

# Application of label-free technologies in a core facility research

Ewa Folta-Stogniew  
*Yale University*



# Biophysics Resource of Keck Laboratory: Yale School of Medicine

**Mission:** quantitative characterization of interactions between biomolecules using in solution biophysical methods

## Common questions:

- how tight is the binding ? ( binding affinity:  $K_d$ ,  $K_a$ )
- how many of each molecule are in the complex (stoichiometry)
- how fast does the complex form? (kinetics)
- is the binding event enthalpy or entropy-driven? (thermodynamics)

## List of technologies:

- Size Exclusion Chromatography coupled with Light Scattering (SEC/LS)
- Dynamic Light Scattering (DLS)
- Isothermal MicroCalorimeter (ITC)
- CD-Spectrophotometer
- Stopped-Flow Spectrofluorometer
- Surface Plasmon Resonance (SPR) Sensor [BiaCore Biosensor; T100]
- Composition Gradient Static Light Scattering (CGSLS)
- Asymmetric flow Field-Flow Fractionation (AFFF)

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# Application of label-free technologies in a core facility research

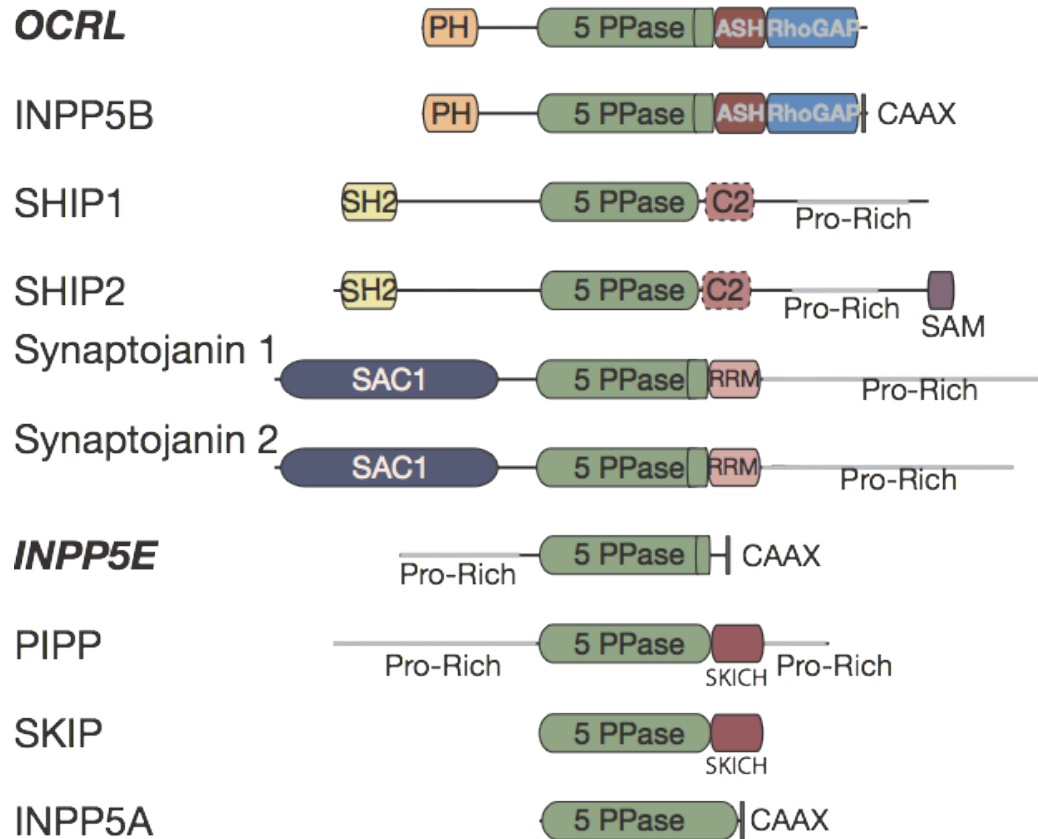
## Recognition of the F&H motif by the Lowe syndrome protein OCRL

Michelle Pirruccello, Laura Swan, Ewa Folta-Stogniew, and Pietro DeCamilli

*Yale University*



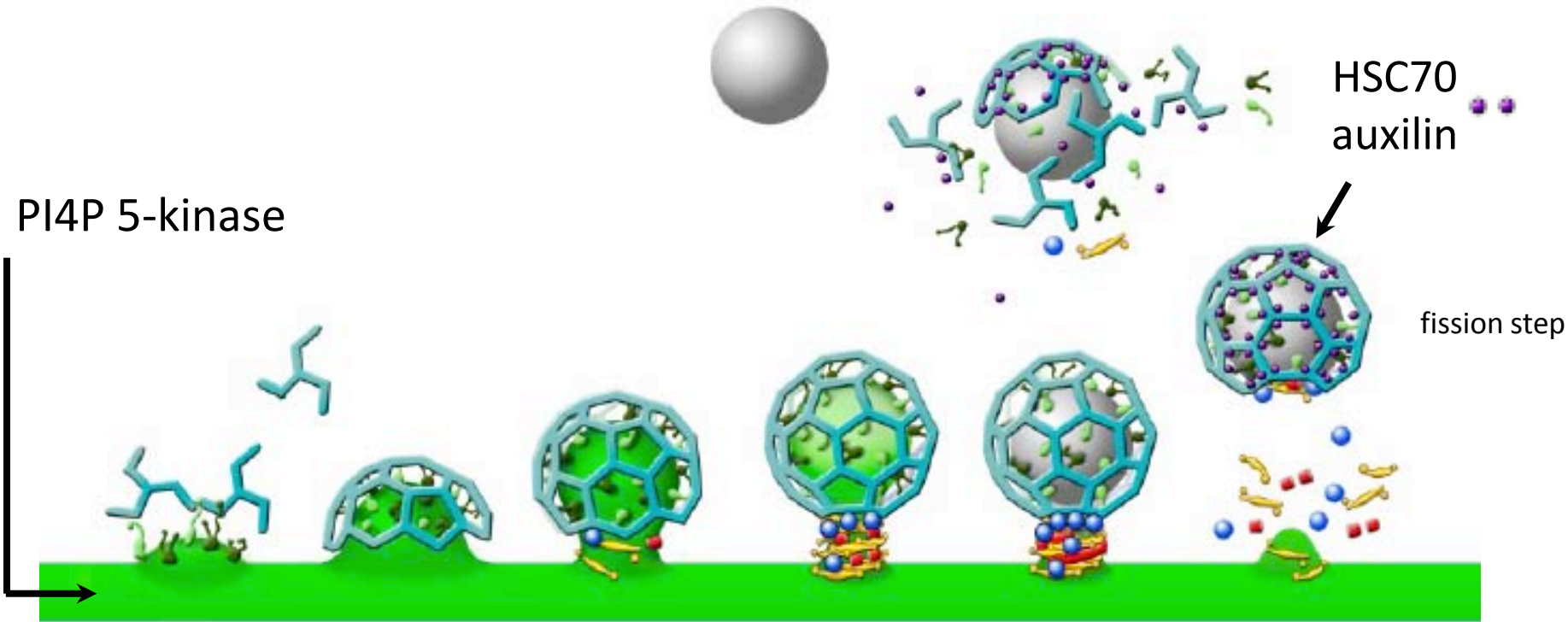
# Phosphoinositide 5-Phosphatases



Common phosphatase domain flanked by regions which direct the enzymes to the correct membrane target

# PI(4,5)P<sub>2</sub> in Clathrin-mediated endocytosis

PI4P 5-kinase



HSC70  
auxilin

fission step

PI(4,5)P<sub>2</sub>

SHIP2

synaptojanin

OCRL

synaptojanin-170



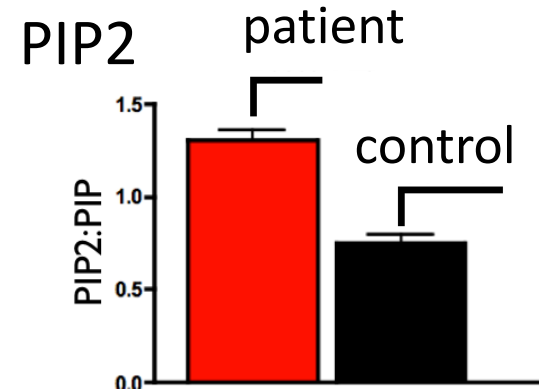
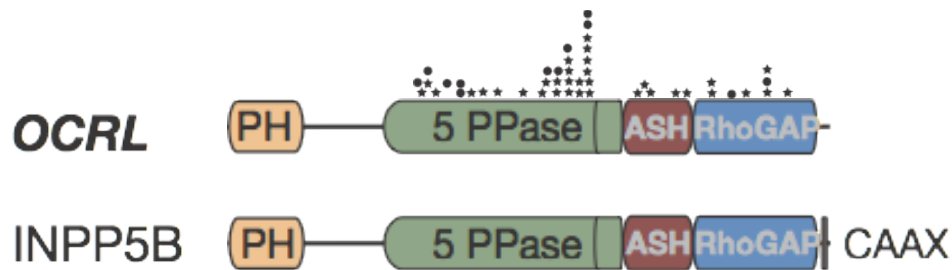
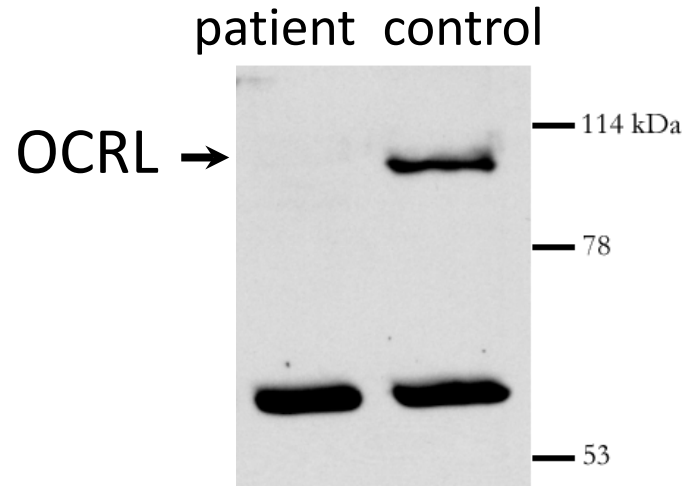
# Loss of function of OCRL: Oculocerebrorenal syndrome of Lowe and Dent's disease (OCRL phosphatase)

## Lowe's syndrome

Kidney Reabsorptive Defects  
 Congenital Cataracts  
 Cognitive Impairment

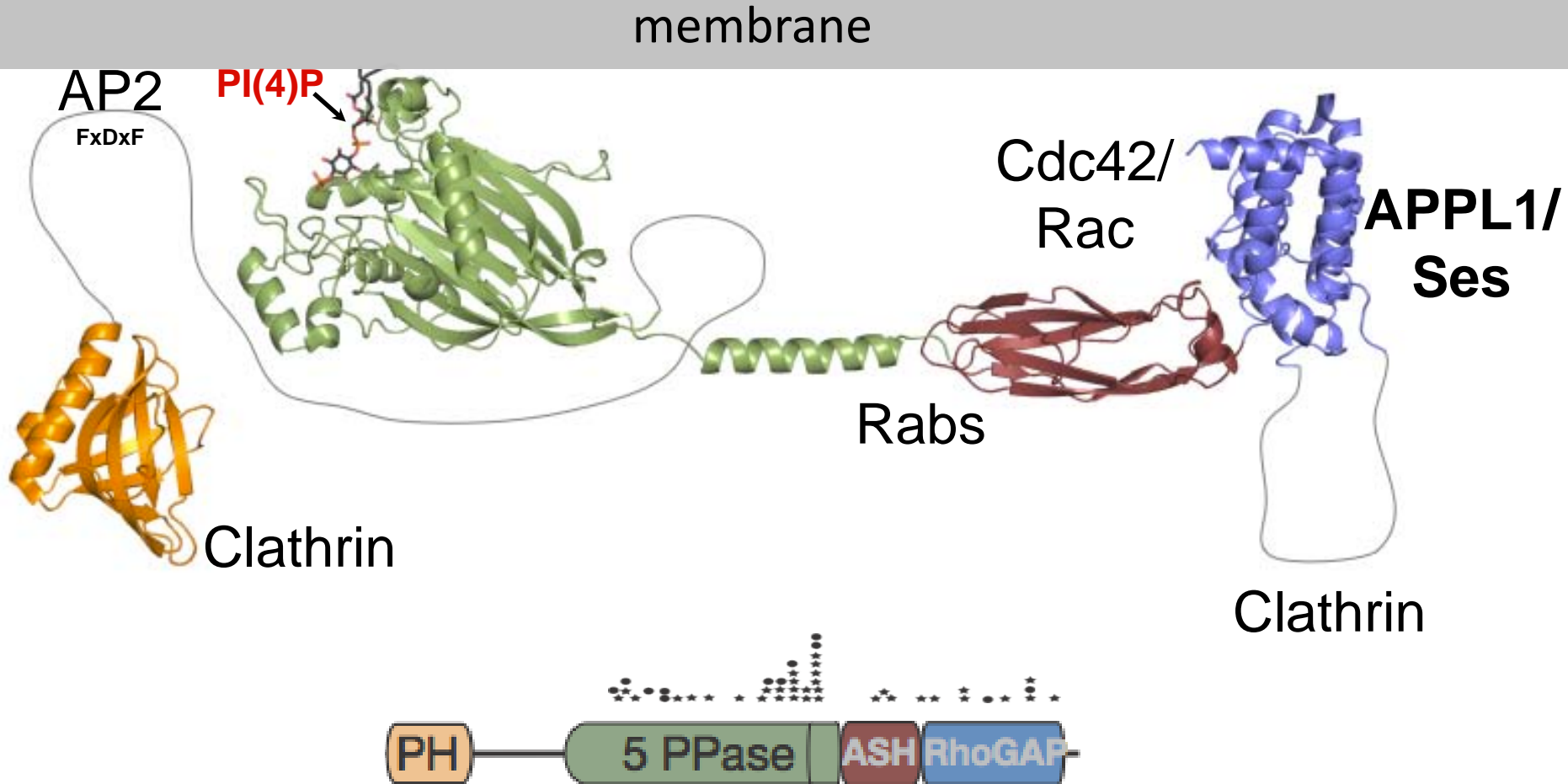
## Dent 2 Disease

Mainly caused by mutations in CIC-5.  
 Some patients identified with OCRL mutations.  
 Kidney Reabsorptive Defects



Disease causing mutations in C-terminus (non-catalytic domain)  
 Increased PIP2 level even for mutations in non-catalytic domains

# OCRL membrane recruitment



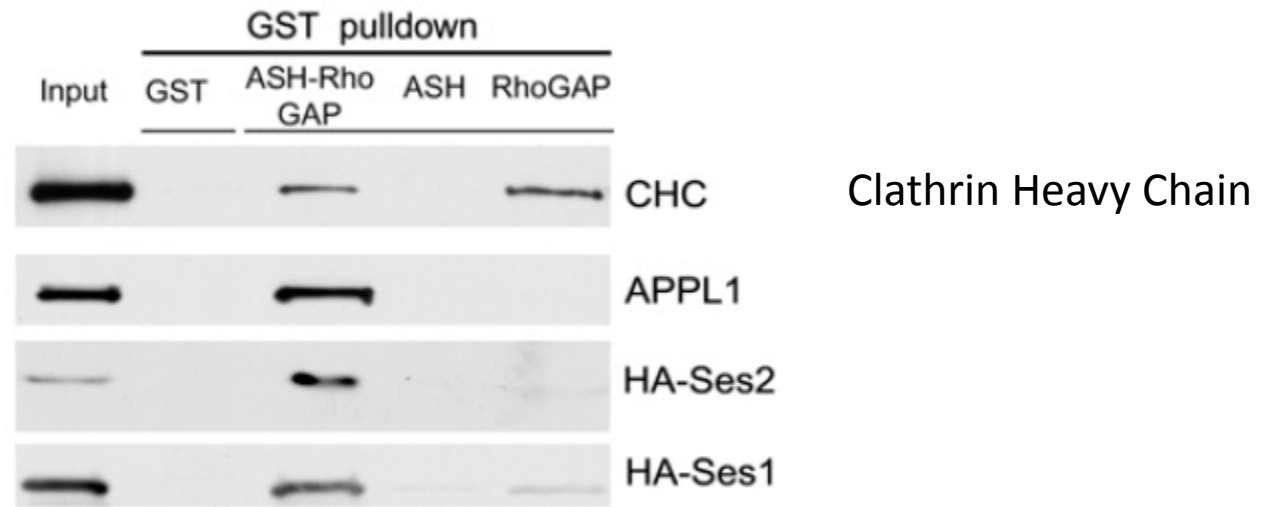
Mao et. al. 2009

3MTC.pdb (Structural Genomics Consortium)

Pirruccello et. al. 2011



# A new class of endocytic adaptors containing an F&H motif



Binding of F&H motif requires the presence of both: the ASH and RhoGAP domains

# Definition of the consensus F&H motif



**Ses1/2**

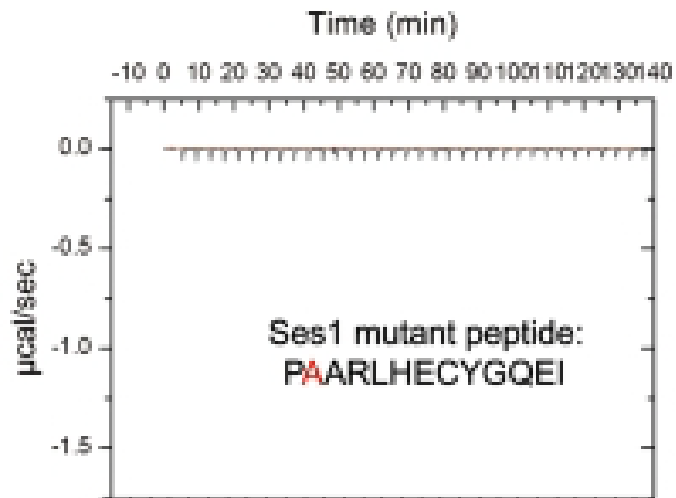
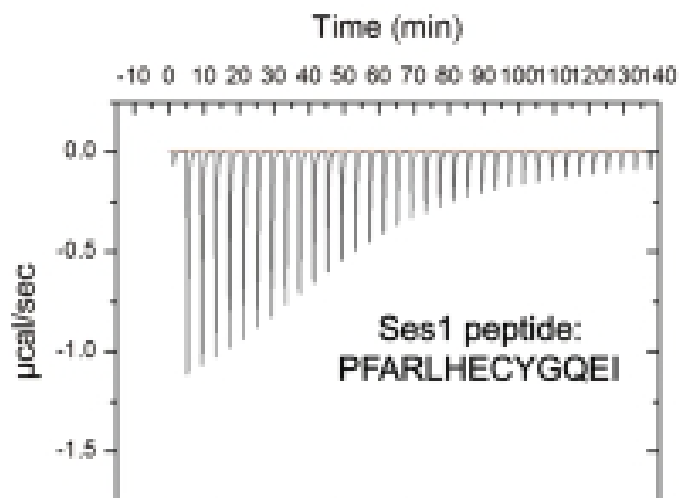


<b>P</b> FAR <b>L</b> HE <b>C</b> Y <b>G</b> Q <b>E</b> I	<b>Ses1</b>
<b>C</b> F <b>S</b> T <b>L</b> H <b>D</b> W <b>Y</b> G <b>Q</b> E <b>I</b>	<b>Ses2</b>
<b>E</b> F <b>A</b> R <b>N</b> H <b>E</b> R <b>F</b> R <b>R</b> E <b>L</b>	<b>DSes</b>
<b>S</b> F <b>Q</b> <b>Q</b> R <b>H</b> E <b>S</b> <b>L</b> R <b>P</b>	<b>APPL1</b>

**OCRL**

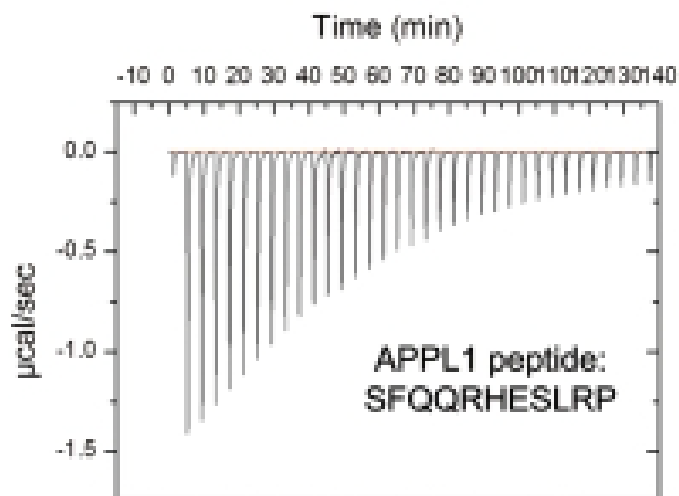


# Direct binding of the minimal consensus peptides to the OCRL ASH-RhoGAP-like domain



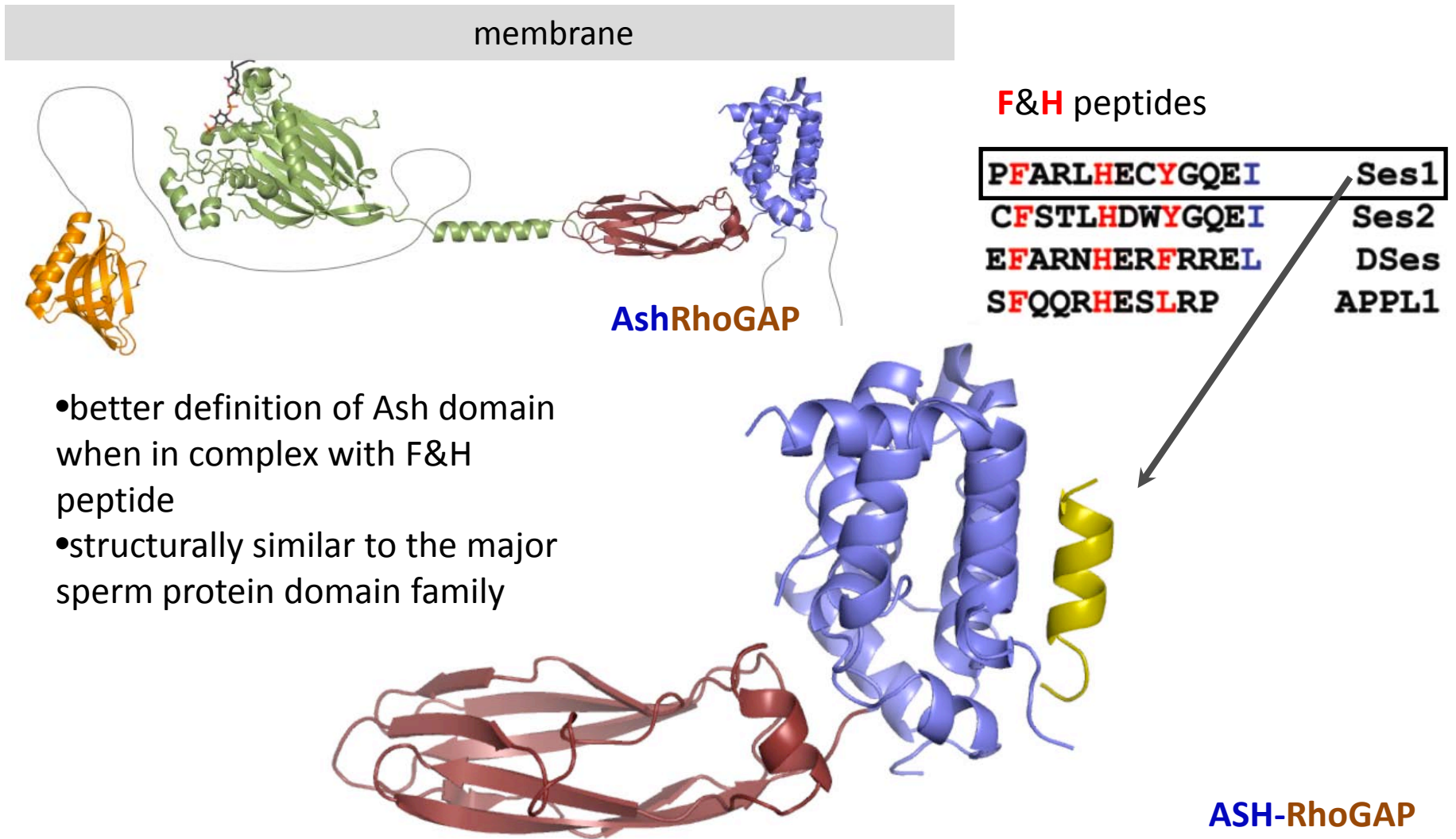
F to A mutant

30 µM OCRL  
2 mM peptide

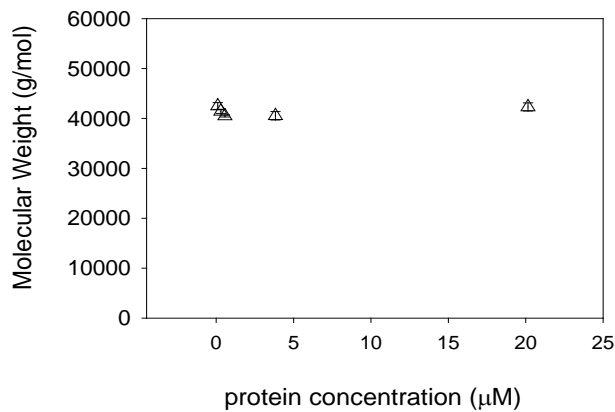
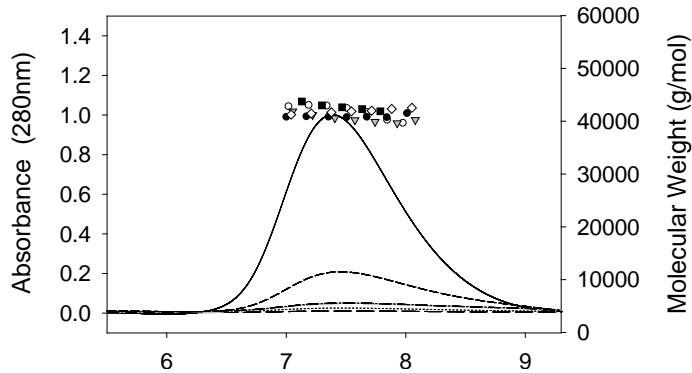


Kd (µM) ITC			
Proteins	F&H Peptides		
	APPL	Ses1	Ses1 (F to A)
Wt OCRL ASH-RhoGAP	12 ± 2	0.7 ± 0.08	NB

# Crystallographic Analysis of the F&H/OCRL interaction

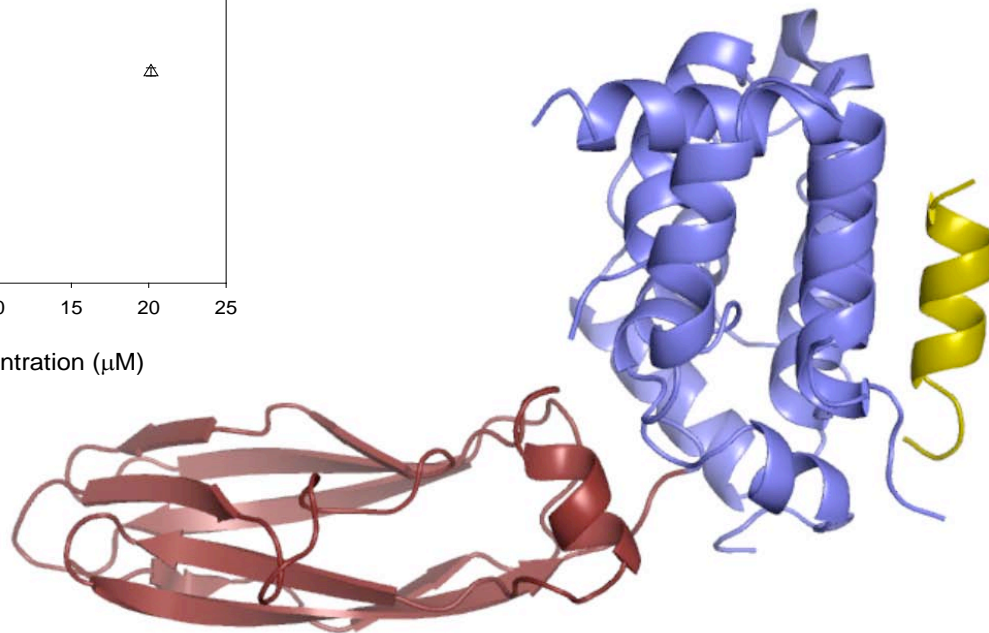


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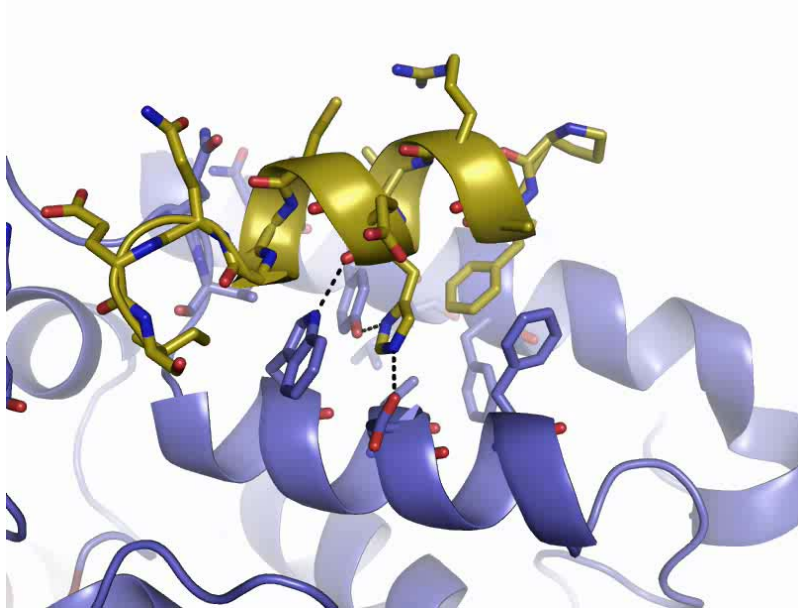
F&H peptides

<b>P</b> FAR <b>L</b> HE <b>C</b> Y <b>G</b> Q <b>E</b> I	<b>Ses1</b>
<b>C</b> F <b>S</b> T <b>L</b> H <b>D</b> W <b>Y</b> G <b>Q</b> E <b>I</b>	<b>Ses2</b>
<b>E</b> F <b>A</b> R <b>N</b> H <b>E</b> R <b>F</b> R <b>R</b> E <b>L</b>	<b>DSes</b>
<b>S</b> F <b>Q</b> Q <b>R</b> H <b>E</b> S <b>L</b> R <b>P</b>	<b>APPL1</b>



ASH-RhoGAP

# Crystallographic Analysis of the F&H/OCRL interaction



## F&H peptide

Phenylalanine  
(F&H motif)

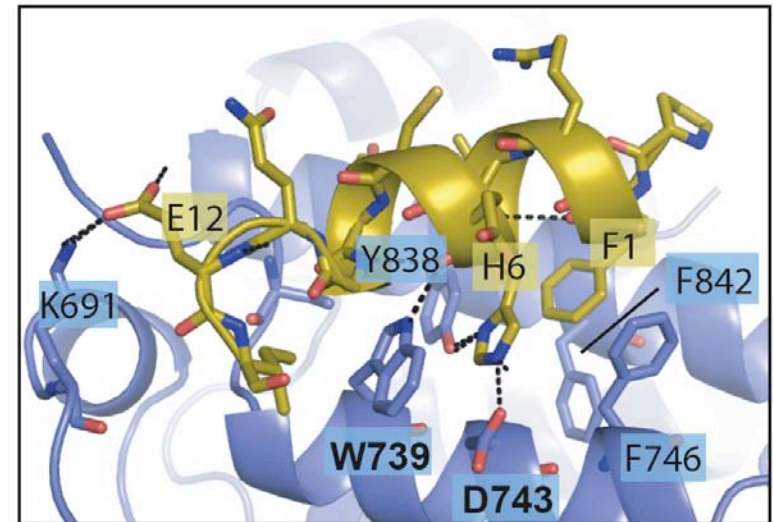
His (F&H motif)

## OCRL

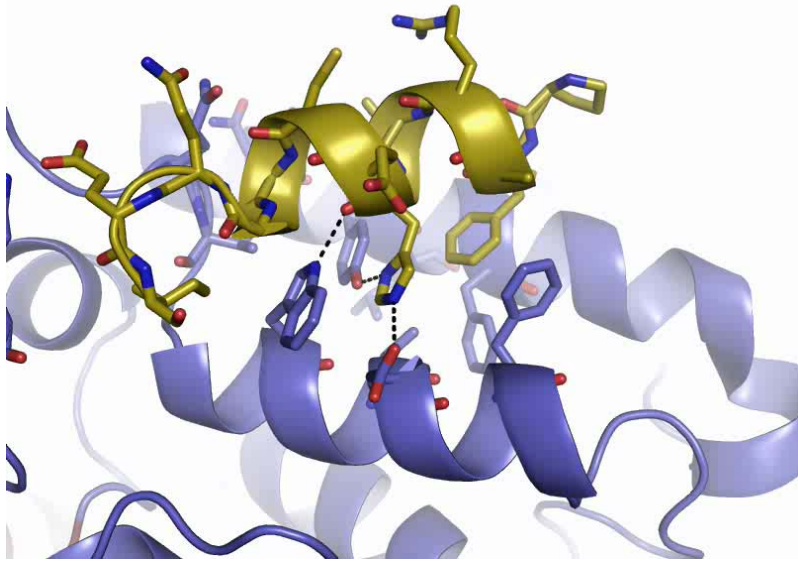
hydrophobic pocket Phe842 and Phe746

Side chain forms hydrogen bond with Asp743  
Main chain carbonyl H- bond with Indole nitrogen of Trp739

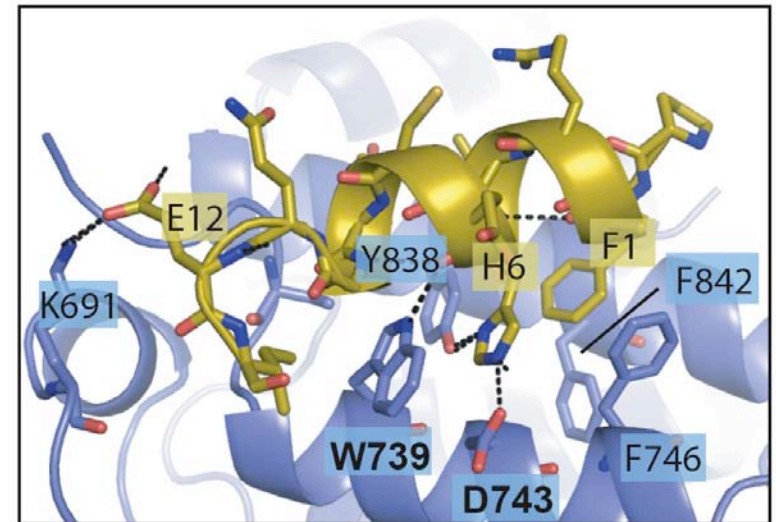
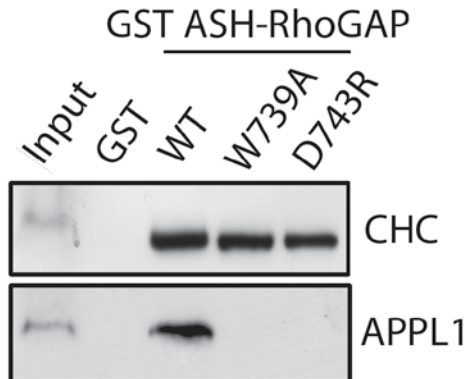
<b>P</b> FAR <b>L</b> HE <b>C</b> Y <b>G</b> Q <b>E</b> I	Ses1
<b>C</b> F <b>S</b> T <b>L</b> H <b>D</b> W <b>Y</b> G <b>Q</b> E <b>I</b>	Ses2
<b>E</b> F <b>A</b> R <b>N</b> H <b>E</b> R <b>F</b> R <b>R</b> E <b>L</b>	Dses
<b>S</b> F <b>Q</b> Q <b>R</b> H <b>E</b> S <b>L</b> R <b>P</b>	APPL1



# Crystallographic Analysis of the F&H/OCRL interaction

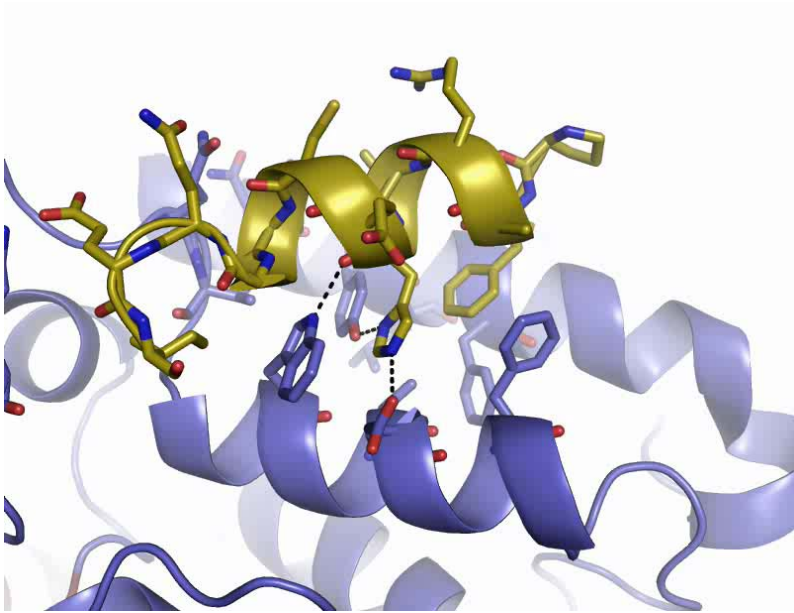


<b>P</b> FAR <b>L</b> H <b>E</b> C <b>Y</b> G <b>Q</b> E <b>I</b>	Ses1
<b>C</b> F <b>S</b> T <b>L</b> H <b>D</b> W <b>Y</b> G <b>Q</b> E <b>I</b>	Ses2
<b>E</b> F <b>A</b> R <b>N</b> H <b>E</b> R <b>F</b> R <b>R</b> E <b>L</b>	Dses
<b>S</b> F <b>Q</b> Q <b>R</b> H <b>E</b> S <b>L</b> R <b>P</b>	APPL1



Binding APPL from rat brain homogenate

# Crystallographic Analysis of the F&H/OCRL interaction



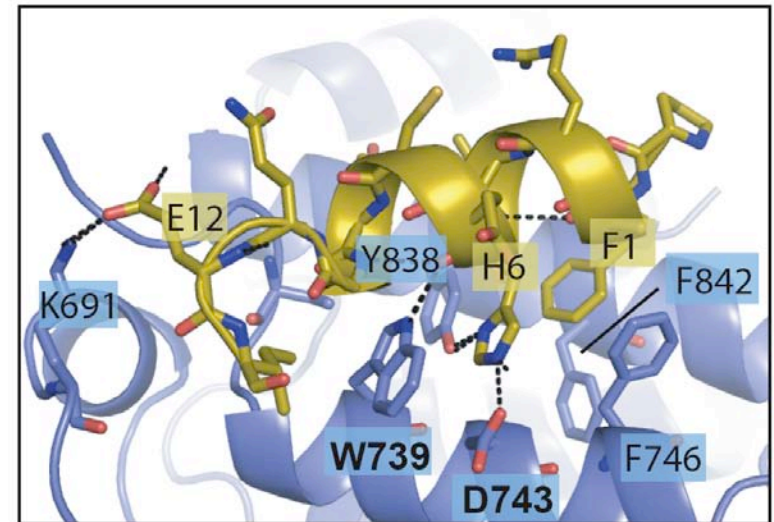
<b>P</b> FARL <b>H</b> ECY <b>G</b> QE <b>I</b>	<b>Ses1</b>
<b>C</b> FSTL <b>H</b> DWY <b>G</b> QE <b>I</b>	<b>Ses2</b>
<b>E</b> FARN <b>H</b> ER <b>F</b> RREL	<b>DSes</b>
<b>S</b> FQQR <b>H</b> ES <b>L</b> RP	<b>APPL1</b>

**Ses1**

**APPL**

Terminal Proline 11  
In the minimal APPL1 F&H peptide

Glu12 H-bond  
Lys691

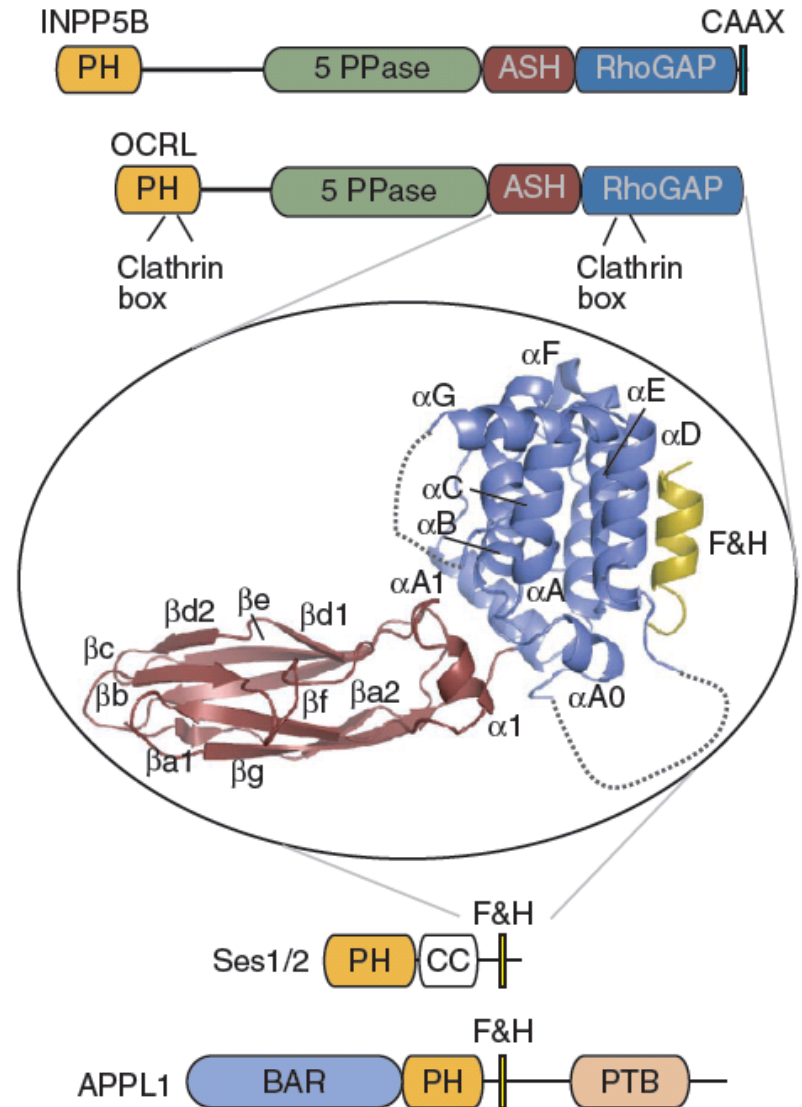




F&H binding site on ASH-RhoGAP domain of OCRL is highly conserved throughout evolution.

This interface is conserved in lower organisms that encode an OCRL and INPP5B homolog but neither APPL1 nor Ses1/2.

What are the interacting partners in these organisms?

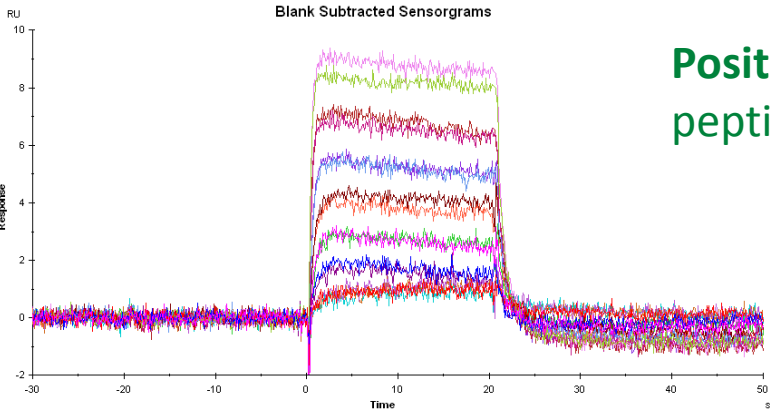


Utilize SPR and ITC to test for binding of F&H peptides to ASH-RhoGAP OCRL and rank their affinities

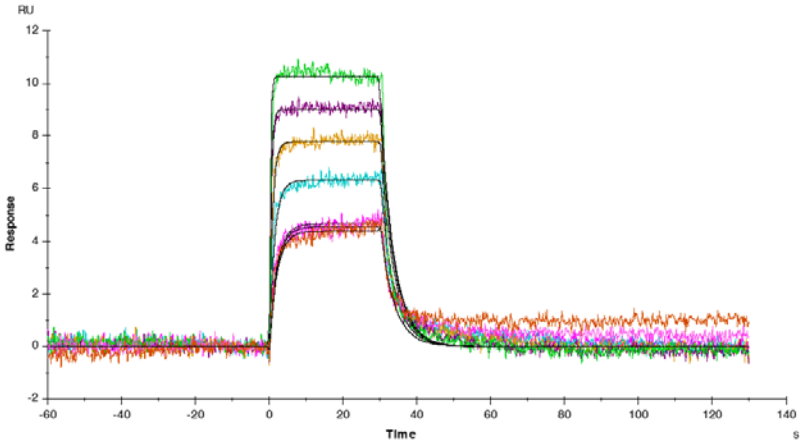
F&H peptides candidates

<b>S</b> FAR <b>L</b> HE <b>C</b> Y <b>G</b> Q <b>E</b> I	<b>Superclamp; positive control</b> (engineered F&H peptide)	
S <b>F</b> Q <b>R</b> H <b>E</b> S <b>L</b> Y <b>R</b> P	APPL1	
P <b>F</b> AR <b>L</b> HE <b>C</b> Y <b>G</b> Q <b>E</b> I	Ses1	Endocytic proteins
C <b>F</b> ST <b>L</b> H <b>D</b> W <b>Y</b> Q <b>E</b> I	Ses2	
<b>K</b> FRR <b>Q</b> H <b>E</b> Q <b>L</b> RA <b>V</b> I	<b>Dynein Heavy Chain</b>	F&H peptide candidates
S <b>F</b> Y <b>V</b> R <b>H</b> S <b>C</b> L <b>R</b> E <b>A</b> L	<b>zFyve26</b> (Spastizin)	Selected through bioinformatics
S <b>F</b> ST <b>V</b> H <b>E</b> K <b>F</b> N <b>K</b> S <b>L</b>	<b>WDR36</b>	
I <b>F</b> GL <b>H</b> H <b>I</b> G <b>M</b> Q <b>M</b> R <b>I</b>	<b>CFTR</b> , cystic fibrosis	
S <b>F</b> ET <b>Q</b> H <b>H</b> H <b>L</b> L <b>H</b> C <b>L</b>	<b>kv4.2</b>	
E <b>F</b> CR <b>N</b> H <b>F</b> L <b>V</b> G <b>L</b> L <b>L</b>	<b>Dock9</b>	
A <b>F</b> IER <b>H</b> R <b>I</b> I <b>E</b> E <b>P</b>	<b>Fly Weeble</b>	

# Utilize SPR and ITC to test for binding of F&H peptides to ASH-RhoGAP OCRL and rank their affinities



**Positive Control:** Superclamp (engineered F&H peptide)

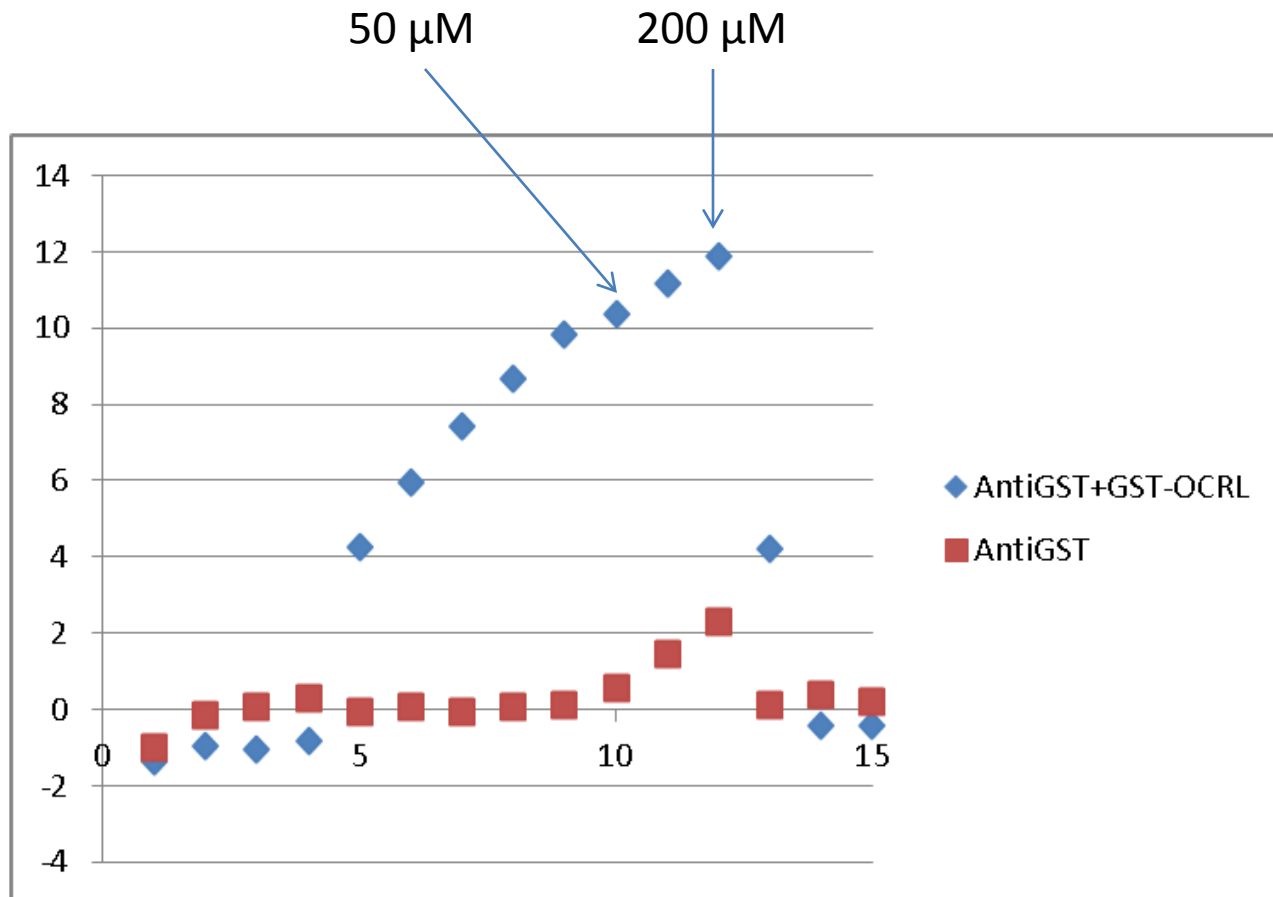


Peptides

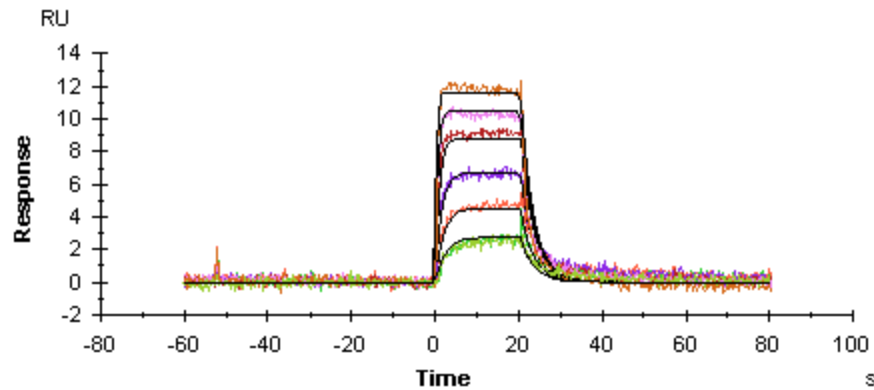


GST-OCRL; captured on anti-GST Ab

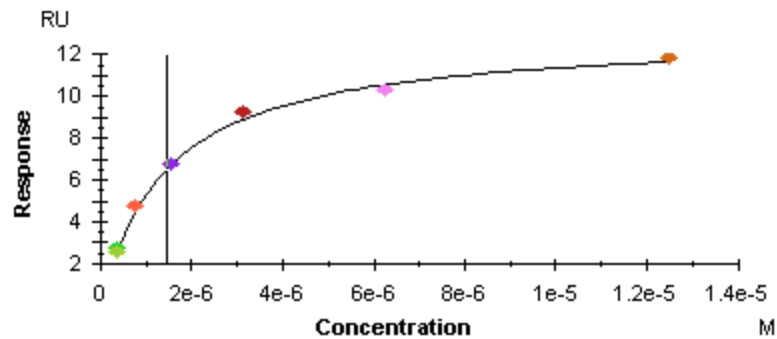
# Non-specific binding to antiGST Ab surface



# Kinetics of binding beyond SPR capabilities; determination of affinity from steady-state amplitudes



Peptides  
→



OCRL; captured on anti-GST Ab

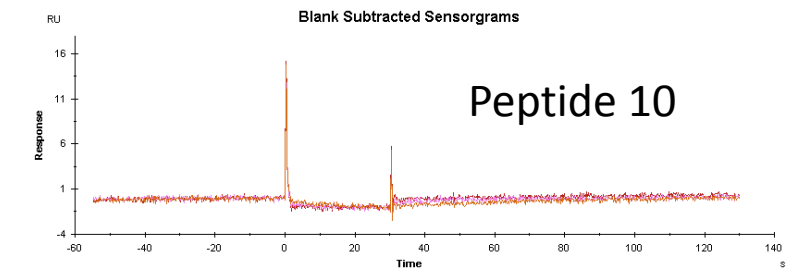
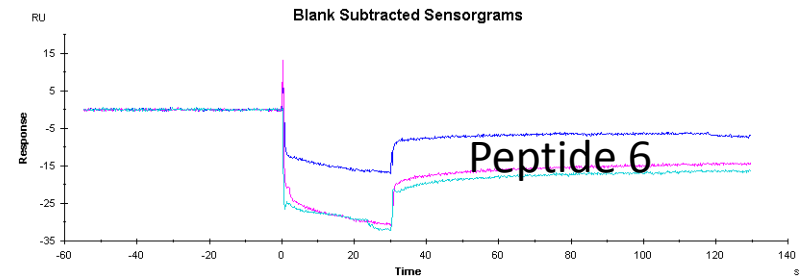
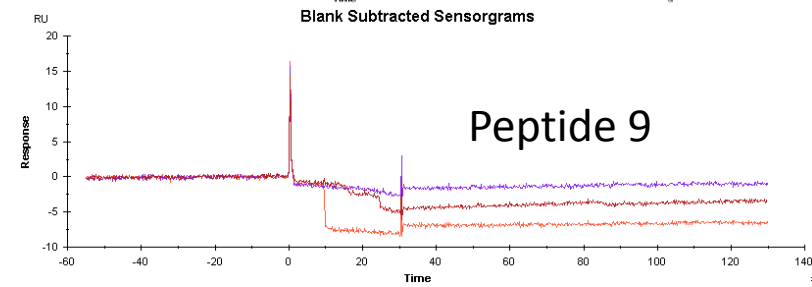
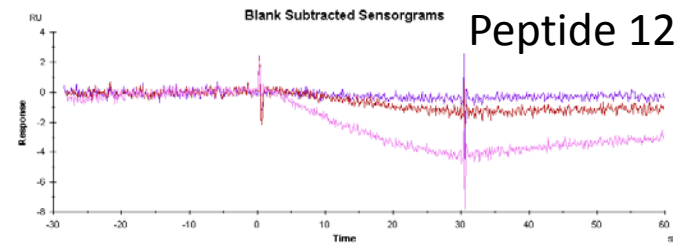
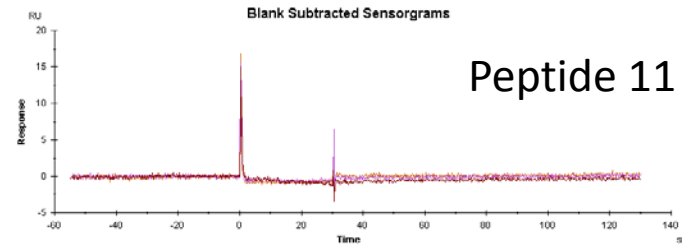
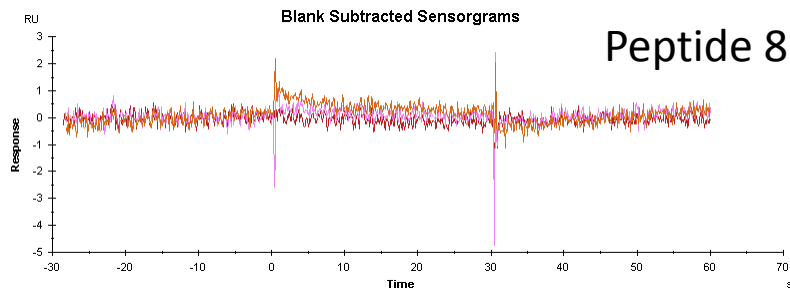
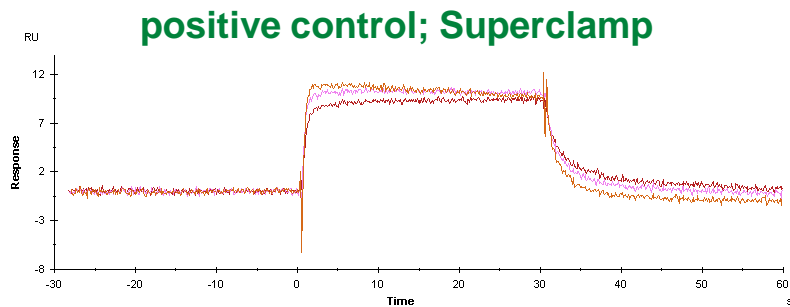
# Testing binding of F&H peptide candidates to ASH-RhoGAP OCRL

GST\_OCRL (WT) captured on anti-GST surface

AntiGST

AntiGST+GST-OCRL

peptides at 50  $\mu$ M, 16,7, and 5.56  $\mu$ M



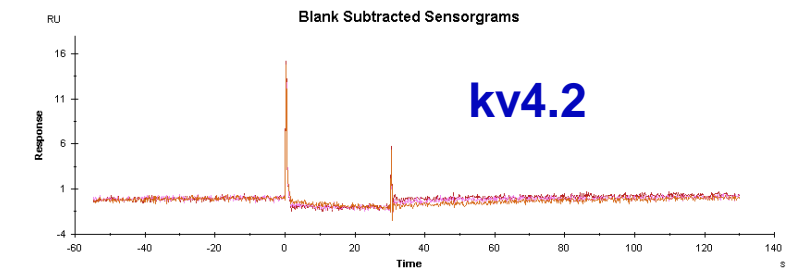
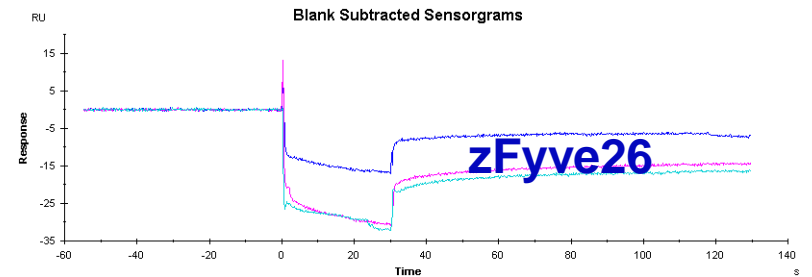
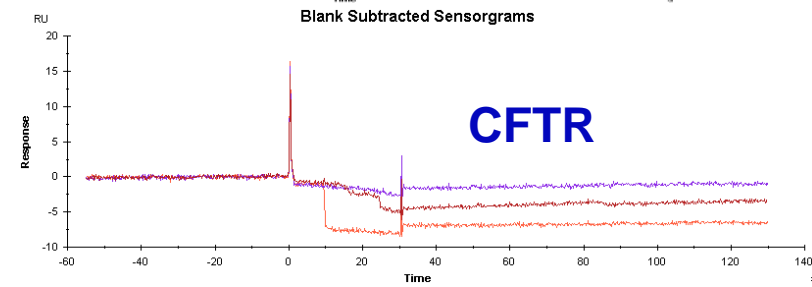
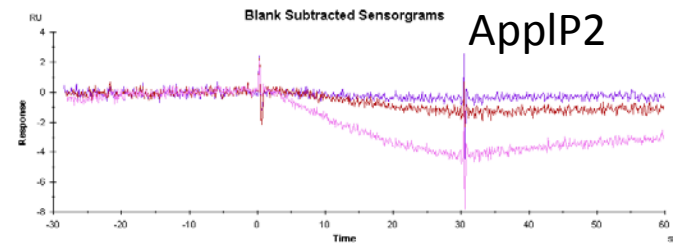
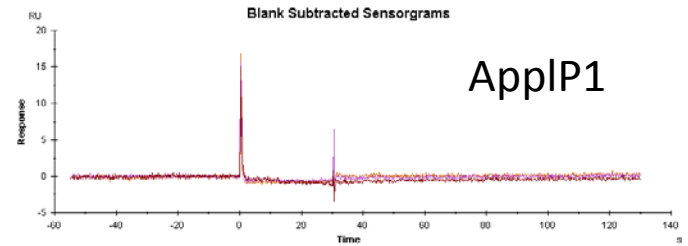
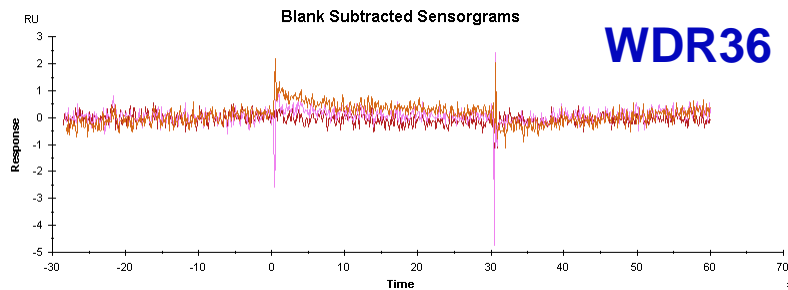
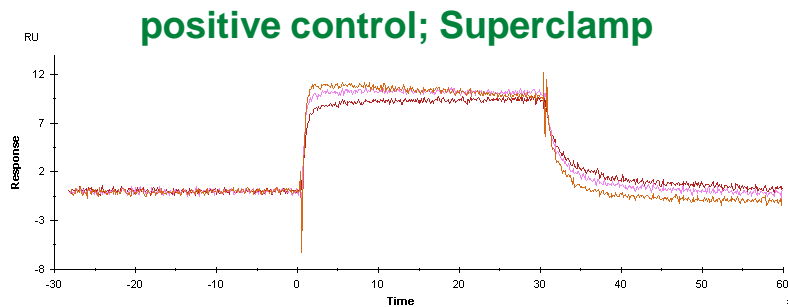
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GST\_OCRL (WT) captured on anti-GST surface

AntiGST

AntiGST+GST-OCRL

peptides at 50  $\mu$ M, 16,7, and 5.56  $\mu$ M



# Testing binding of F&H peptide candidates to ASH-RhoGAP OCRL by SPR

SFARLHECYGQEI Superclamp; positive control  
SFQQRHESLYRP APPL1  
PFARLHECYGQEI Ses1  
CFSTLHDWYGQEI Ses2

Candidates for SPR

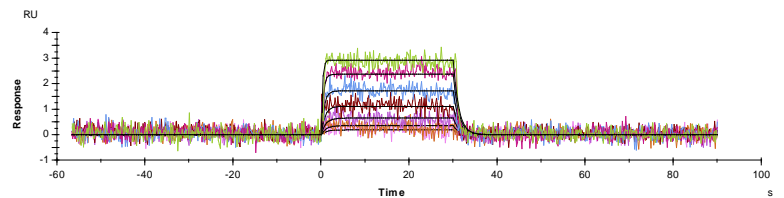
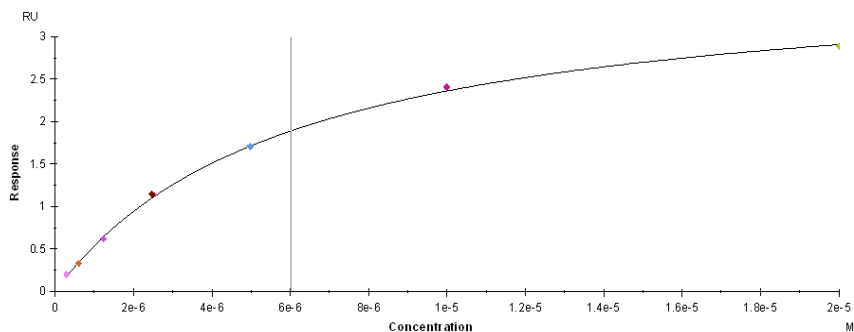
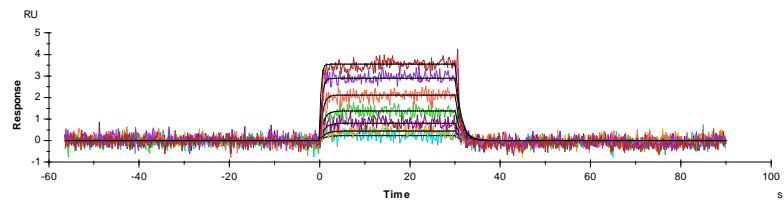
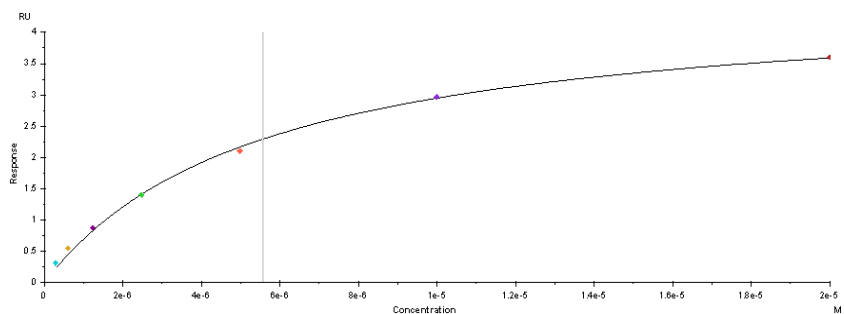
KFRRQHEQLRAVI **Dynein Heavy Chain**  
SFYVRHSCREAL **zFyve26** (Spastizin)  
SFSTVHEKFNKSL **WDR36**  
IFGLHHIGMQMRI **CFTR**, cystic fibrosis  
SFETQHHLLHCL **kv4.2**  
EFCRNHFLVGLLL **Dock9**  
AFIERHRIIEEP **Fly Weeble**

anti-GST binder  
anti-GST binder  
no binding in SPR  
no binding in SPR (?)  
no binding in SPR  
insoluble in SPR buffer  
anti-GST binder



# Affinity for binding of F&H peptides to ASH-RhoGAP OCRL

## Peptide #1

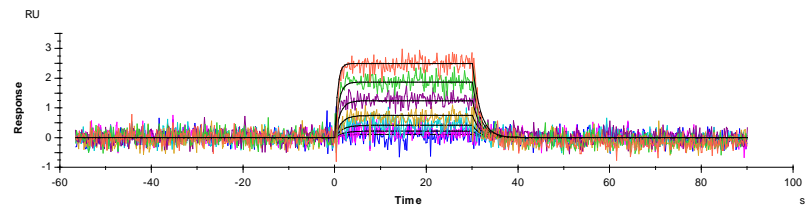
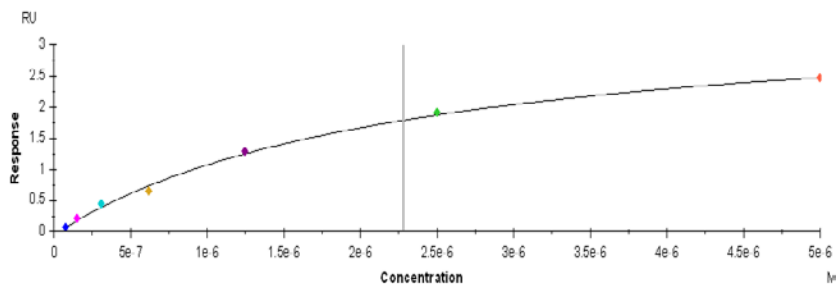
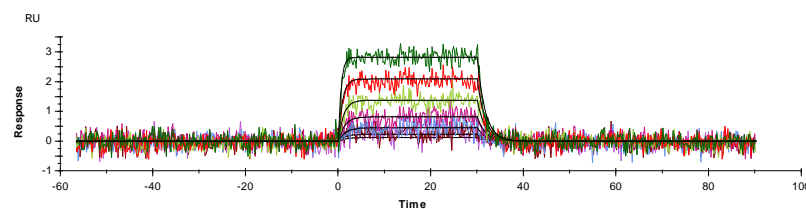
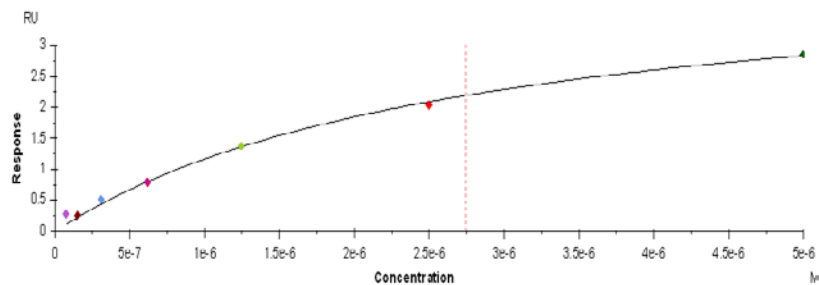


Ses #1

Kd  $5.8 \pm 0.3$  uM

# Affinity for binding of F&H peptides to ASH-RhoGAP OCRL

Peptide #2

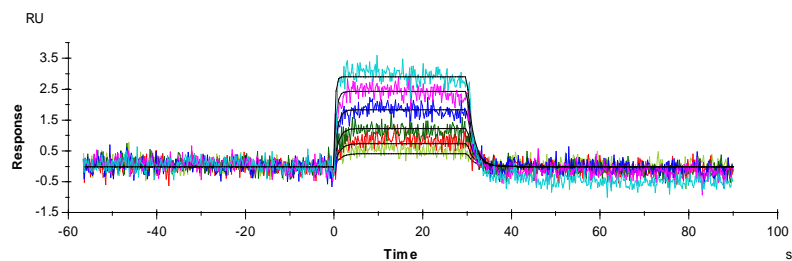
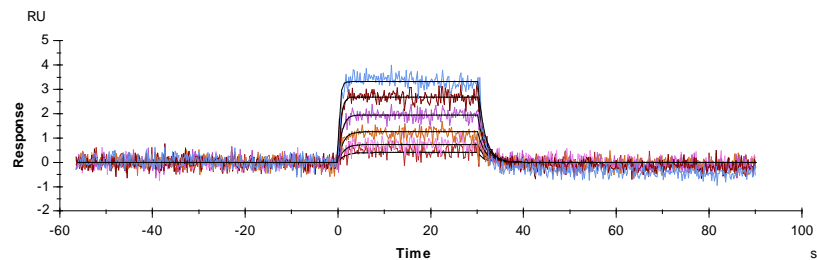
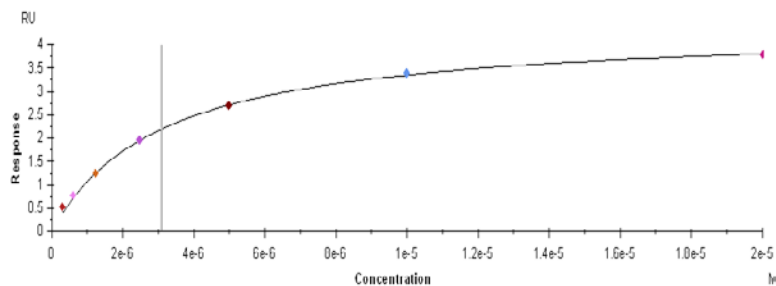
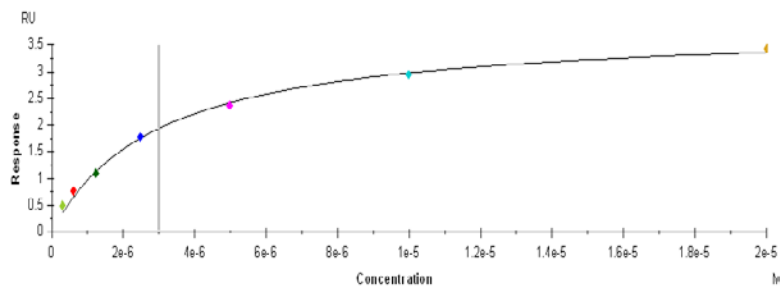


Ses2

Kd  $2.5 \pm 0.4 \mu\text{M}$

# Affinity for binding of F&H peptides to ASH-RhoGAP OCRL

## Peptide #5



Superclamp

Kd  $3.0 \pm 0.2$   $\mu$ M

# Testing binding of F&H peptide candidates to ASH-RhoGAP OCRL by SPR

SFARLHECYGQEI	Superclamp; positive control
SFQQRHESLYRP	APPL1
PFARLHECYGQEI	Ses1
CFSTLHDWYGQEI	Ses2

Kd measured by SPR

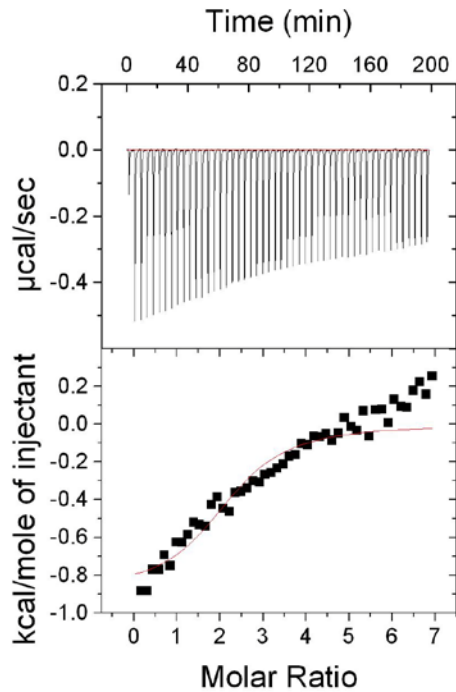
## Candidates for ITC follow up

KFRRQHEQLRAVI	<b>Dynein Heavy Chain</b>
SFYVRHNSCLREAL	<b>zFyve26</b> (Spastizin)
SFSTVHEKFNKSL	<b>WDR36</b>
IFGLHHIGMQMRI	<b>CFTR</b> , cystic fibrosis
SFETQHHLLHCL	<b>kv4.2</b>
EFCRNHFLVGLLL	<b>Dock9</b>
AFIERHRIIEEP	<b>Fly Weeble</b>

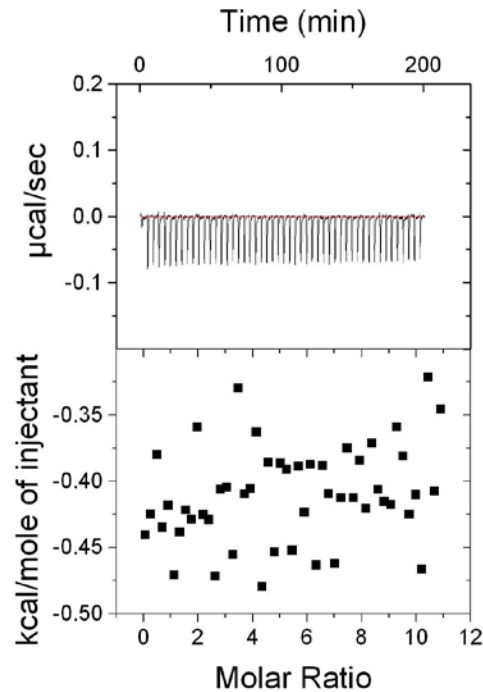
NON-SPECIFIC  
anti-GST binder  
no binding in SPR  
no binding in SPR (?)  
no binding in SPR  
insoluble in SPR buffer  
anti-GST binder

# Testing binding of F&H peptide candidates to ASH-RhoGAP OCRL by ITC

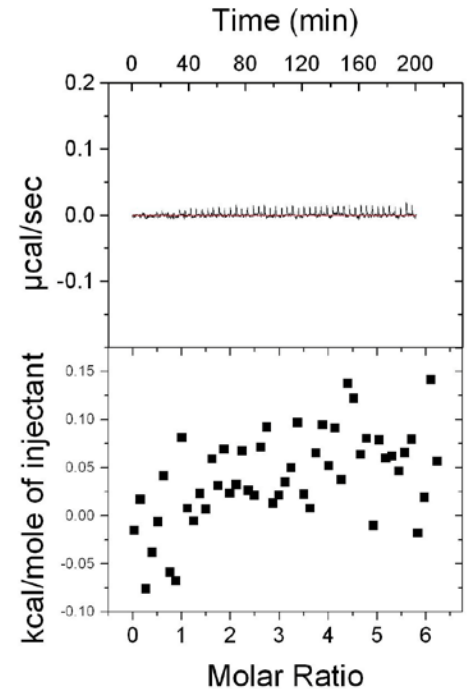
## Zfyve-26



## Fly Weeble

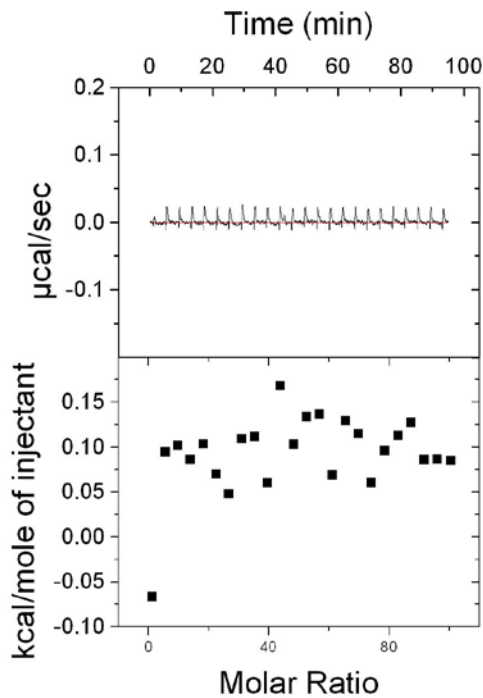


## Kv 4.2

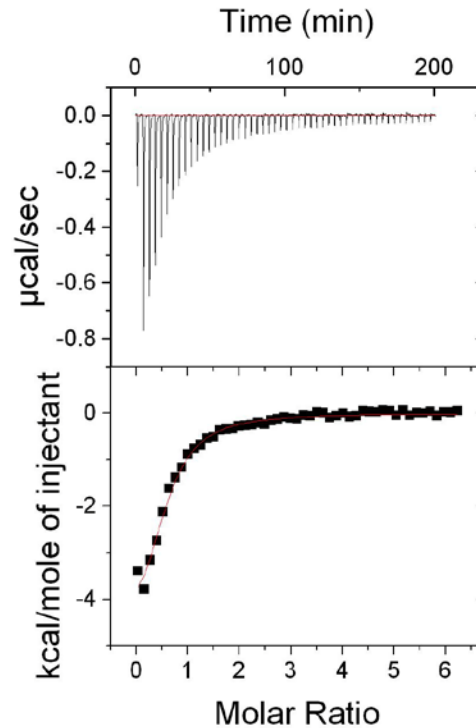


solubility artifact?

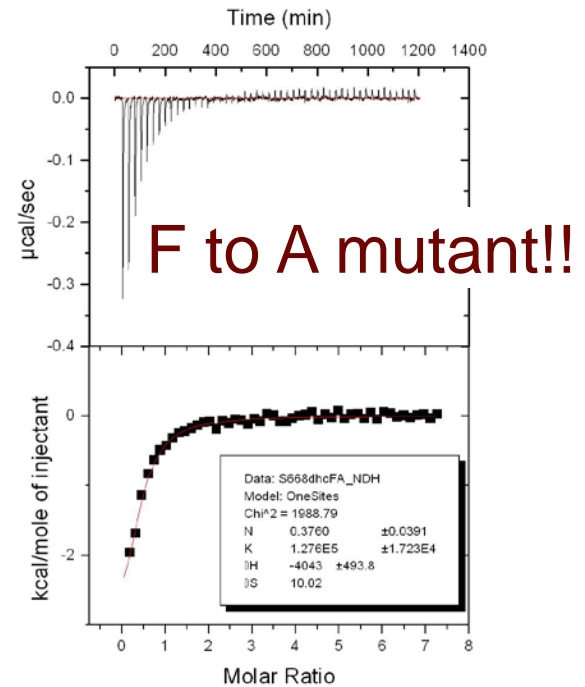
# Identification of Dynein heavy chain as a potential interactor



**Dynein Heavy Chain  
Into Buffer**

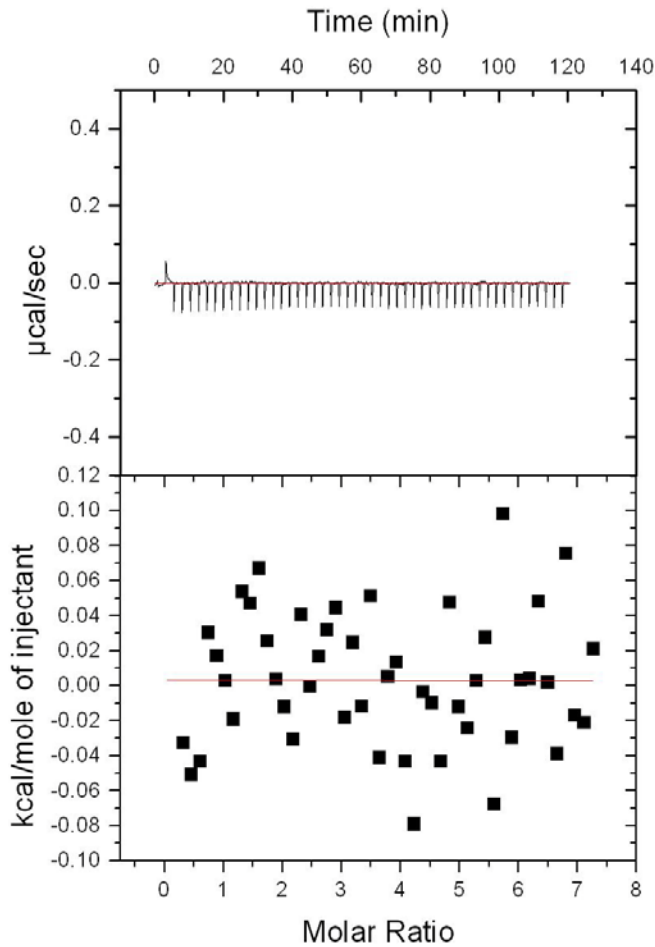


**Dynein Heavy Chain  
Into OCRL**

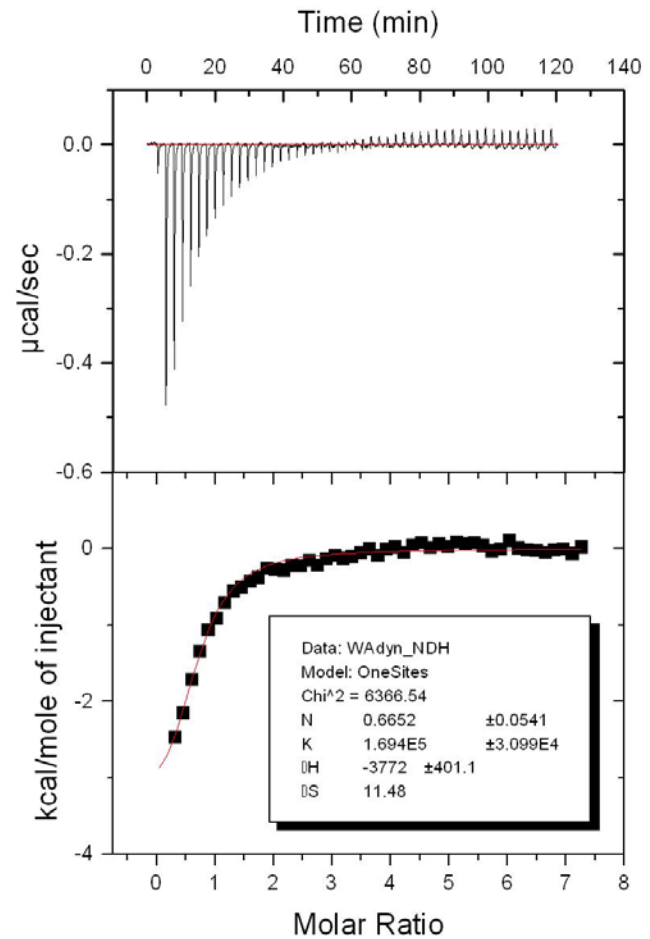


**Dynein F to A Mutant  
Into OCRL**

# Control ITC: OCRL W739A mutant

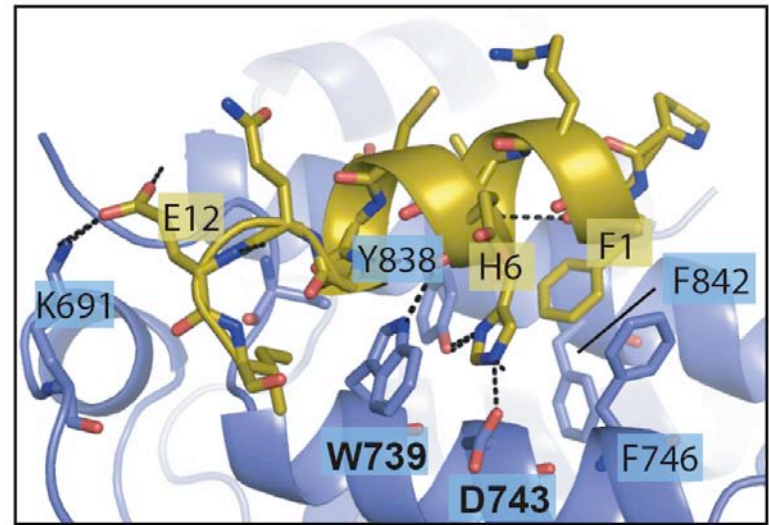
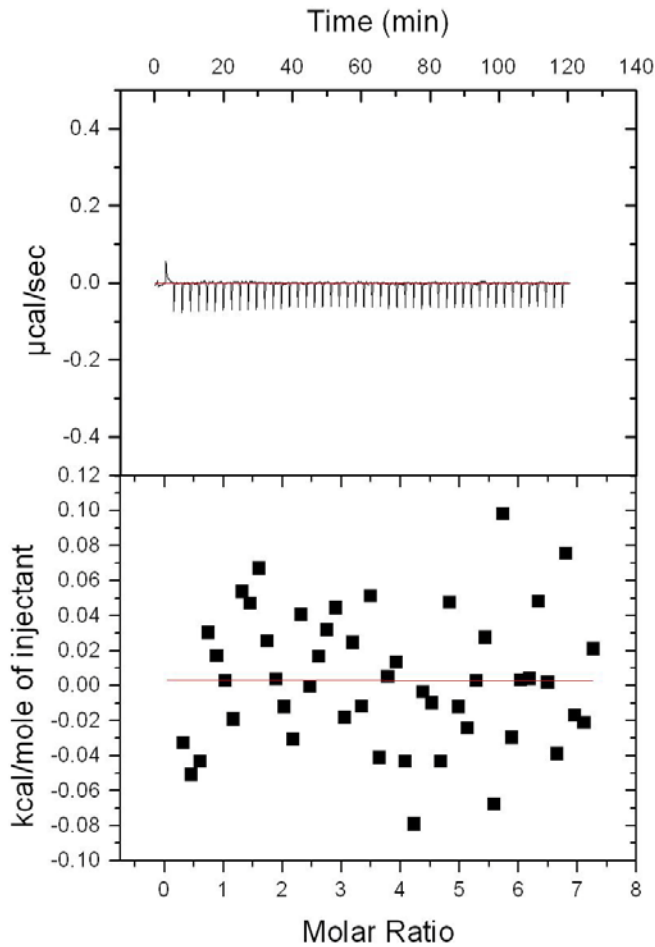


**Ses2 peptide**  
**W739A ASH-RhoGAP**



**Dynein peptide**  
**W739A ASH-RhoGAP**

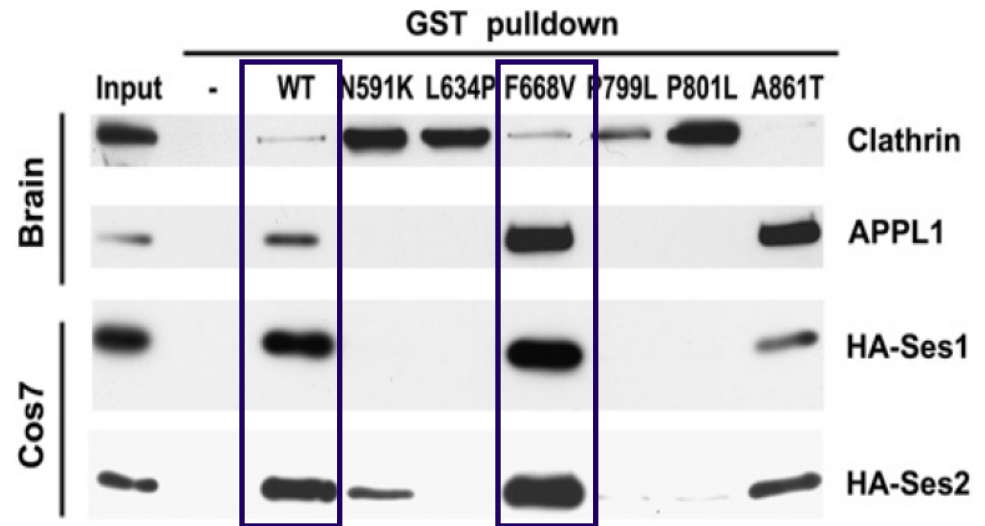
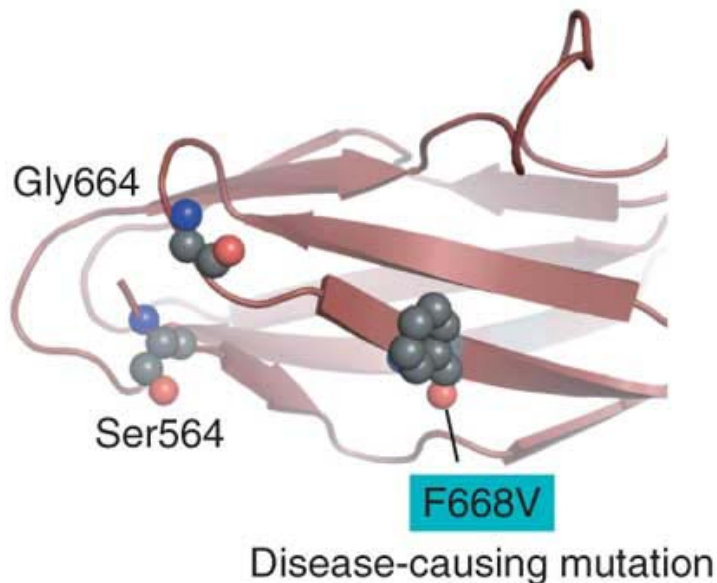
# Control ITC: OCRL W739A mutant



**Ses2 peptide**  
**W739A ASH-RhoGAP**

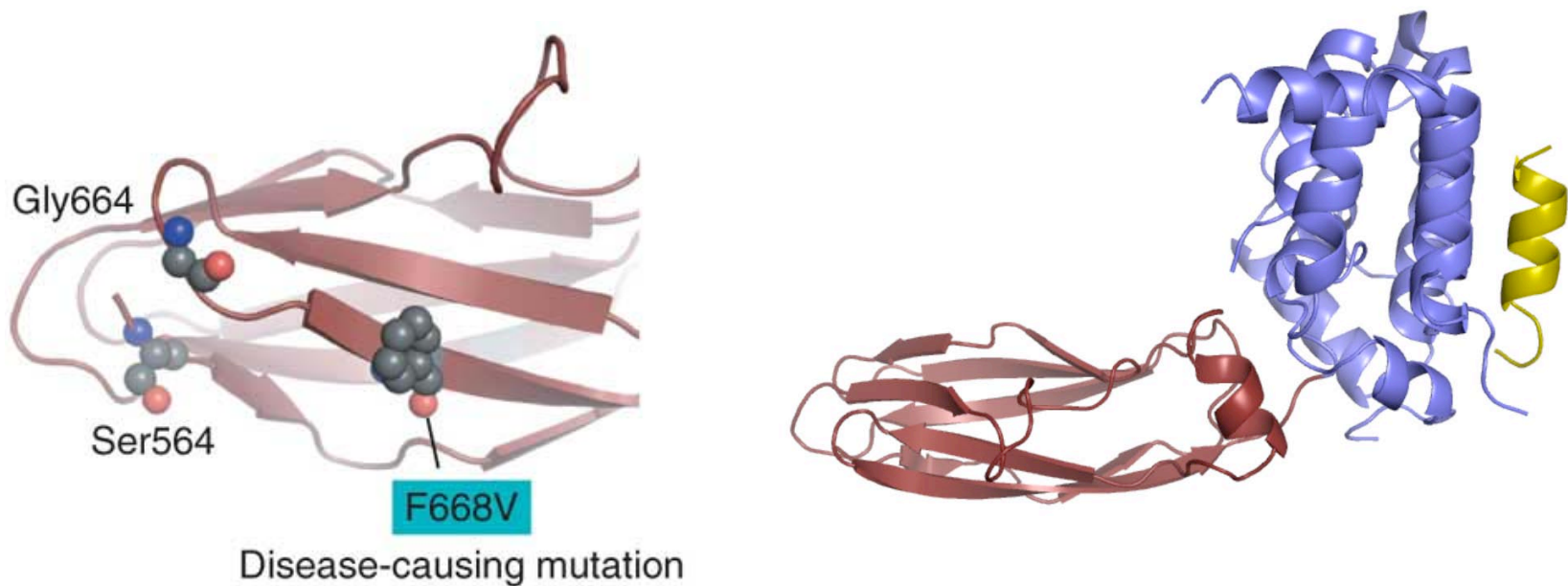


Utilize SPR and ITC to determine affinity for binding of F&H peptides to ASH-RhoGAP OCRL (wt and disease-causing mutants)



Determine whether the F668V mutant binds more tightly to F&H peptides

Utilize SPR and ITC to determine affinity for binding of F&H peptides to ASH-RhoGAP OCRL (wt and disease-causing mutants)

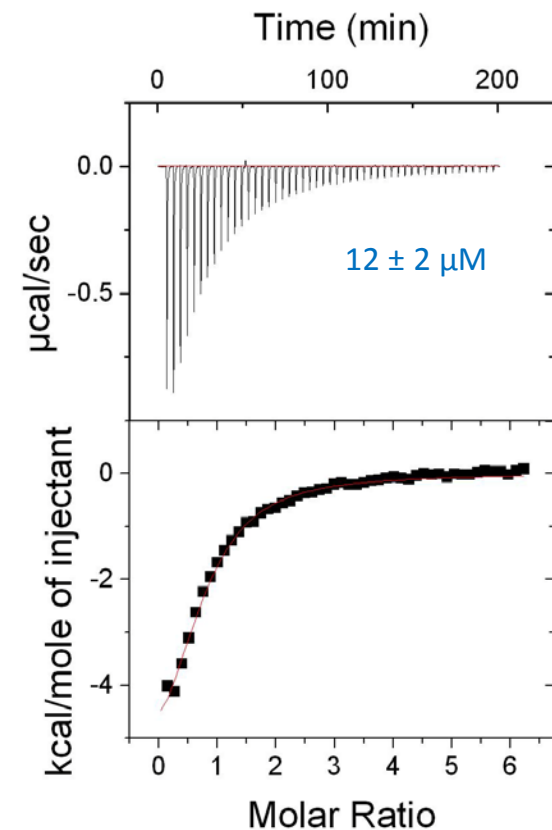


Determine whether the F668V mutant binds more tightly to F&H peptides

# Utilize SPR and ITC to determine affinity for binding of F&H peptides to ASH-RhoGAP OCRL (wt and disease-causing mutants)

Proteins	Kd ( $\mu\text{M}$ ) SPR/ <i>ITC</i>	
	Peptides	
	APPL	Superclamp
Wt OCRL	$12 \pm 2$	$3.0 \pm 0.2$
F668V	$12 \pm 2$	$2 \pm 1$

F668V mutant has identical binding affinities to the F&H proteins as the wt protein



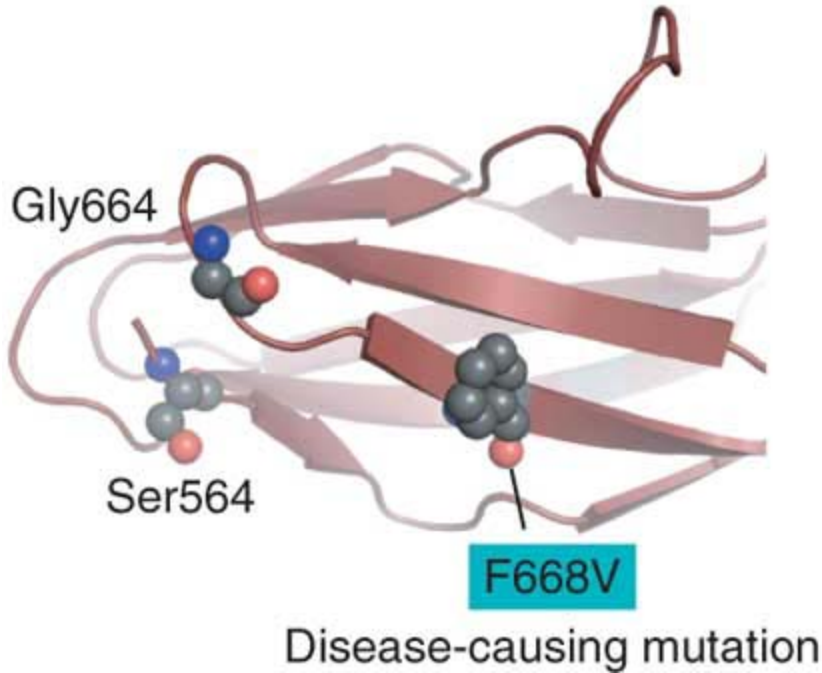
APPL binding to F668V OCRL

# Summary

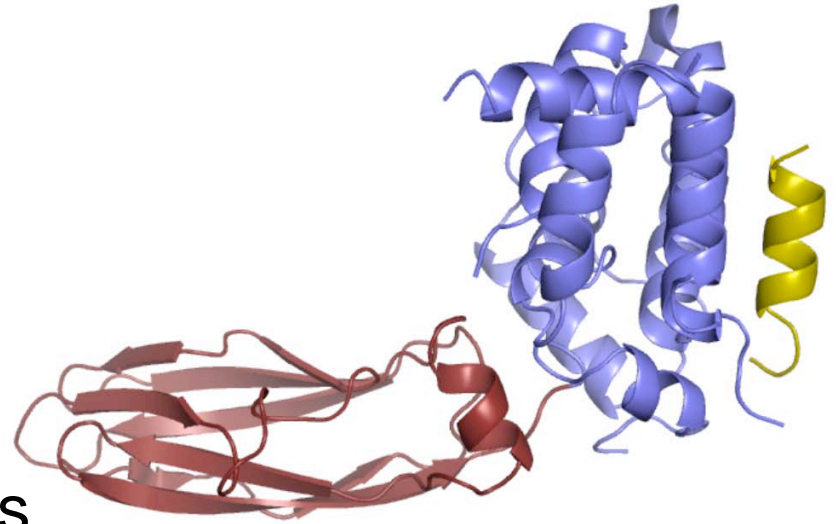
Kd ( $\mu$ M) SPR/ <i>ITC</i>								
Proteins	Peptides							
	APPL	Ses1	Ses2	SesFA	Superclamp	APPLP1	APPLP2	Dynein Heavy Chain
Wt OCRL	43.1 $\pm$ 0.4 <i>12 <math>\pm</math> 2</i>	5.8 $\pm$ 0.3 <i>0.70 <math>\pm</math> 0.08</i>	2.5 $\pm$ 0.4	ND <i>ND</i>	3.0 $\pm$ 0.2	ND <i>ND</i>	ND <i>ND</i>	<i>92 <math>\pm</math> 4</i>
W739A (engineered)	ND	ND	ND	ND	ND	ND	ND	<i>8 <math>\pm</math> 1</i>
F668V (disease causing)	<i>12 <math>\pm</math> 2</i>				2 $\pm$ 1			

- **F668V** and wt **AshRhoGAP** bind to F&H peptides with similar affinities
- No binding of F&H peptides was observed for **W739A** mutant
- Bioinformatics approach did not yield new F&H candidates that interact with F&H motif binding surface of OCRL

## Future Direction



Rabs



Characterize the **Rab binding surface** of a full length OCRL

Michelle Pirruccello and Laura Swan

Pietro DeCamilli

Department of Cell Biology Yale University and HHMI

Ken Williams

Yale/NIDA Neuroproteomics Center

W.M. Keck Biotechnology Resource Laboratory at Yale University School of Medicine

NIH (Biacore T100 purchase, Yale/NIDA Center support)

HHMI (VP-ITC purchase)

<http://info.med.yale.edu/wmkeck/biophysics>

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# Patient mutations affecting F&H binding in the ASH-RhoGAP domain of OCRL destabilize the protein

