The background of the slide is a grayscale electron micrograph showing a synaptic cleft. It features various cellular structures such as mitochondria, endoplasmic reticulum, and synaptic vesicles. The text is overlaid on this image.

Mapping the Proteome of the Synaptic Cleft Through Proximity Labeling

Thomas Biederer

**Department of Neuroscience
Tufts University School of Medicine
Boston, Massachusetts**

Molecular organization of the excitatory synaptic cleft

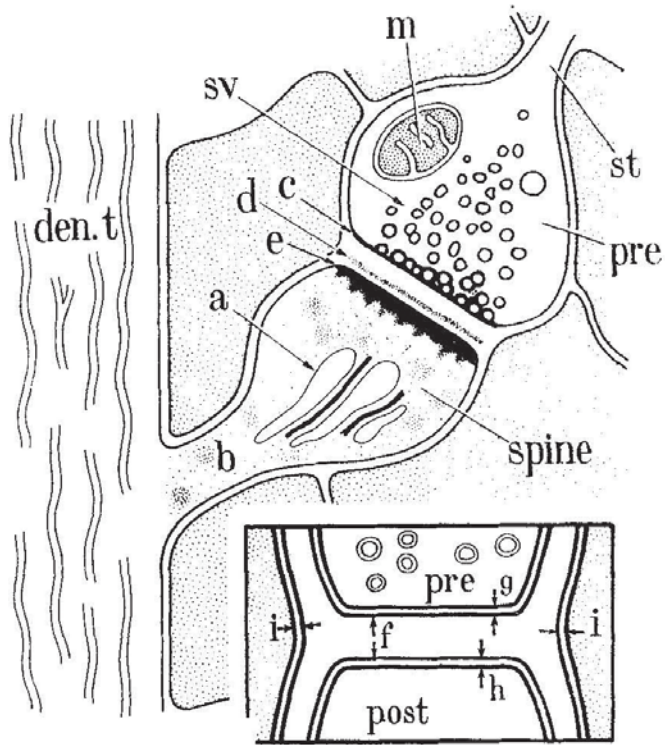
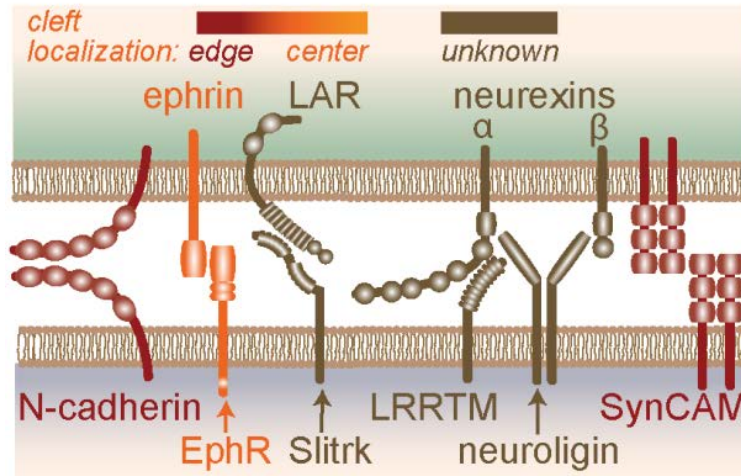
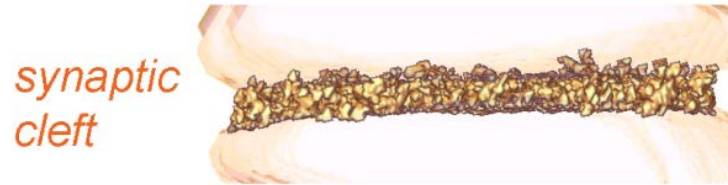


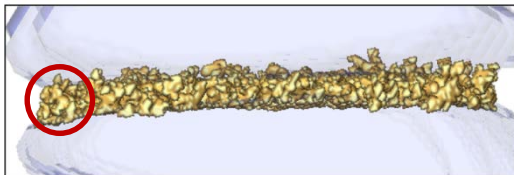
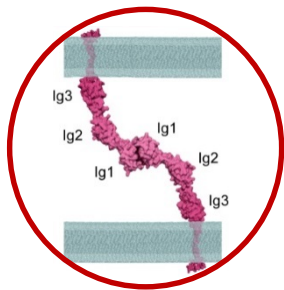
Fig. 1. Diagram of a synaptic contact on a dendritic spine, observed with the electron microscope after osmium tetroxide fixation. The stippled regions represent neuronal and glial processes of the neuropil

Inset. The opposed regions of the pre- and post-synaptic membranes seen after potassium permanganate fixation. The membranes (i) are of neighbouring processes of the neuropil



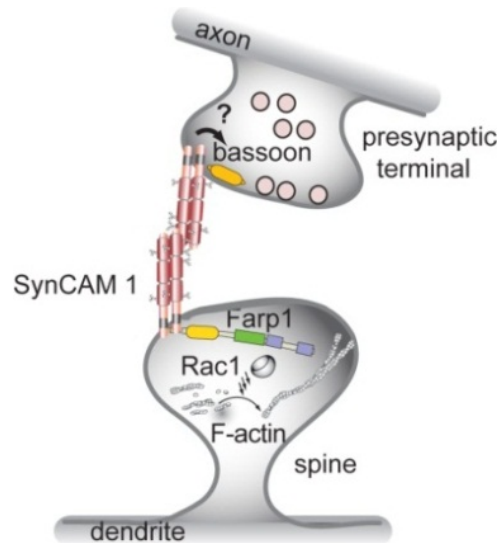
Our approaches to investigate synapse development

I. Molecular properties of synaptic adhesion



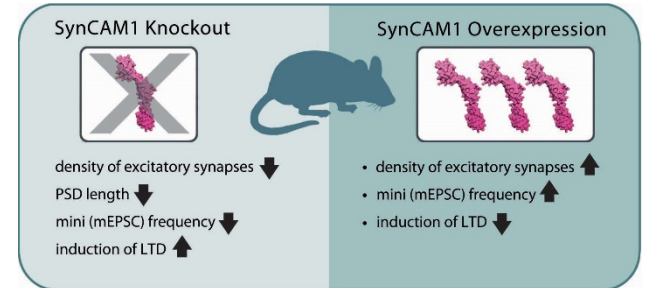
Biederer, *Genomics* 2006
 Fogel et al. *J Neurosci* 2007
 Thomas et al. *J Comp Neurol* 2008
Fogel et al. *J Biol Chem* 2010
 Fogel et al. *EMBO J* 2011
 Perez de Arce et al. *Neuron* 2015
Cijsouw et al. *Proteomes* 2018

II. Synaptogenic signaling

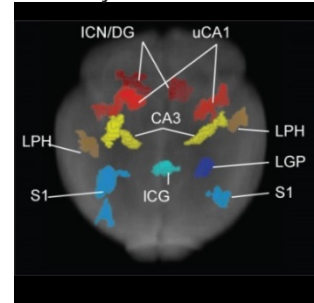


Stagi et al. *PNAS* 2010
Cheadle & Biederer *J Cell Biol* 2012
 Cheadle & Biederer *J Neurosci* 2014

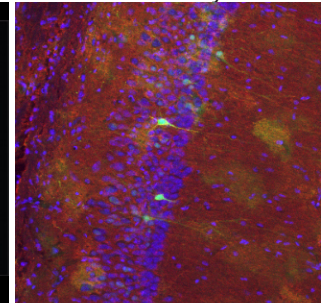
III. Wiring neurons into circuits



DTI SynCAM1 KO



CA1 c-fos OE SynCAM1^{flag}

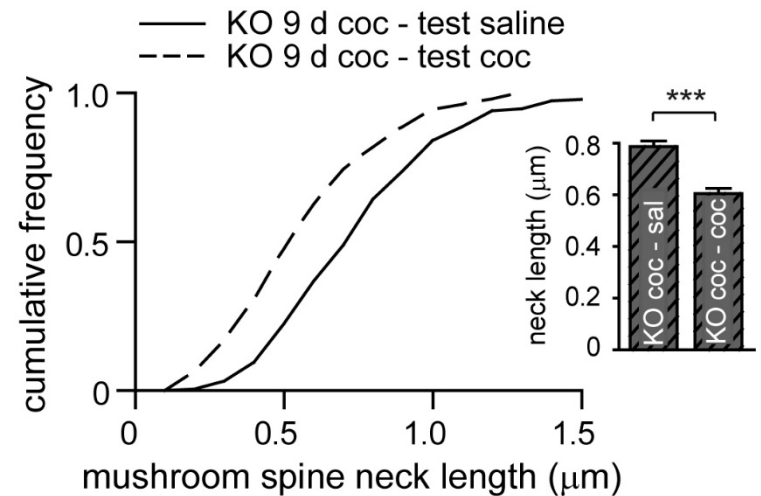
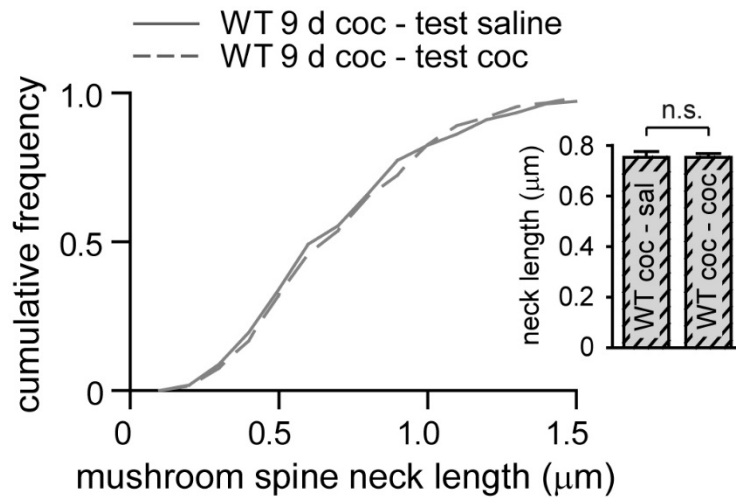
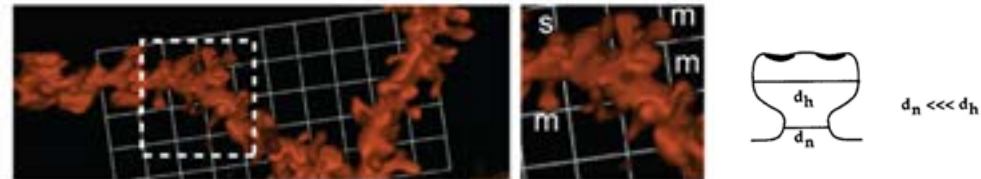
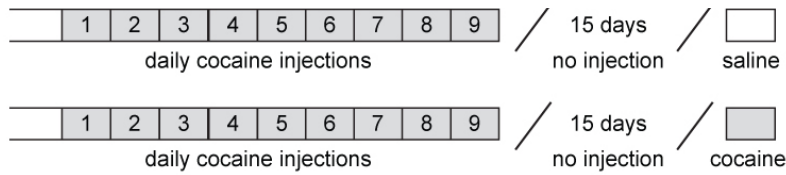


flag / SynCAM 1 / NeuN

Robbins et al. *Neuron* 2010
 Giza et al. *Neuropsychopharm* 2013
 Ribic et al. *J Comp Neurol* 2014
 Park et al. *J Neurosci* 2016
 Ribic, Crair and Biederer *Cell Reports* 2019

Trans-synaptic interactions impact synapse structure effects of psychostimulants

Loss of SynCAM 1 sensitizes in NAc medium spiny neurons the mushroom spines to cocaine-induced shortening:



Unraveling the complexity and heterogeneity of synaptic composition

Resource



Proteomic screening of glutamatergic mouse brain synaptosomes isolated by fluorescence activated sorting

Christoph Biesemann¹, Mads Grønberg^{2,10,†}, Elisa Luquet^{3,4}, Sven P Wichert^{5,‡}, Véronique Bernard^{6,7,8}, Simon R Bungers¹, Ben Cooper¹, Frédérique Varoqueaux¹, Liyi Li¹, Jennifer A Byrne⁹, Henning Urlaub^{10,11}, Olaf Jahn¹², Nils Brose^{1,7} & Etienne Herzog^{1,3,4,6,7,8,**}



Resource

A multiregional proteomic survey of the postnatal human brain

Becky C. Carlyle, Robert R. Kitchen, Jean E. Kanyo, Edward Z. Voss, Mihovil Pletikos, André M. M. Sousa, TuKiet T. Lam, Mark B. Gerstein, Nenad Sestan & Angus C. Nairn

OPEN ACCESS Freely available online

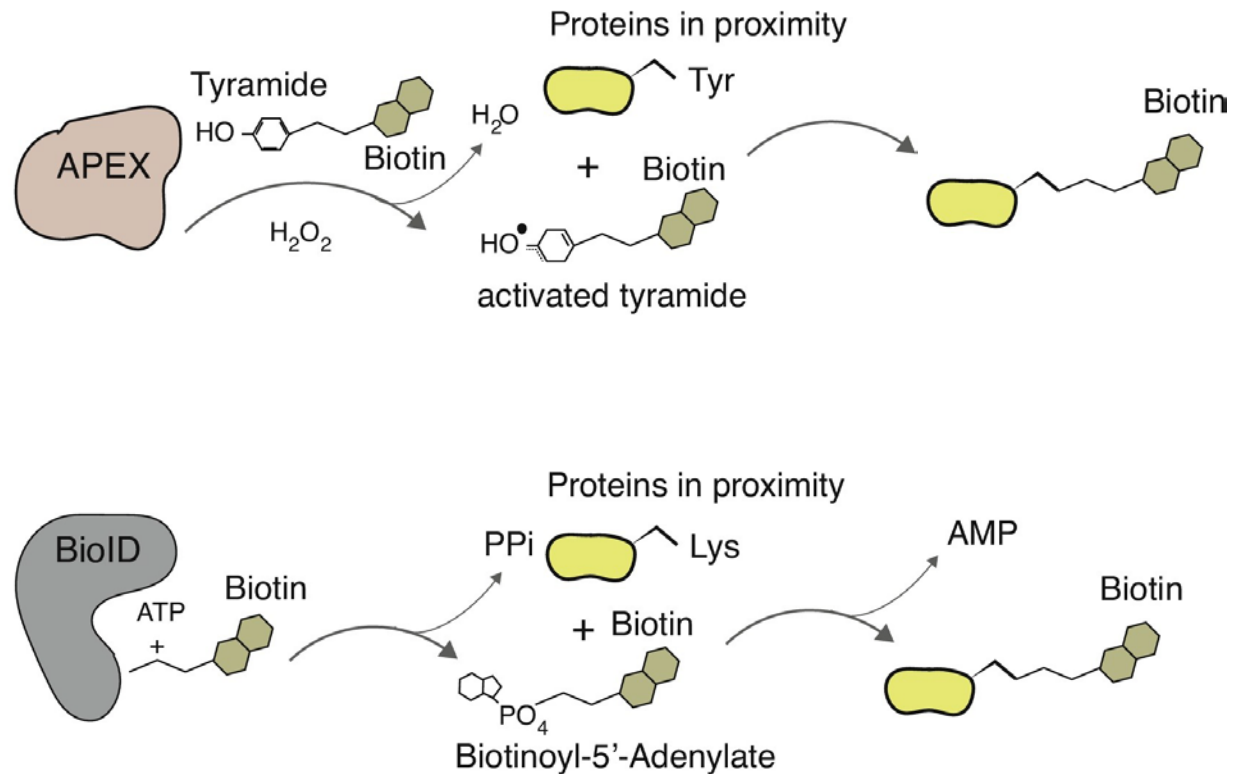


The Biochemical Anatomy of Cortical Inhibitory Synapses

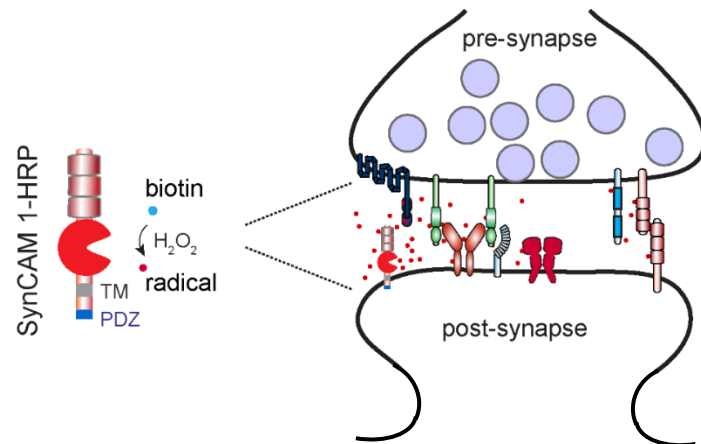
Elizabeth A. Heller¹, Wenzhu Zhang², Fekrije Selimi³, John C. Earnheart¹, Marta A. Šlimak⁴, Julio Santos-Torres⁴, Ines Ibañez-Tallon⁴, Chiye Aoki⁵, Brian T. Chait², Nathaniel Heintz^{1*}

¹ Howard Hughes Medical Institute, Laboratory of Molecular Biology, The Rockefeller University, New York, New York, United States of America, ² Laboratory for Mass Spectrometry and Gaseous Ion Chemistry, The Rockefeller University, New York, New York, United States of America, ³ CIRB, Collège de France, Paris, France, ⁴ Molecular Neurobiology Group, Max-Delbrück-Center for Molecular Medicine, Berlin, Germany, ⁵ Center for Neural Science, New York University, New York, New York, United States of America

Proximity labeling: a method to tag proximal proteins with biotin



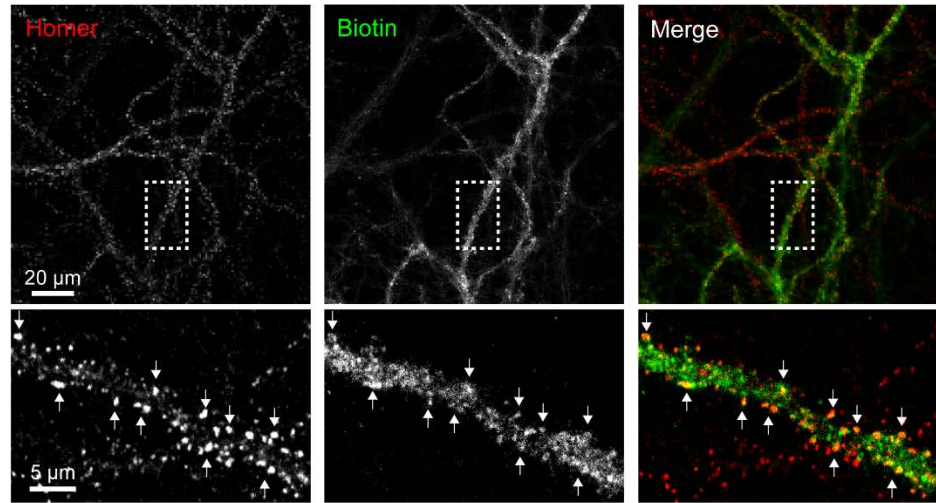
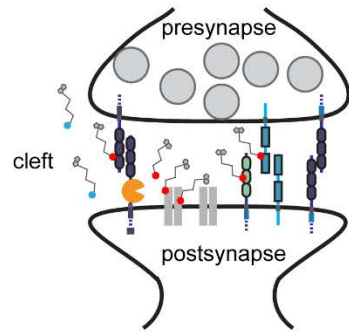
A SynCAM 1-HRP reporter to target the excitatory synaptic cleft



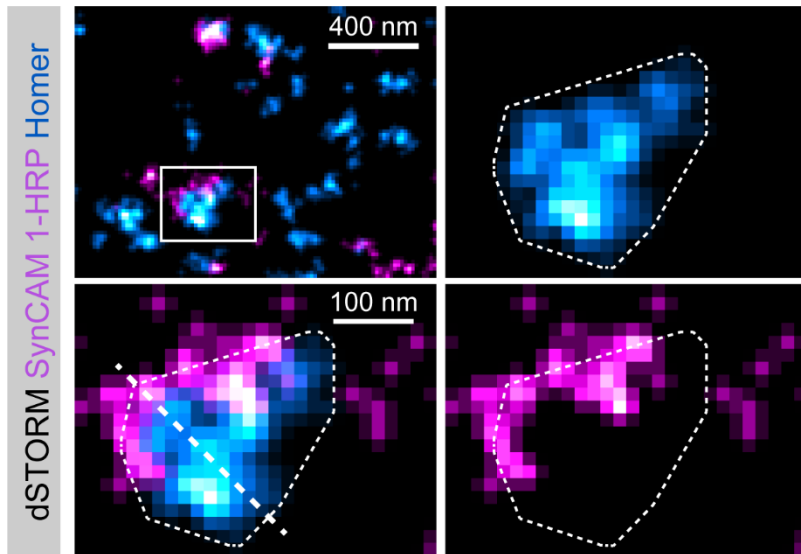
Based on proximity labelling approach developed by Alice Ting and colleagues.

Synaptic protein labeling by the SynCAM1-HRP reporter vs dendritic control

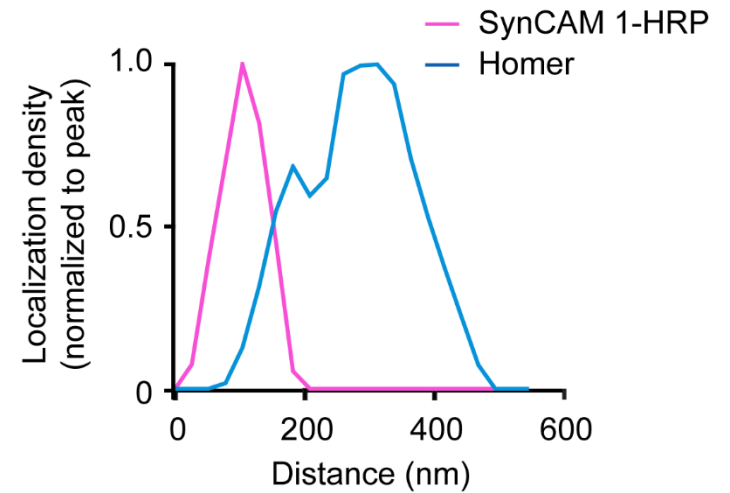
SynCAM 1-HRP



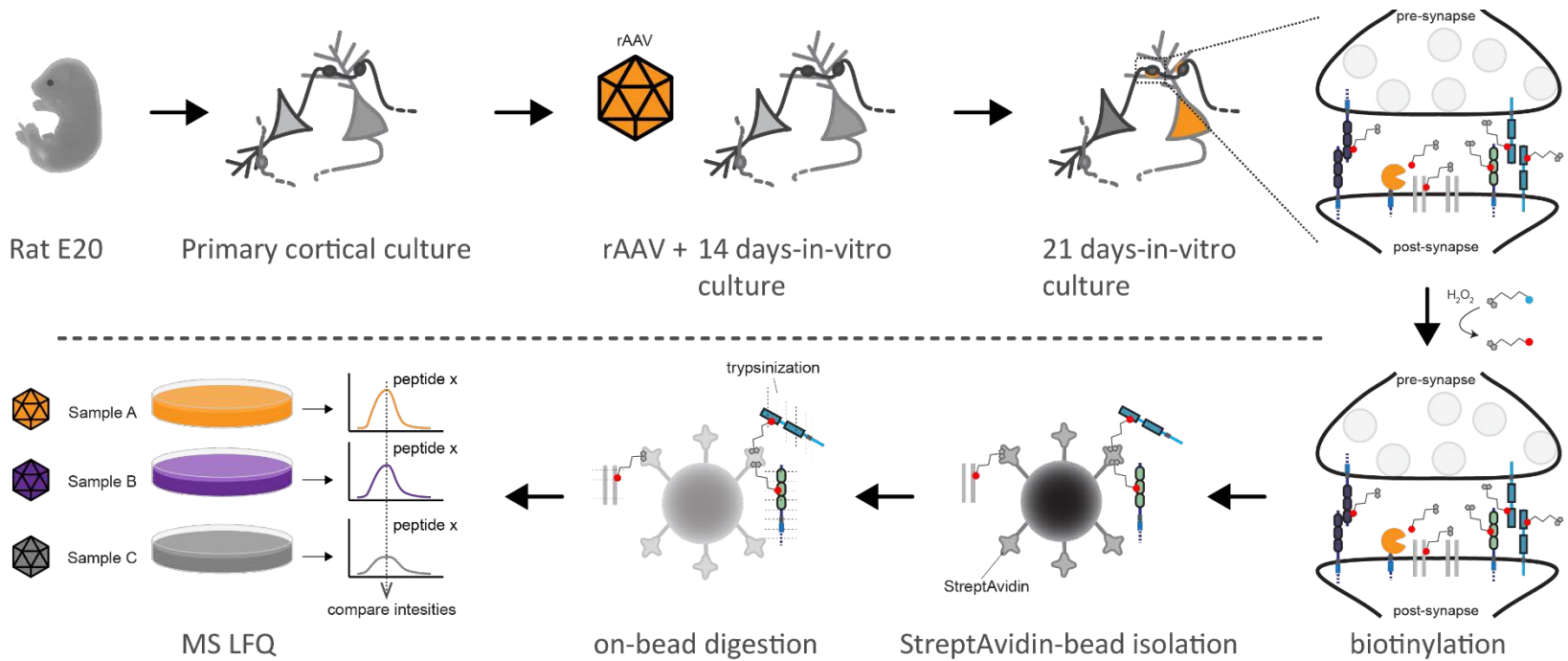
Sub-synaptic localization of the SynCAM 1-HRP reporter at the synaptic edge



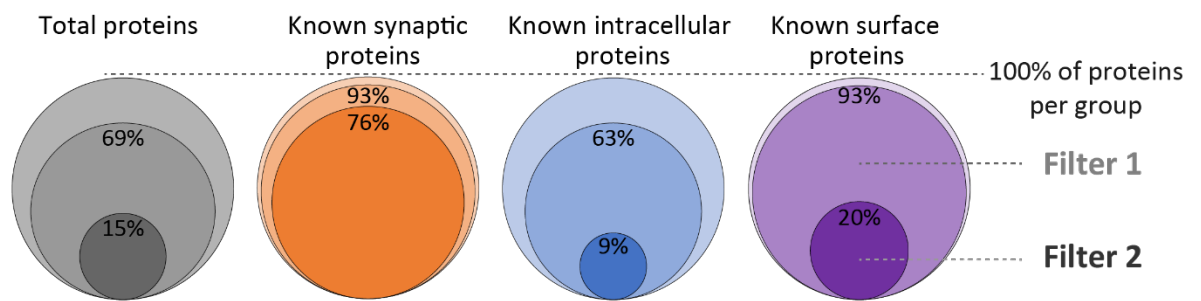
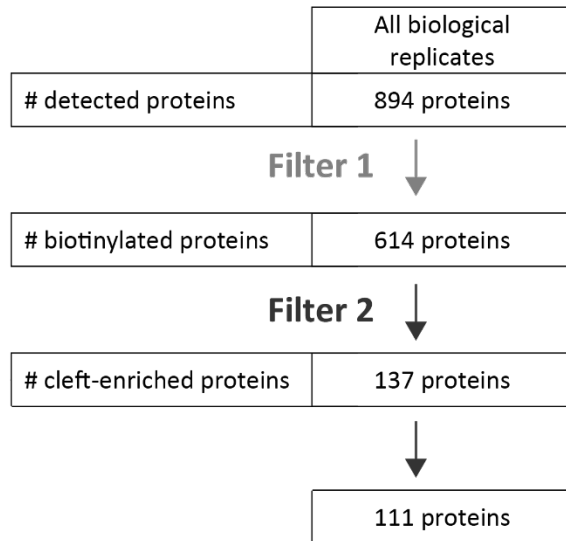
3D dSTORM imaging



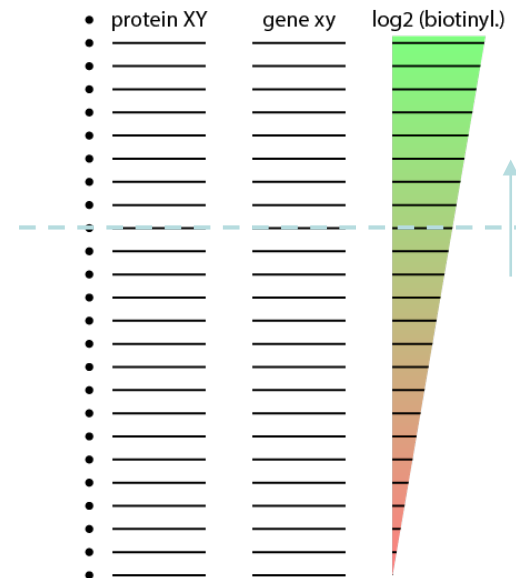
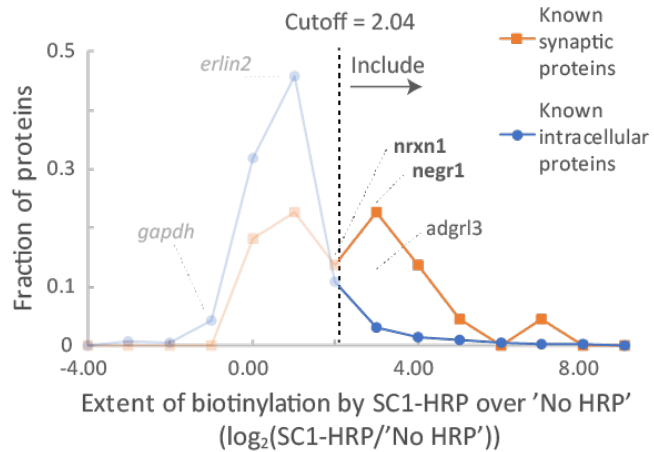
Experimental design and workflow



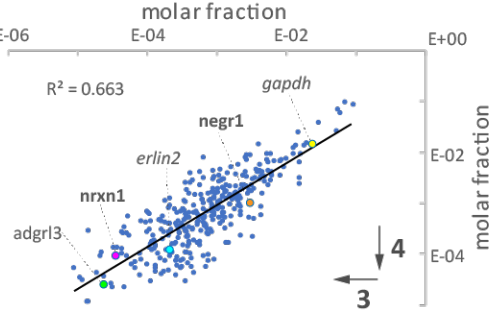
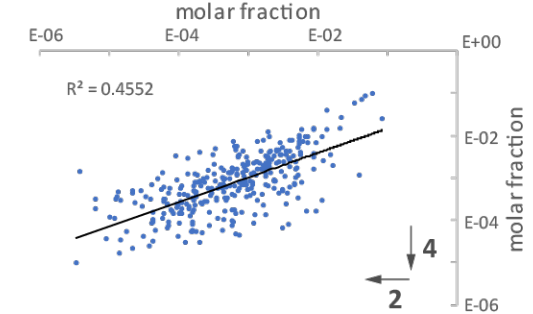
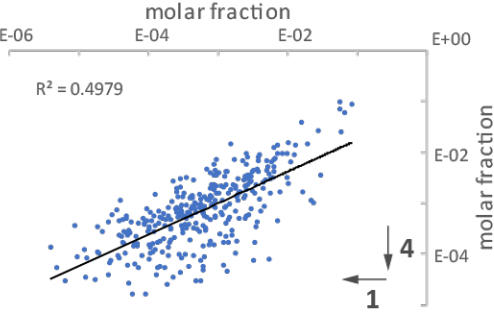
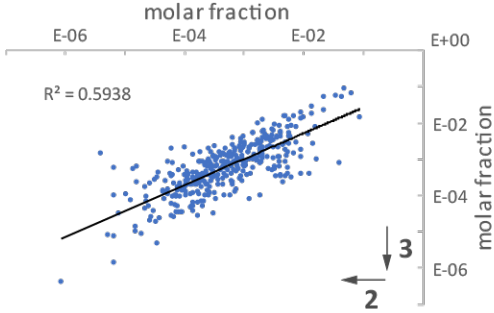
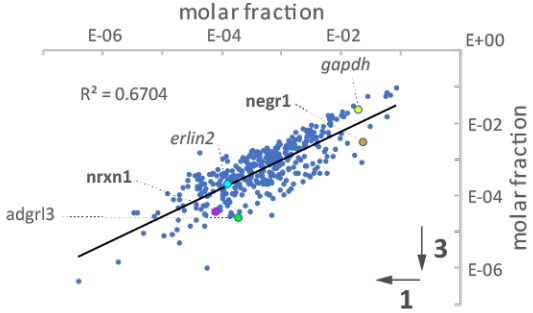
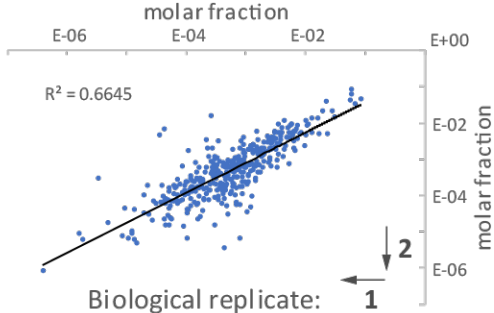
Data analysis steps for selection of putative cleft proteins



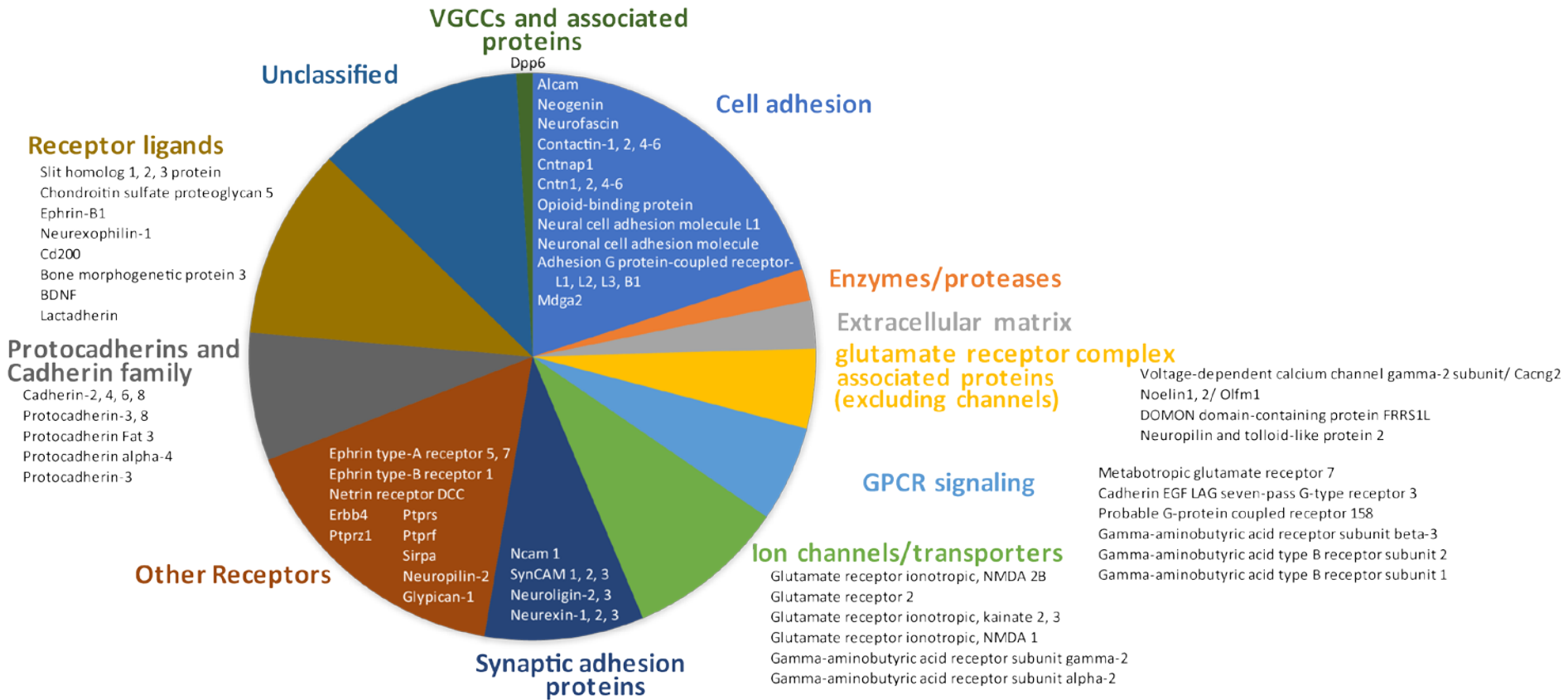
A 'true positive – false positive' filter for synaptic cleft proteins



Identified proteins correlate well across biological replicates



Synaptic cleft candidates are found across protein classes

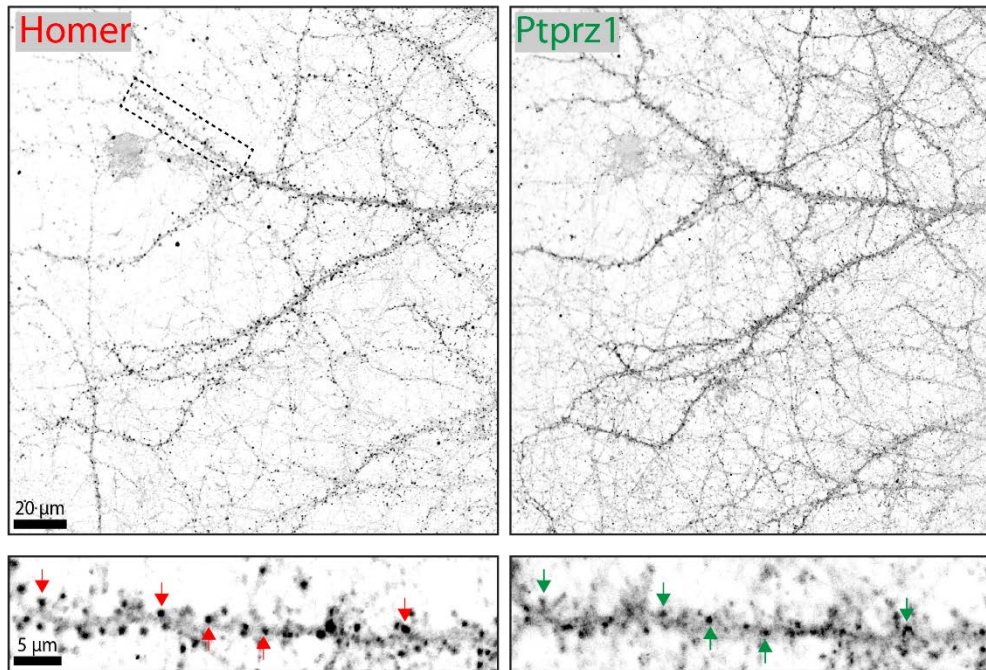


Validation: The tyrosine phosphatase Ptpnz1 colocalizes with excitatory synapses

Protein-Tyrosine Phosphatase, Receptor-type, Zeta-1:



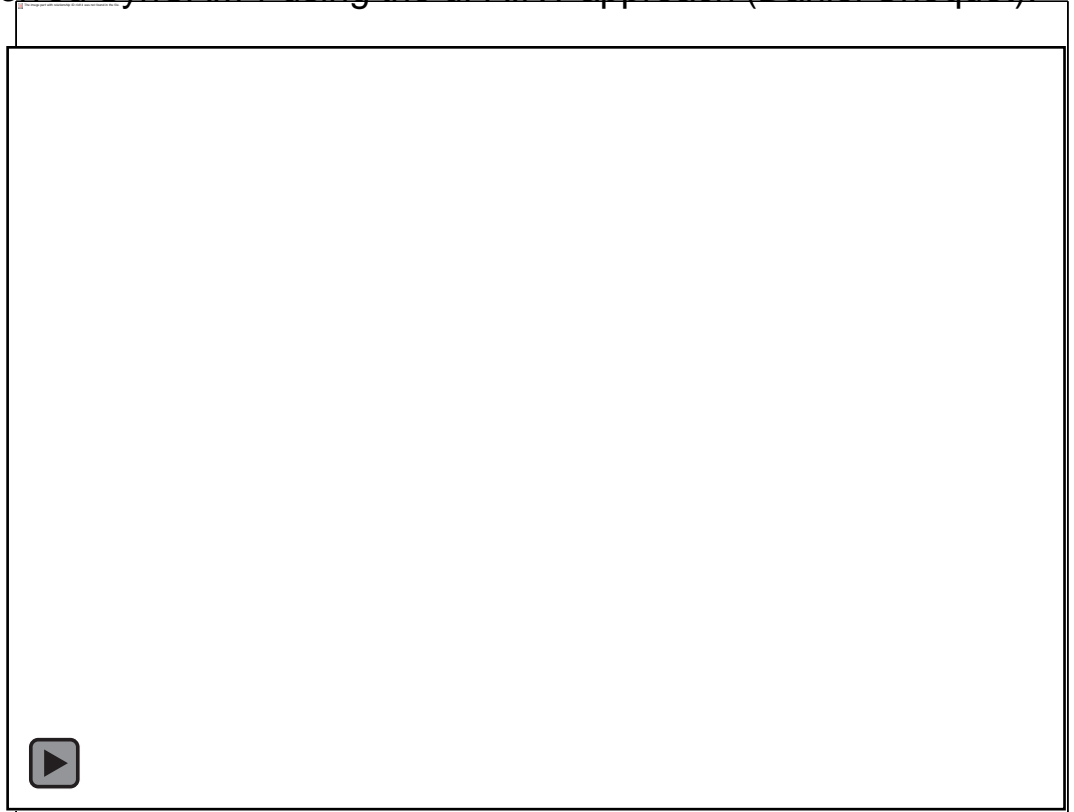
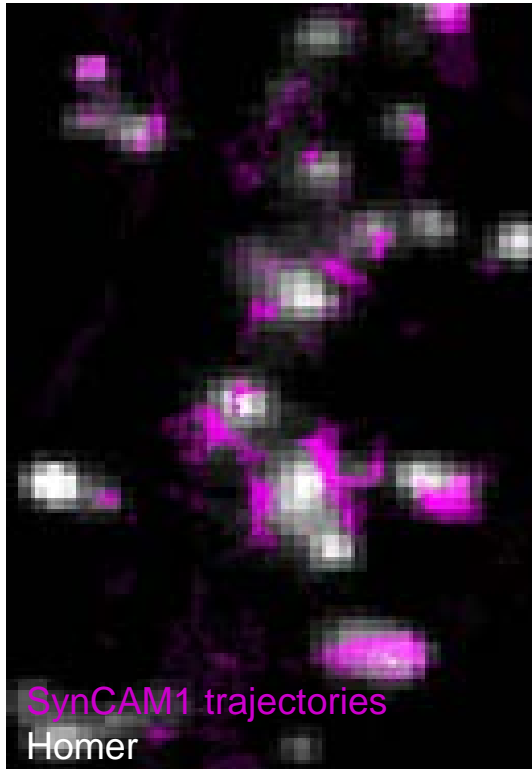
Ptpnz1 expression is regulated by morphine in rodents (Garcia-Perez et al. (2017) *Mol Neurobiol* 54:495-510)



Detected using immunocytochemistry

The next step: Defining the dynamic activity-dependent remodeling of the synaptic cleft

Single particle tracking of super-resolved SynCAM 1 using the uPAINT approach (Daniel Choquet):



SynCAM 1-pHluorin with single molecule nanobody labeling
20 ms per frame

Acknowledgements

Adema Ribic, Ph.D.

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University of Maryland
Tom Blanpied



Yale/NIDA Neuroproteomics Center

TuKiet Lam



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NIH/NINDS R21 NS10963

NIH/NIDA P30 DA018343



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