

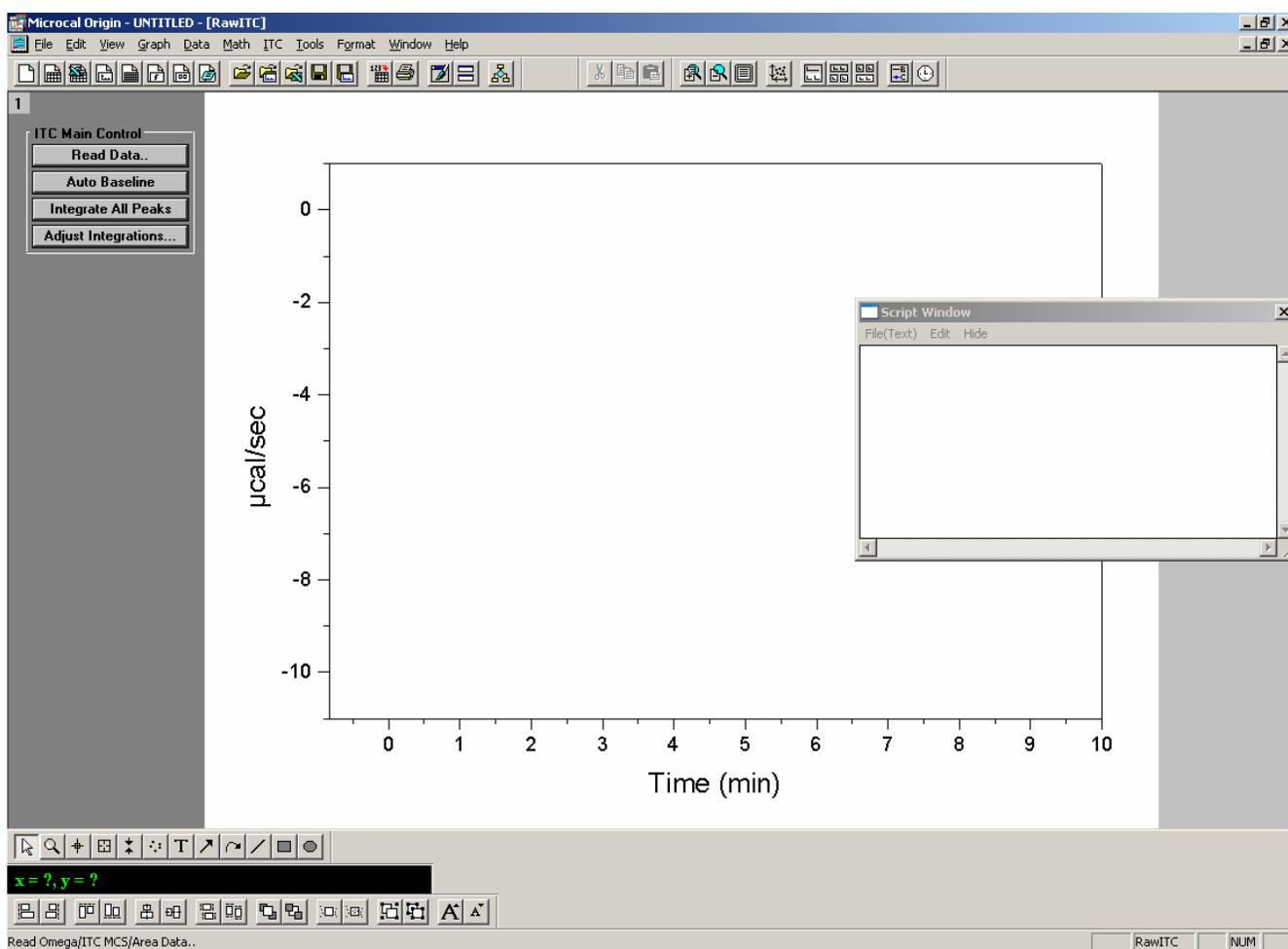
## Processing ITC data using ORIGIN 5.0

1. Launch the MicroCal processing software (Origin 5.0 with Microcal ITC

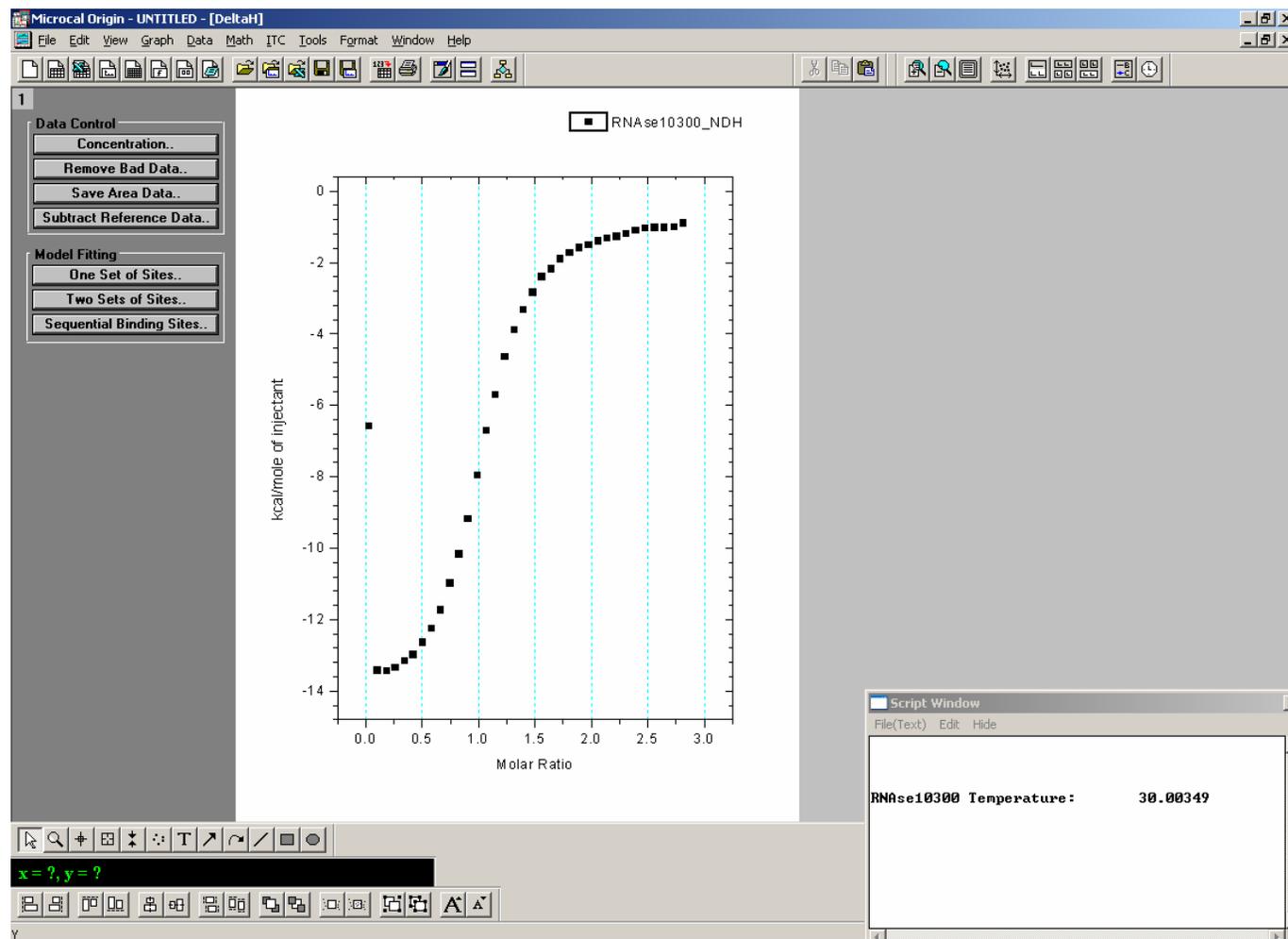


feature) Microcal Inc. ITC.Ink icon on a desktop.

2. The following window appears:



3. Click “Read Data” and open the data file : “\*.itc” in this case “RNAse\*\*\*\*.itc”
4. Once the data file is loaded, the following window appears:

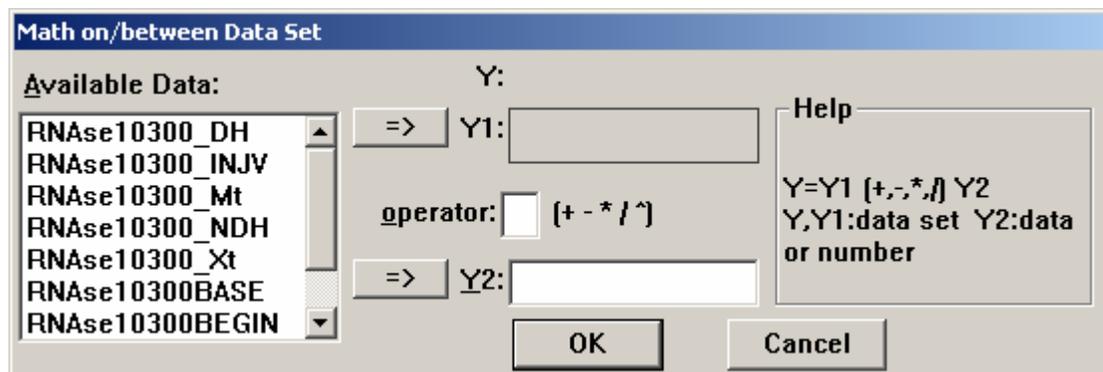


5. At this point the data can be corrected for heat of dilution of the ligand (which are the heats observed at the end of titration, where no binding events are happening and majority of heat observed are from dilution of titrant).
6. Go to “Window” (on the main tab) “Name of the data file” selection (in this case “window” → “RNAse103007”)
7. The following window appears:

The screenshot shows the Microcal Origin software interface. The main window displays a table with the following columns: DH (Y), INJV (Y), Xi (Y), Mt (Y), XMt (X), and NDH (Y). The data rows are numbered 1 through 36. A 'Script Window' is open in the bottom right corner, displaying the text: 'RNase10300 Temperature: 30.00349'.

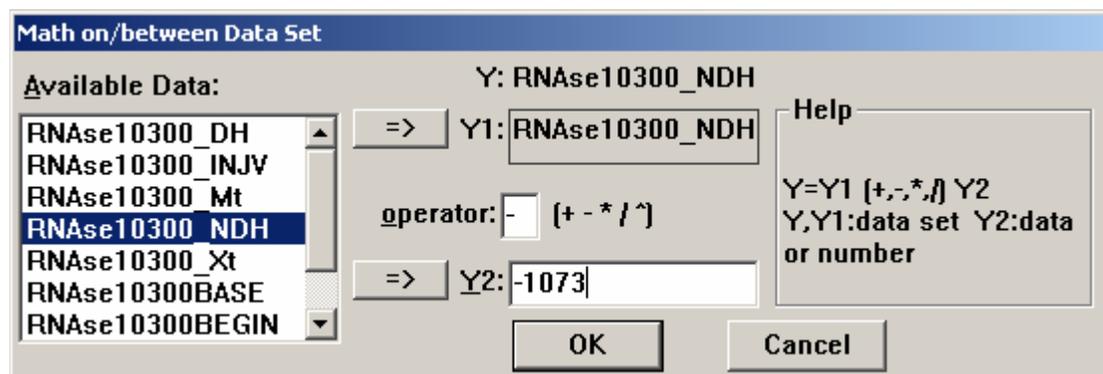
	DH (Y)	INJV (Y)	Xi (Y)	Mt (Y)	XMt (X)	NDH (Y)
1	-12.49578	1	0	0.0504	0.02643	-6576.72582
2	-76.5581	3	0.00133	0.05036	0.10584	-13431.24642
3	-76.67572	3	0.00532	0.05026	0.18542	-13451.88099
4	-76.03736	3	0.0093	0.05015	0.26516	-13339.88808
5	-75.05513	3	0.01327	0.05005	0.34507	-13167.567
6	-74.03893	3	0.01723	0.04994	0.42515	-12989.28575
7	-72.08799	3	0.02119	0.04984	0.50539	-12647.01591
8	-69.89562	3	0.02513	0.04973	0.5858	-12262.38961
9	-66.87644	3	0.02907	0.04963	0.66638	-11732.70942
10	-62.58374	3	0.033	0.04952	0.74712	-10979.60306
11	-57.9531	3	0.03692	0.04942	0.82803	-10167.20969
12	-52.3641	3	0.04084	0.04932	0.9091	-9186.68393
13	-45.3917	3	0.04474	0.04921	0.99035	-7963.4559
14	-38.27674	3	0.04864	0.04911	1.07176	-6715.21693
15	-32.49515	3	0.05252	0.04901	1.15333	-5700.9037
16	-26.51929	3	0.0564	0.0489	1.23508	-4652.50678
17	-22.25841	3	0.06027	0.0488	1.31699	-3904.98409
18	-18.94247	3	0.06413	0.0487	1.39906	-3323.24007
19	-16.17843	3	0.06799	0.0486	1.48131	-2838.32117
20	-13.72115	3	0.07183	0.04849	1.56372	-2407.21972
21	-12.47567	3	0.07567	0.04839	1.64629	-2188.7138
22	-10.91302	3	0.0795	0.04829	1.72903	-1914.56508
23	-9.88032	3	0.08332	0.04819	1.81194	-1733.38907
24	-9.08917	3	0.08713	0.04809	1.89502	-1594.59175
25	-8.59761	3	0.09094	0.04799	1.97826	-1508.35218
26	-8.0037	3	0.09473	0.04789	2.06167	-1404.15791
27	-7.57929	3	0.09852	0.04778	2.14525	-1329.70062
28	-7.27702	3	0.1023	0.04768	2.22899	-1276.6695
29	-6.82534	3	0.10606	0.04758	2.3129	-1197.42795
30	-6.35986	3	0.10983	0.04748	2.39698	-1115.76492
31	-5.97998	3	0.11358	0.04738	2.48122	-1049.11853
32	-5.86642	3	0.11732	0.04728	2.56563	-1029.19613
33	-5.87551	3	0.12106	0.04719	2.65021	-1030.79105
34	-5.78283	3	0.12479	0.04709	2.73495	-1014.53233
35	-5.15273	3	0.12851	0.04699	2.81986	-903.98838
36		-	0.13222	0.04689	-	-

8. Scroll down to the last injections and compute an average value for NDH for the last 5-15 injections; make sure these injections are really of similar heats and represent pure dilution effects; for the RNase103007 file, this value is  $\sim -1073$ .
9. Go to “Math” → “Simple Math” on the main tab.
10. The following window appears:



11. Select “RNase\*\*\*\_NDH” press “=>” button and populated the Y1 window

12. Choose the “operator” as “-” and populate the “Y2” window with the average heat of dilution computed in step 8 (for RNase10300, the value is “-1073”).

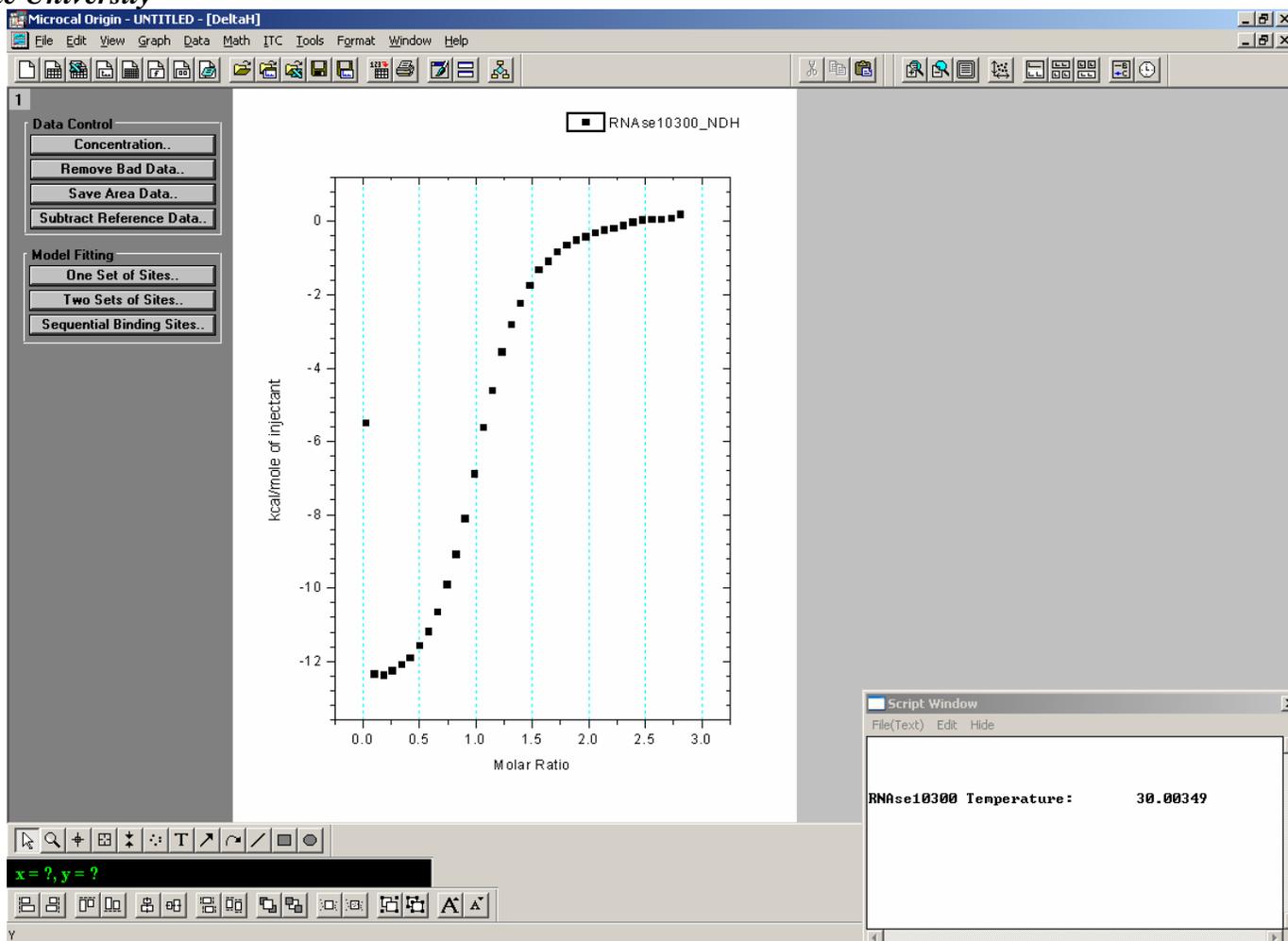


13. Click “OK” to subtract the heats of dilution from all measured heats.

14. Change display window to “DeltaH” (“window” → “DeltaH”).

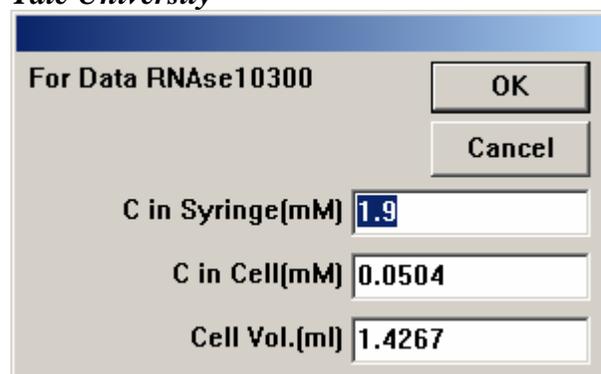
the heats should be corrected for heat of dilution as illustrated:

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15. You may correct the concentrations at this point if needed, but choosing “Concentration” and changing the values for cell and ligand.

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For Data RNAse10300

OK

Cancel

C in Syringe(mM) 1.9

C in Cell(mM) 0.0504

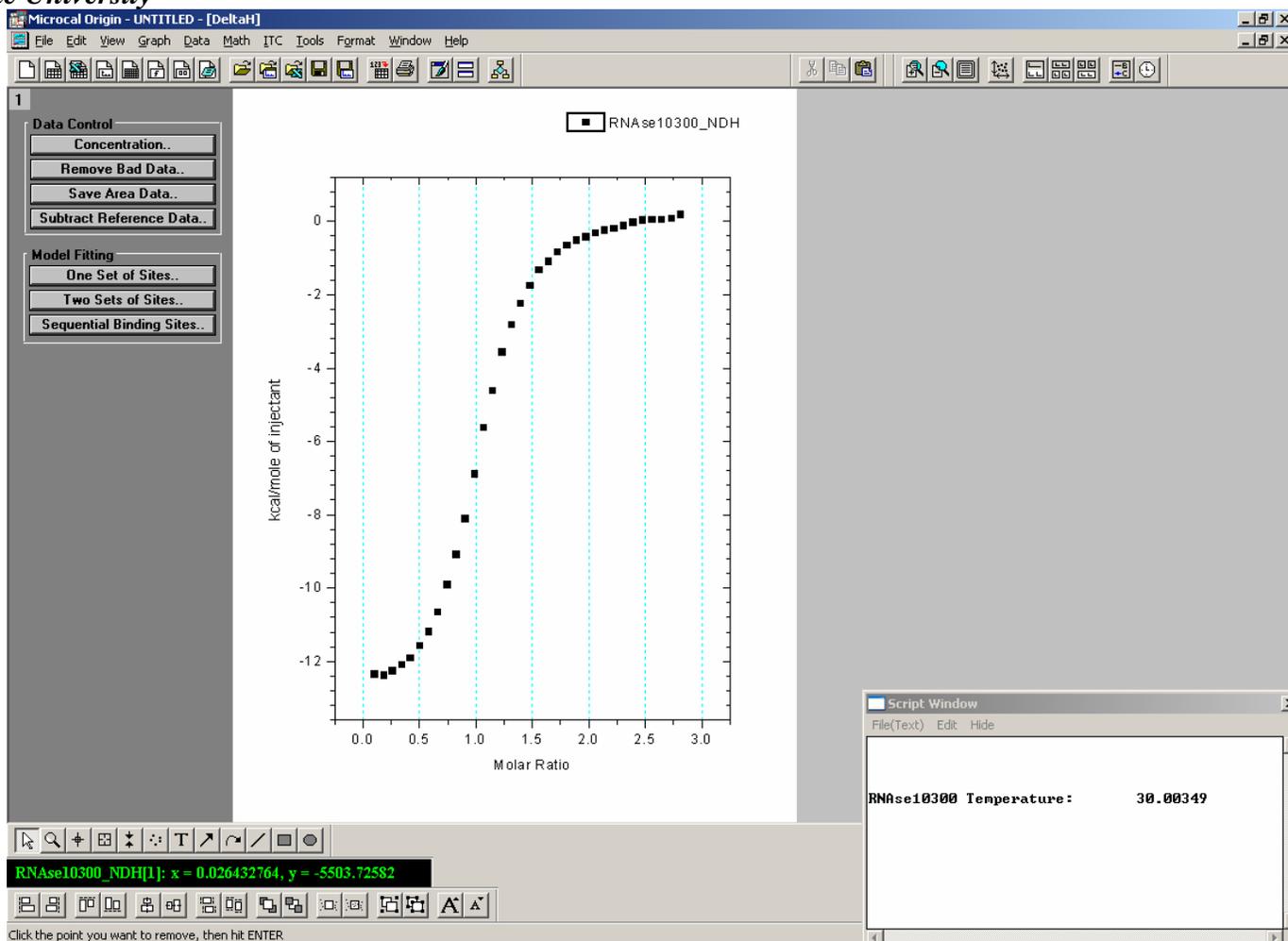
Cell Vol.(ml) 1.4267

16. Remove the results for first injection by choosing “Remove Bad Data” (under “Data Control” section on left hand side); select the data point for the first injection

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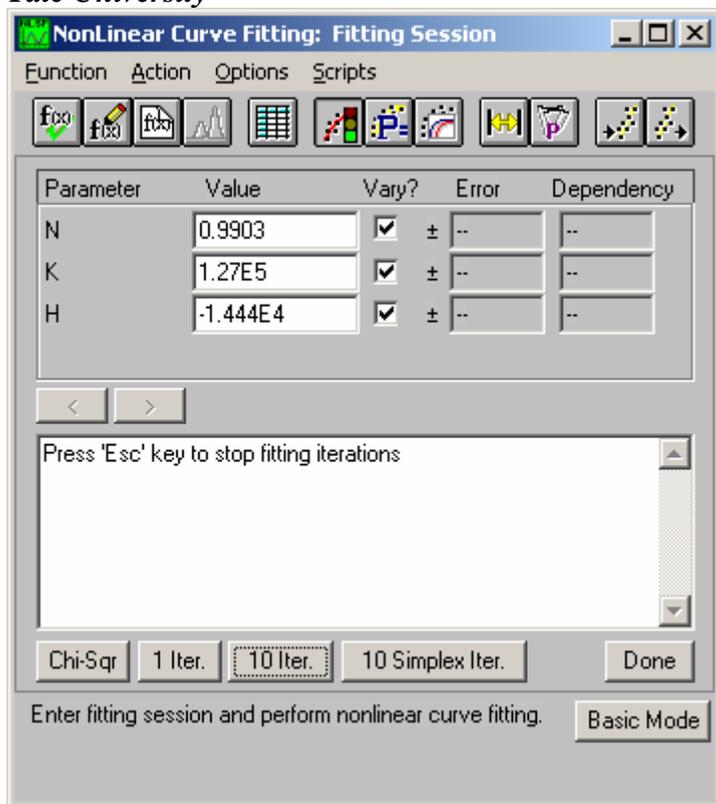
and hit “enter” key, which should delete the result for first injection:

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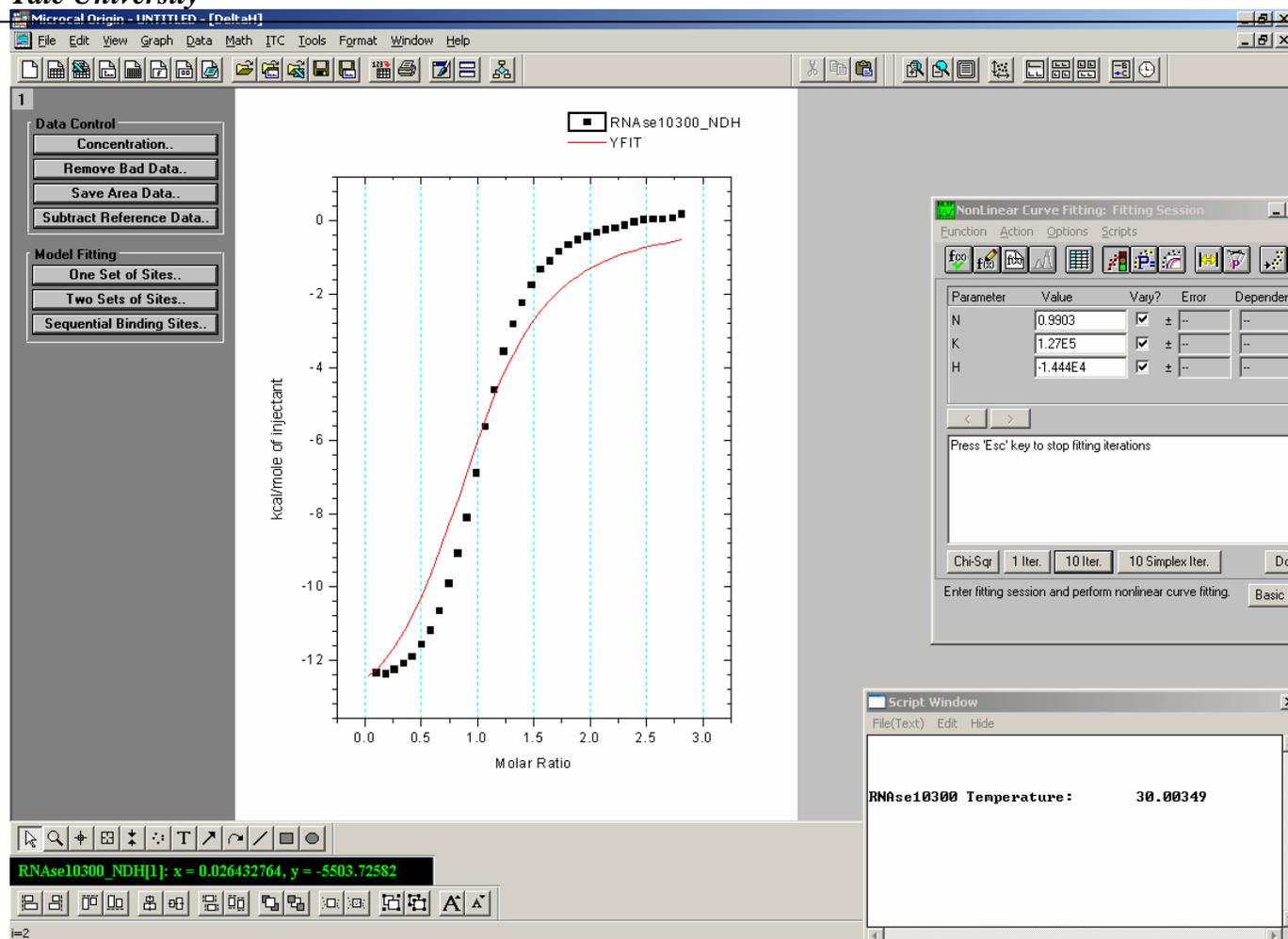
17. Now you are ready to choose the model for fitting the data from the “Model Fitting” section.
18. For single sets of sites: choose “One Set of sites” and the following window appear:

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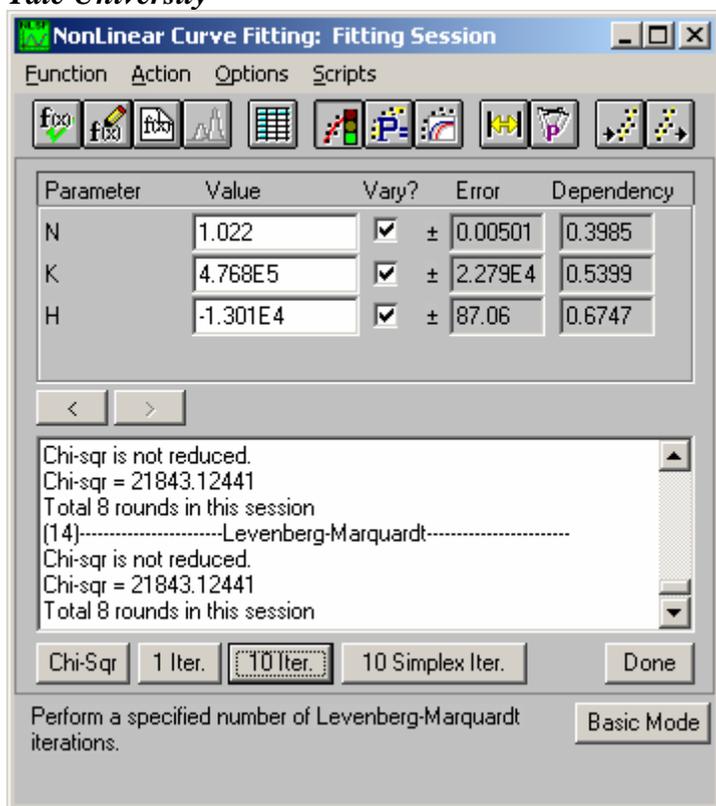
with the initial guesses computed by the software and the red curve for fitted values displayed on the “DeltaH” window:

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19. Choose “1 Iter.” (single iteration and monitor the progression to make sure the fit is converging properly and eventually do “10 Iter.” until the Chi-Sqr value does not change with subsequent iterations.

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20. Choose “Done” and the results will be displayed on the DelatH page:

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