement therapy for spinal muscular atrophy. *New Engl. J. Med.*, **377**, 1713–1722.
142. Warren Olanow, C., Bartus, R.T., Baumann, T.L., Factor, S., Boulis, N., Stacy, M., et al., (2015). Gene delivery of neurturin to putamen and substantia nigra in Parkinson disease: a double-blind, randomized, controlled trial. *Ann. Neurol.*, **78**, 248–257.
143. Kuzmin, D.A., Shutova, M.V., Johnston, N.R., Smith, O. P., Fedorin, V.V., Kukushkin, Y.S., et al., (2021). The clinical landscape for AAV gene therapies. *Nature Rev. Drug Discovery*, **20**, 173–174.
144. Wang, D., Tai, P.W.L., Gao, G., (2019). Adeno-associated virus vector as a platform for gene therapy delivery. *Nature Rev. Drug Discovery*, **18**, 358–378.
145. Mendell, J.R., Al-Zaidy, S.A., Rodino-Klapac, L.R., Goodspeed, K., Gray, S.J., Kay, C.N., et al., (2021). Current clinical applications of in vivo gene therapy with AAVs. *Mol. Ther.*, **29**, 464–488.

146. Finkel, R.S., Fischbeck, K.H., (2021). Maybe too much of a good thing in gene therapy. *Nature Neurosci.*, **24**, 901–902.
147. Van Alstyne, M., Tattoli, I., Delestree, N., Recinos, Y., Workman, E., Shihabuddin, L.S., et al., (2021). Gain of toxic function by long-term AAV9-mediated SMN overexpression in the sensorimotor circuit. *Nature Neurosci.*, **24**, 930–940.
148. Kormann, M.S., Hasenpusch, G., Aneja, M.K., Nica, G., Flemmer, A.W., Herber-Jonat, S., et al., (2011). Expression of therapeutic proteins after delivery of chemically modified mRNA in mice. *Nature Biotechnol.*, **29**, 154–157.
149. Pardi, N., Weissman, D., (2020). Development of vaccines and antivirals for combating viral pandemics. *Nature Biomed. Eng.*, **4**, 1128–1133.
150. Han, Z., Chen, C., Christiansen, A., Ji, S., Lin, Q., Anumonwo, C., et al., (2020). Antisense oligonucleotides increase Scn1a expression and reduce seizures and SUDEP incidence in a mouse model of Dravet syndrome. *Sci. Transl. Med.*, **12**.
151. Lim, K.H., Han, Z., Jeon, H.Y., Kach, J., Jing, E., Weyn-Vanhentenryck, S., et al., (2020). Antisense oligonucleotide modulation of non-productive alternative splicing upregulates gene expression. *Nature Commun.*, **11**, 3501.
152. Roberts, T.C., Langer, R., Wood, M.J.A., (2020). Advances in oligonucleotide drug delivery. *Nature Rev. Drug Discovery*, **19**, 673–694.
153. Zhao, X., Reebye, V., Hitchen, P., Fan, J., Jiang, H., Saetrom, P., et al., (2019). Mechanisms involved in the activation of C/EBPalpha by small activating RNA in hepatocellular carcinoma. *Oncogene*, **38**, 3446–3457.
154. Berchtold, N.C., Cribbs, D.H., Coleman, P.D., Rogers, J., Head, E., Kim, R., et al., (2008). Gene expression changes in the course of normal brain aging are sexually dimorphic. *Proc. Natl. Acad. Sci. USA*, **105**, 15605–15610.
155. Vogler, T.O., Wheeler, J.R., Nguyen, E.D., Hughes, M.P., Britson, K.A., Lester, E., et al., (2018). TDP-43 and RNA form amyloid-like myo-granules in regenerating muscle. *Nature*, **563**, 508–513.
156. Yasuda, K., Clatterbuck-Soper, S.F., Jackrel, M.E., Shorter, J., Mili, S., (2017). FUS inclusions disrupt RNA localization by sequestering kinesin-1 and inhibiting microtubule detyrosination. *J. Cell Biol.*, **216**, 1015–1034.
157. Yasuda, K., Zhang, H., Loiselle, D., Haystead, T., Macara, I.G., Mili, S., (2013). The RNA-binding protein Fus directs translation of localized mRNAs in APC-RNP granules. *J. Cell Biol.*, **203**, 737–746.